



Review

Mast cells, mediators, and symptomatic activation

Amolak S Bansal, Alex Nicholas, Nazira Sumar and Veronica Varney*

Dept of Allergy and Immunology, St Helier Hospital, Wrythe Lane, Carshalton, Surrey, UK

* **Correspondence:** Email: veronica.varney@btinternet.com.

Abstract: Mast cells (MC) are central effectors of allergic disease and distinct subsets with varying amounts of tryptase, chymase, and carboxypeptidase A3, and cathepsin G is distributed throughout the body. Their involvement in a diverse range of non-allergic illnesses mediated by a complex range of preformed and newly synthesized mediators is now increasingly recognized. The latter especially include conditions under the umbrella term of mast cell activation syndrome. In allergic disease, much has been written about the mechanisms by which the early and rapidly acting mediators produce both localized and systemic allergic symptoms. The role of chymase is presently underappreciated but there is increased awareness that MCs contain significant amounts of preformed TNF alpha and synthesize and releases a wide range of inflammatory cytokines such as interleukins (IL) 1 β , IL6, IL31, and IL33. These can aggravate itching and perpetuate inflammation and likely contribute to the late constitutional symptoms seen in allergic reactions. Importantly, their involvement helps to clarify the role of MCs in stress and non-parasitic infections. Presently, unexplained is the increasing incidence of significant acute allergic reactions within a relatively short time frame. In this context, there is increasing interest in the environmental, menstrual, endocrine, circadian, and psychological factors that influence MC activation as well as the endocrine pathways involving the renin angiotensin system that oppose hypotension. In non-allergic diseases with normal numbers of MCs, reduced thresholds for activation may be produced by various combinations of life and dietary factors. Diagnosing these conditions is difficult but may be helped by urinary analysis of prostaglandin metabolites. The investigation and management of mastocytosis with and without mutations of c-kit is also relevant to allergic disease and the new medications used may also be helpful in idiopathic anaphylaxis. This knowledge may open a new chapter in human diseases and mast cell regulation.

Keywords: mast cells; anaphylaxis; renin & angiotensin system; genes of renin angiotensin system; endothelial nitric oxide; c-KIT; mast cell disorders; mastocytosis

Abbreviations: ACE: angiotensin converting enzyme; RAS: renin angiotensin system; AII: angiotensin-2; AI: angiotensin-1; PAF: platelet activating factor; eNO: endothelial nitric oxide; ATR-1: angiotensin-2 receptor; BK: bradykinin; BP: blood pressure; C5a: complement protein C5a; C3a: complement protein C3a; mRNA: messenger RNA; IgE: Immunoglobulin E

1. Introduction

Mast cells (MC) are widely distributed and have central roles in allergic disease as well as an increasing range of illnesses caused by either raised numbers of these cells and/or lower thresholds for activation and degranulation. They contain numerous mediators, some of which are preformed and some newly synthesized on activation. While many of the preformed mediators are critical for the acute cardiorespiratory, cutaneous, and gastrointestinal symptoms some of the cytokines also contribute to the late phase reaction and many of the constitutional symptoms. The role of some of the mediators such as chymase is also being redefined along with a range of cofactors that can facilitate MC activation.

Anaphylaxis can be defined as an acute systemic hypersensitivity reaction resulting from the release of mast cell histamine with other mediators that have widespread effects upon 2 or more organs (skin, lungs, and gastrointestinal and cardiovascular systems) [1]. While many cases of anaphylaxis are caused by re-exposure to allergen and cross linkage of adjacent specific IgE on the MC surface, some arise from specific IgG and others from direct activation of normal or raised numbers of MCs. Numerous factors are now considered to modulate the severity of an allergic reaction. These include genetic, endocrine and nutritional variables, enzymes and proteins involved blood pressure regulation and even stress and concomitant drug use.

In the U.K, 2% of the adult population carries adrenaline due to prior anaphylactic reactions [2]. Anaphylaxis is increasing on a global scale without explanation. Indeed a 9-year Australian study showing a 150% increase in anaphylaxis admissions and a 300% increase in fatalities, especially in children [3]. Demographically, there appears to be a very high incidence of anaphylaxis in young children < 5 years from food allergies and in pre-menopausal women. A 15-year study in the UK confirmed this trend, and showed a 7-fold increase in hospital admission especially for young children but without a clear explanation for the increase [4–7]. Regardless, subjects usually repeat their prior allergic reactions on further exposure unless the allergen dose is small [1,4,8].

2. The central role of Mast cells in allergy anaphylaxis, and mast cell disorders

2.1. Clinical and genetic aspects of allergic disease and anaphylaxis

MC activation can result in clinical symptoms ranging from mild cutaneous effects (Grade I) to worsening bronchospasm, angioedema and cardiac arrest from profound hypotension and circulatory collapse (Grade 4) as per Terr's original Classification 1985 [8]. Thankfully, severe anaphylaxis is a relatively rare but the manifestations can go unrecognized in some cases. Autopsy studies have shown 80% of deaths to have upper airway oedema with secondary pulmonary hyperinflation and circulatory collapse as their only features [9,10]. Unsurprisingly, prior cardio-respiratory disease and poorly controlled asthma increases the risk of death in adults. The presence of asthma in small children with allergic reactions is a significant risk factor for a severe outcome, as small children have a lower

lung capacity with histamine-induced bronchoconstriction producing greater respiratory difficulty than seen for adults [5,6,11].

Presently, there are few links between anaphylaxis and specific “allergy genes” [2]. Additionally, the host cofactors, which influence the severity of reactions that remain ill-defined [11–13]. Surprisingly, some subjects experiencing near-fatal anaphylaxis have low levels of circulating allergen-specific IgE, while others with higher levels have no symptoms upon exposure [14]. Unfortunately, many causes of anaphylaxis remain unidentified although foods are a leading cause in children, whereas venom or drugs account for the majority of adult cases [15,16]. In the hospital setting, the risk of cardiac arrest and death was greater in obese patients, those with coronary artery disease or in those taking beta-blockers or ACE-inhibitor drugs. The commonest triggers were antibiotics, neuromuscular blocking drugs, the anti-septic chlorhexidine, patent blue dye, blood products and gelatin [17]. Serum tryptase analysis can help in supporting a diagnosis of anaphylaxis but confirm only mast cell activation and the clinical history is critical. This is especially so in the case of patients manifesting the symptoms of allergic mediator release but without demonstrable specific IgE.

2.2. Mast cell subtypes and distribution

It is now abundantly clear that mast cells (MC) are distributed throughout the body including the myocardium and in the retina where they have been noted in the bursa premacularis and especially in certain ocular conditions [18]. Within the central nervous system, they are located in several areas and can influence stress mediated allergic type reactions [19] and are also involved in regulating sleep [20]. They are central players in the initiation and progression of an allergic immune response and critical in the majority of anaphylactic reactions [21]. They are also critically important in all types of mast cell activation syndrome [22–24] and especially in mastocytosis [22,24] where there is an increase in the numbers of irritable MCs. Developmentally, MC are derived from CD34+/CD117+/CD13+ multipotent, hematopoietic progenitors stimulated by the action of stem cell factor on the c-kit transmembrane tyrosine kinase [25,26]. They are broadly divided into mucosal and tissue MCs based on morphological differences and variations in tryptase, chymase, cathepsin G, and carboxypeptidase A3 [27,28]. As such connective tissue MCs are located predominantly in the skin and submucosa of the gastrointestinal tract mucosa and produce all four proteins while mucosal MCs are found in the alveoli and mucosa and produce only tryptase [26,29–31]. There are also differences in the precise amounts of allergic mediators and cytokines held and released by these subsets, which likely reflect subtle differences in their primary function [32].

2.3. Mast cell release of preformed mediators

It is now clear that complexes of IgE and FcεR remain on the surface of the MC for a long time and are the main mechanism of sensitization to the allergen. MCs are activated by allergen cross-linking of pre-existing allergen specific IgE bound to FcεRI, by non-IgE mediated mechanisms [33] and by IgG anti-IgE antibodies evident in some patients with atopic dermatitis [34]. These results in rapid release several preformed mediators held in numerous granules as part of the initial allergic response [35] (Table 1). Important amongst these are histamine, tryptase, chymase, heparin, tumor necrosis factor alpha (TNFα), platelet activating factor (PAF), and others detailed in Table 2. The action of histamine is well known and includes smooth muscle contraction leading to vasodilation and

bronchoconstriction and disruption of the micro-circulation leading to angioedema and hypotension and central nervous system symptoms [36–38]. Tryptase promotes inflammation and stimulates fibrin formation as well as the conversion of IL33 into a more active form [39]. Importantly, MCs are the only cells that hold preformed TNF α [40], which is rapidly secreted and can contribute to attraction and activation of several inflammatory cells [41], including T cells [42–44]. It is possible that TNF α contributes to the exhaustion, drowsiness and flu-like sensation that follows acute severe allergic reactions although histamine stimulation through the H3 receptor is likely also involved [45].

Table 1. Mast cell mediators.

Pre-stored Mediators and enzymes	Actions
Histamine	Vasodilation, hypotension, itch
5-Hydroxytryptamine	Vasoconstriction
Tryptase	Inflammation, tissue damage, pain
Chymase	Neutralises bradykinin, angiotensin II synthesis
Heparin	Angiogenesis, anticoagulant, stabilizes nerve growth factor
Kinogenases	Synthesis of vasodilatory kinins, pain
Carboypeptidase A	Peptide processing
Metalloproteinases	Tissue damage
Phospholipases	Arachidonic acid generation
Interleukin-8	Neutrophil attraction
Platelet activating Factor	Causes hypotension in anaphylaxis and further mast cell degranulation
Peptides/proteins	
Corticotropin-releasing hormone	Inflammation, vasodilatation
Endorphins	Analgesia, vasodilatation
Endothelin	Sepsis
Bradykinin	Inflammation, pain,
Neurokinin	Inflammation, pain
Renin	vasoconstriction
Angiotensin-I	Vasoconstriction
Somatostatin	Anti-inflammatory
Substance P	Inflammation, pain
Vasoactive Intestinal Peptide	Vasodilatation
Vascular endothelial growth factor	Neovascularization, Vasodilatation

Table 2. Classification of mast cell disorders.

Cutaneous Mastocytosis	1a. Diffuse cutaneous mastocytosis
	1b. Maculopapular mastocytosis (Urticaria Pigmentosa)
	1c. Cutaneous mastocytoma
Systemic Mastocytosis	2a. Indolent systemic mastocytosis
	2b. Smoldering systemic mastocytosis
	2c. Advanced Systemic Mastocytosis
	2d. Mastocytosis associated hematological neoplasia or Mast cell leukemia
	2e. Non-clonal disorders associated with mast cell sarcoma
Mast Cell Activation Syndrome	3a. Primary Mast cell Activating Syndrome/Monoclonal Mast Cell Activating Syndrome/Clonal Mast Cell* Activating Syndrome
	3b. Secondary Mast Cell Activating Syndrome (IgE-mediated allergy)
	3c. Idiopathic Mast Cell Activation Syndrome (no obvious etiology)
	3d. Combined primary disorder with allergy-triggered mast cell activation
Idiopathic Anaphylaxis	4a. No obvious allergens
	4b. Idiopathic anaphylaxis associated with bone marrow mastocytosis + Clonality*
	4c. Idiopathic anaphylaxis associated with Clonal mast cells* or hereditary alpha-tryptasemia
Associations of hymenoptera venom and clonal mast cell disease	5a Mastocytosis and clonal mast cells*
	5b Mast cell activation syndrome ± clonal mast cells*
	5c Bone marrow mastocytosis
	5d Heredity alpha tryptasaemia ± mastocytosis or clonal cell syndrome*
Combined Disorders	Various combination of above conditions
	*clonal mast cells associated with D816V mutation aberrant mast cells associated with CD2 or CD25 surface marker

The role of chymase in anaphylaxis is presently underestimated. It is a serine protease present in the mast cells of the heart, blood vessels, skin, and gut. In the skin, 98% of mast cells produce chymase compared with only 7% of pulmonary and bronchial mast cells. Chymase is stored in secretory granules bound to its inhibitor heparin and was believed to be rapidly degraded by proteases following its release. However, it now appears that it is rapidly captured by circulating alpha-2 Macroglobulin and sequestered within a cage-like structure that precludes its detection in standard assays, limits access to inhibitors, and allows access to the systemic circulation [46]. The Chymase/alpha-2 Macroglobulin complex remains accessible to small substrates like Angiotensin-1 where Chymase plays a role in systemic BP control that could also protect against cardiovascular collapse following massive mast cell mediator release. Newer assays can detect captured Chymase and suggests a steady leak of chymase from mast cells as responsible for the background production of Angiotensin-II. This in turn can stimulate vasoconstriction of the blood vessel smooth muscle cells and is thought to account for up to 80% of the circulating angiotensin-II levels and blood pressure control [47–50]. Importantly, chymase can hydrolytically inactivate peptides such as Bradykinin, kallikrein and substance P.

Mast cell heparin released with chymase activates the ‘contact system’ with auto-activation of clotting factor XII leading to activation of Factor X-mediated Bradykinin formation. Unsurprisingly, heparin is linked to circulating Bradykinin levels and the severity of anaphylaxis [51–55]. In mice

deficient in factor X and bradykinin, systemic anaphylaxis does not produce hypotension or death, indicating an important role for these mediators in severe reactions (Figure 1).

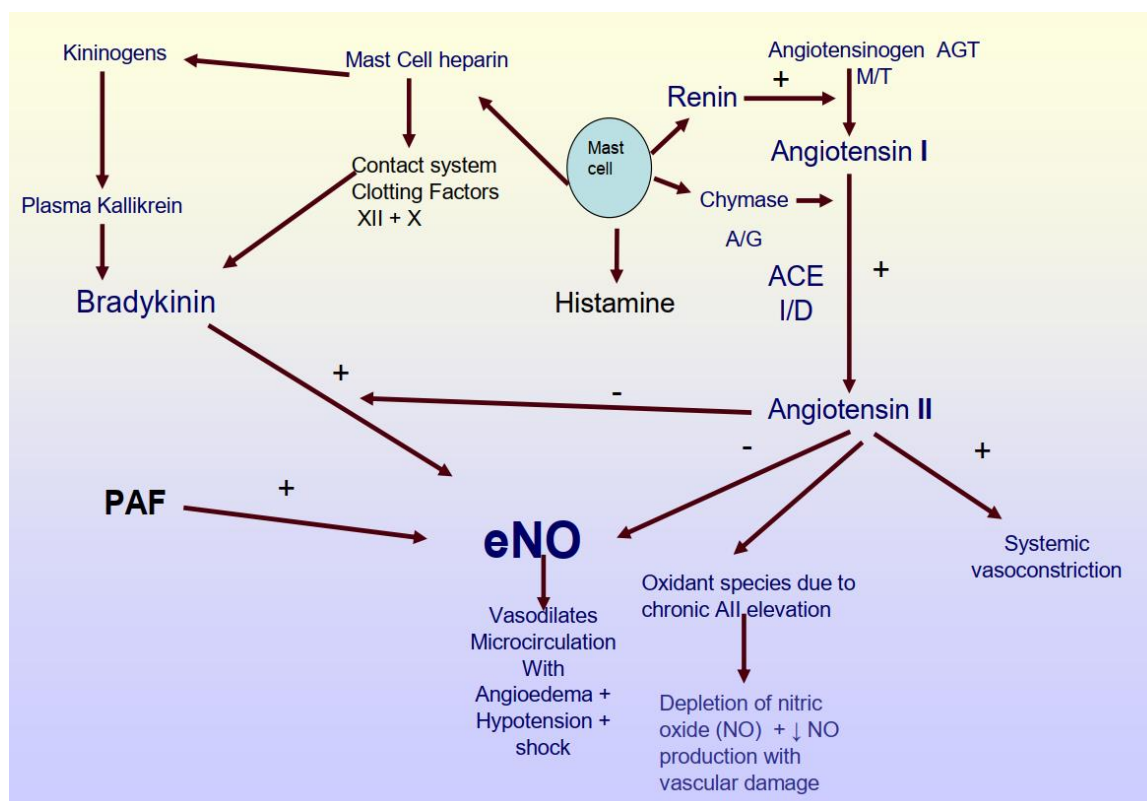


Figure 1. Renin-angiotensin system.

Rodent studies, suggest that platelet-activating factor (PAF) is also involved in anaphylaxis-induced cardiovascular collapse [56] that can be prevented by PAF antagonists [57,58]. In PAF-knockout mice, neither hypotension nor death in anaphylaxis occurs. However, PAF levels have been measured in man, and are reduced immediately following anaphylaxis, but are normal if measured away from the event [56,58]. PAF has been shown to further enhance mast cell histamine release and anaphylaxis in wildtype mice can be blocked by anti-histamines and a PAF antagonist. Animal models examining the depletion of monocytes and macrophage in anaphylaxis, suggests that they may also be the source of PAF along with mast cells [14,59,60]. PAF, like bradykinin may also act through endothelial nitric oxide (eNO) when mediating its hypotensive effect in anaphylaxis, as the nitric oxide-inhibitor L-NAME (L-Nitro Arginine Methyl Ester) can block severe hypotension and death induced by PAF. Endothelial NO is now of interest in murine models of anaphylaxis, where chronic blockade of eNO does increase the expression of mRNA for renin, ACE, and ATR1-receptors in the aorta suggesting a relationship [61]. Endothelial nitric oxide is a critical mediator of shock and death in animal models [62,63] and can be attenuated/prevented by endothelial nitric oxide inhibitors. Bradykinin also exerts a powerful arterial vasodilating effect on the microcirculation through stimulation of nitric oxide synthase that produces endothelial nitric oxide (eNO) (Figure 1).

2.4. Delayed release and newly synthesised mediators

In a more delayed manner MC activation leads to the release of several newly synthesised mediators such as prostaglandins and leukotrienes [64,65]. These encourage smooth muscle contraction and mucous formation that are central to the reduced bronchial airflow and asthma symptoms. Platelet activating factor (PAF) is also produced in a similar time frame [56] as are several cytokines including IL-1 β , IL6, IL31, IL33 [66,67], and TGF β amongst many others. Interestingly, several of these cytokines activate and stimulate the secretion of others in an amplification of inflammatory pathways. In this regard, TGF β can encourage Th17 cells [68] that promotes inflammation and IL-33 amplifies the effect of IgE on histamine release from mast cells and basophils [69–71]. IL33 also stimulates MCs to secrete IL-1 β , which then fuels IL6 production to promote inflammation [72]. Additionally, IL33 has also been shown to boost the facility of substance P (SP) to stimulate secretion of vascular endothelial growth factor by MCs and without inducing obvious degranulation [73]. IL31 is especially relevant to allergic symptomatology as it is highly potent at inducing itching [74] and thus adds to the pruritus caused by histamine and cutaneous discomfort produced by SP. Interestingly, activated MCs can also synthesise and secrete chemokines CCL2 and CCL8 that are important in inflammatory cell recruitment [75]. MC activation through Fc ϵ R1 can also stimulate autocrine hemokinin-1 synthesis and release, which via neurokinin 1 receptor stimulation can increase MC mediator release [76] and encourage renin production [77] with the latter being important in countering the hypotension of anaphylaxis.

2.5. Non-IgE stimulants of mast cell activation

MCs are not infrequently activated by specific IgG binding to Fc γ RII/III, activated complement proteins C3a/C5a binding to complement receptors CR3 and CR5 and by several low molecular agents [65]. The latter include several drugs [78–80], organophosphates, heavy metals, and specific neuropeptides such as corticotropin-releasing hormone [81], neurotensin (NT) [82,83], and substance P (SP). In the case of drug induced allergy/anaphylaxis, the reactions are often seen with the first dose. The increase in the occurrence of severe allergic reactions during periods of stress may be explained by the complex interaction and facilitation of MC activation by neuropeptides and these agents [84,85]. In regards to drug mediated allergic reactions, there is now abundant evidence supporting the importance of the low affinity G-protein-coupled receptor MRGPRX2 [86]. This becomes especially relevant in reactions occurring during anaesthesia and which cannot be identified following extensive skin prick and intradermal testing. Recent evidence also implicates this receptor in aetiology of chronic prurigo [87]. Reactions to opiate painkillers and codeine is thought to be through the surface mast cell opioid μ -receptor. These reactions often present with early itching and rash but maybe more florid in some cases but generally are easily treated with anti-histamines and steroids [88,89].

Reactions following intravenous radiocontrast injections and nanotechnology drugs (liposomal or micelle-solubilized drugs) [90] involve complement activation by direct complement C3 adsorption with conformational changes that resemble C3b, from which onward downstream complement activation of C3b convertase occurs. Occasionally, IgG + IgM antibodies are present or the complement component C1q is directly activated by the liposome [91]. Drugs in this category include the anti-fungal drug amphotericin (ambisome) and many chemotherapy drugs related to Taxol. Complement activation can be via the classical, lectin, or alternative pathway all of which can activate

Complement C3a and C5a giving surface activation of the mast cell and basophil surface complement receptors and mediator release [92,93]. C3a and C5a can induce respiratory distress with bronchoconstriction by direct binding to specific C3a and C5a receptors expressed on bronchial smooth muscle and epithelial cells in both mice and human lungs [92,93]. These anaphylatoxins (C3a and C5a) stimulate chemotaxis and mobilization of intracellular free Ca²⁺ in mast cells with C5a causing the rapid release of histamine and tryptase from mast cells.

3. Factors influencing mast cell activation and anaphylaxis

3.1. Blocking IgG antibodies

It is unclear why some individuals with allergen specific IgE experience life-threatening reactions while others have no symptoms. In this respect a possible protective effect of allergen-specific IgG which has a much higher concentration than allergen specific IgE has been discussed [94]. Elevation of allergen specific IgG4 is well described following both airborne allergen and venom immunotherapy even though its exact role is unclear. With food allergy, allergen-specific IgG levels are higher in individuals that are sensitized but unresponsive to the allergen or no longer respond to the food allergen as a result of oral allergen desensitization. Animal studies have demonstrated the presence of allergen specific IgG can inhibit the production of specific IgE by promoting the induction of T-regulatory cells to reduce immune responses. The factors driving IgE and IgG responses to ingested antigens are complex and not yet fully understood, but the gut microbiome may have an important role. Human and mouse MC and basophils express Fcγ receptors (FcγR's) that bind to IgG antibodies and their patterns of expression vary among leukocytes [95–97]. Unlike humans, mice monocytes appear to express FcγRI but do not express FcγRIIB, which is expressed by human B cells and basophils. Further, important differences include expression of FcγRIIIA by humans but not mice but is limited to NK cells and monocytes/macrophages. Additionally, FcγRIV exists in mice but not in humans and FcγRIIA, FcγRIIC and FcγRIIB exist in humans but not in mice. Overall, however, human mast cells and basophils are believed to express two IgG receptors (FcγRIIa and FcγRIIb) with FcγRIIb believed to be an inhibitory IgG receptor able to reduce activation of the mast cell and this is a focus for future research [98].

3.2. Oestrogen

Oestrogen's role in allergic disease was initially suggested by rodent studies [99]. Clinically, allergic disease is three times more common in females and more active especially during their peak reproductive years [100]. Furthermore, 30–40% of women with asthma have observed increased symptoms in the pre-menstrual phase when oestrogen and progesterone levels are changing and leukotriene C4 levels are shown to be elevated [101,102]. Mechanistically, human mast cells have mRNA for oestrogen receptor α but not β [103] and basophils and mast cells pre-incubated with physiological concentrations of oestrogen show increased histamine release following IgE cross-linking of the surface receptor [99] with mast cells similarly exposed undergoing partial degranulation. As expected this could be blocked by tamoxifen, suggesting a direct link to an oestrogen receptor [101]. Finally, the severity of anaphylaxis may be greater in females as oestrogen has an inhibitory effects on ACE biosynthesis [104,105] (see later).

3.3. Vitamin D

The role of Vitamin D in allergic and immune disease is interesting [106,107]. In the United States, prescriptions for auto-injecting adrenaline pens for anaphylaxis is 4-fold higher in the northern US states than the southern states where Vitamin D levels are lower from reduced sunlight exposure. Case reports also show benefit in reducing the symptoms of physical urticaria following supplementation of Vitamin D for severe cases. This benefit may be explained by ability of vitamin D3 to stabilize mast cells and reduces IgE dependant pro-inflammatory mediator release including reduced histamine release [108,109]. Moreover, 1,25 dihydroxy-Vitamin D receptors are present on mast cells, macrophages, T and B lymphocytes and other antigen presenting cells and it appears important for the functioning of the glucocorticoid receptor. Micro-array studies show that within the CD4+ T-lymphocyte alone, a total of 102 genes are targeted by Vitamin D with 57 being down-regulated and 45 up-regulated [110,111]. Within the skin, the presence of inflammation and inflammatory cytokines increases the local production of 1,25 hydroxy-Vitamin D usually by macrophages, which then reduces the production of inflammatory cytokines (including Interleukin-6,8,12,17,23 and tumor necrosis factor- α) along with the infiltration of neutrophils and eosinophils into the skin [112,113].

3.4. The renin-angiotensin system

In the early 1990s, Hermann and Ring showed a stepwise reduction in renin and angiotensin levels in patients with increasing severity of hymenoptera venom anaphylaxis (grade 1–3) [114,115]. Subsequently, Summers et al. [116] showed that patients with serum ACE levels below 37 mmol/L were 9.7 times more likely to develop pharyngeal oedema than those with mean serum ACE levels above 47 mmol/L [116]. With the widespread use of ACE-Inhibitor drugs, case reports of prolonged and profound anaphylaxis have appeared in the literature from the early 1990's [117]. Importantly, the ACE gene exists in 2 allelic forms, with the presence of "I" (insertion) or its absence "D" (deletion) of a 287 base pair intron of the gene. The "D" genotype is associated with higher ACE activity and AII levels and increased plasma bradykinin catabolism and individuals with the homozygous DD genotype have the highest serum ACE levels while the I/I or I/D genotype has the lowest serum ACE and AII levels [118]. In this respect, Niedoszytko et al. [119] showed 80% of 30 patients with insect venom allergy of grades 3–4 on the Mueller scale [120] had the ID or II genotypes along with 50% lower levels of serum ACE activity and higher basal bradykinin levels. Similarly, we found II/ID genes linked to anaphylaxis involving hypotension and angioedema and associated with lower serum ACE levels relative to both atopics and healthy controls [118]. This may be explained by serum ACE being important in the catabolism of bradykinin, which can cause angioedema and hypotension [121] (Figure 1). Additionally, we noted that gene polymorphisms that coded for low activity of RAS were more prevalent in severe anaphylaxis [118]. These genes included ACE, Renin, angiotensinogen, angiotensin Receptor-1, Chymase, and the bradykinin B2-receptor gene [122].

4. Diagnosing mast cell dysfunction and anaphylaxis

The clinical history is critical and aims to check for the symptoms and signs related to the release of mast cell mediators previously discussed. The clinical features of anaphylaxis need assessment as

well as a search for any causative and contributory factors for mast cell activation such as alcohol ingestion, exercise, NSAID usage, and stress. However, activated mast cells release tryptase and detecting raised serum levels 2–4 hours later can be very helpful in suspected anaphylaxis. In contrast, the release of mast cell chymase in allergic reactions is largely overlooked but warrants discussion. Blood Chymase levels increase after 1 hour following allergic reactions and remain raised for 8–24 hrs. Autopsy studies show mean Chymase blood levels of 89.8 ng/mL in anaphylaxis compared with <3 ng/mL in cases without anaphylaxis [123,124].

5. Mast cell disorders

5.1. Broad categorization

Disorders arising from mast cell mediator release may be differentiated into those with increased numbers of mast cells (mastocytosis) and those with aberrantly raised function (mast cell activation syndrome) (Table 2). There is, however, a continuum of variation between the two that influences the clinical presentation and especially with features of anaphylaxis [23,125] and especially idiopathic anaphylaxis [126]. Notwithstanding, genetic and hormonal predisposition is likely important [125,127–129] as is the circadian facility of MC mediator release [41,130].

From a clinical perspective, three sets of criteria are required to suggest a mast cell problem [23,126,127].

(1) Episodic symptoms of mast cell activation eg itchy rashes, flushing, bronchospasm, abdominal discomfort and diarrhoea, faintness/syncope etc.

(2) Laboratory evidence of released mast cell mediators in blood and urine related to these episodes such as raised serum tryptase levels at baseline and within 4 hours of an acute event.

(3) Symptomatic improvement after regular MC mediator antagonist therapy, MC reduction, and/or stabilizing treatment.

5.2. Mastocytosis

Here there is clonal proliferation of mast cells in several organs, including the skin (urticaria pigmentosa, maculopapular areas, or mastocytomas), bone marrow, and gastrointestinal tract, (liver, spleen, lymph nodes). This is usually caused by mutations of the c-KIT receptor [131] that potentiates mast cell activation by IgE and non-IgE receptor mechanisms. There is also constitutive activation of the MC with mediator release leading to systemic symptoms and even anaphylaxis. Somatic missense mutation involving substitution of aspartic acid (D) to valine (V) at amino acid B16 in exon 17 is the most frequent mutation but others are also recognized. C-KIT binding to the stem cell factor receptor (a tyrosine kinase receptor) initiates a signaling cascade within the mast cells that regulates their growth, migration, and proliferation. In consequence, increased c-KIT activity can be associated with increased mast cell numbers (mastocytosis) and neoplastic disorders of mast cell expansion in man due to its oncogene properties [131,132]. In addition to c-KIT, there are also other markers of mast cell irritability including CD25 (alpha chain) of the IL-2 receptor or CD2 (lymphocyte function antigen-2) present on neoplastic mast cells. More variants will no doubt be described but the exact details of how this makes the mast hyperactive are currently sketchy [133,134].

The World Health Organization has classified mastocytosis into 2 major groups:

1. Cutaneous mastocytosis
2. Systemic mastocytosis (involving at least 1 extracutaneous organ).

The true prevalence of mastocytosis is unknown and probably goes unrecognized in milder cases, but is estimated to be 9–13 per 100,000 [135,136]. Cutaneous mastocytosis is commonest in young children and is often a skin-limited disease that regresses with time. Fuchs et al. [137] have proposed a scoring system allowing the prediction of systemic mastocytosis (SM) in those with cutaneous mastocytosis and based on the tryptase level as well as constitutional/cardiovascular symptoms and bone pain/osteoporosis. In adults, multi-organ involvement is more common, can be checked by biopsy examination of skin and the bone marrow, and may give more symptoms with a possible progression to mast cell leukemia. This was considered more likely in the presence of activating C-kit and a range of other mutations [138] which clearly affects prognosis [139]. Regardless, 85% of subjects with SM the condition is indolent and symptomatic patients have local or systemic episodes of mast cell mediator release (including histamine, proteases, leukotrienes, and prostaglandins). Often, there are recognized triggers for the “Mast Cell Release Episodes” such as physical exertion, heat and cold, insect stings, alcohol consumption, oral non-steroidal anti-inflammatory drugs, and emotional distress. Specific triggers vary between patients but generally symptoms predominately include flushing, pruritus, palpitations, dizziness, hypotension, and syncope. Additional reported symptoms may include breathing difficulties, abdominal pain and diarrhea. A history of flushing is a cardinal symptom in many of these patients that should alert suspicion. Some subjects may experience symptoms resembling anaphylaxis that may last 15–30 minutes although some episodes may be life threatening due to severe hypotension [23,126,135].

5.3. *Secondary Mast cell activation syndrome*

Secondary MC activation through IgE and non-IgE-mediated processes (food, drugs, venom) can produce variable allergy type symptoms and systemic anaphylaxis without evidence of a clonal mast cell population and without evidence of a primary mast cell activation disorder. Symptoms are episodic and similar to those seen in mastocytosis as detailed above [23,140,141]. Interestingly, there is evidence that MC irritability leading to non-allergic mediator release is more frequent in those with joint hypermobility syndrome [142,143] although the precise mechanism of this association has yet to be defined. While a number of non-allergic symptoms such as fatigue, headaches, myalgia etc have now been associated with this secondary MCAS, these are unlikely to be due to the release of preformed mediators but may possibly arise from inflammatory cytokine release [143]. Regardless, the diagnosis of MCAS is based on a combination of several suggestive symptoms in the clinical history supplemented by urine tests looking for elevated levels of methylhistamine, leukotriene E4, and 2,3-dinor-11beta-prostaglandin F2 alpha [144]. The precise role of gastrointestinal dysbiosis, increased permeability of the intestinal epithelium and the absorption of microbial factors able to stimulate MCs and leading to constitutional symptoms and allergic type rashes is certainly interesting and warrants further in depth investigation [145].

5.4. *Hereditary alpha-tryptasaemia*

This is found in 5–6% of the general population and is an autosomal dominant condition producing excess copies of the alpha-tryptase gene (TPSAB1). It is a common cause of elevated basal

serum tryptase levels >8 ng/ml, due to raised pro-alpha tryptase synthesis rather than increased mast cell activation [146,147]. It is associated with some increase in of mast cell numbers in the bone marrow and gut and some increase in the urinary excretion of mast cell mediators such as methylhistamine, 9α - 11β -Prostaglandin F₂ and the breakdown product PGDM. Its prevalence is increased in both clonal and non-clonal disease suggesting an assessment of its presence is made in the evaluation of mast cell disorders [148]. The risk for severe spontaneous or insect venom-triggered anaphylaxis is reported to be increased in this condition [149,150].

6. Treatment options for anaphylaxis and mast cell disorders

Adrenaline is first line for acute anaphylaxis [151–153] in mast cell disorders. In refractory cases of severe hypotension not responding to repeated doses of intramuscular epinephrine or where cardiac arrest has occurred, intravenous epinephrine should be given with continuous monitoring of the cardiac response, blood pressure, and oxygen saturation. Anti-histamines, oxygen and fluid replacement should be given once the cardiovascular status is stabilized then beta2-agonists and corticosteroids are usually recommended [153].

Long term management is aimed at the prevention of repeat episodes of anaphylaxis including specific IgE and skin tests for any potential triggers and discussion of foods, medication, inhalational triggers and cofactors such as alcohol, exercise, and non-steroidal anti-inflammatory drugs (NSAIDs) to be avoided. Elimination of histamine-rich foods is not routinely recommended. Those sensitized to venom should have at least 3 years and in some cases lifelong venom immunotherapy to reduce recurrent anaphylaxis risk following a sting [154].

There are no randomized studies to show which prophylactic therapy options are superior for mastocytosis. A stepwise approach is important with step one utilizing increasing doses of oral antihistamines (H₁ receptor blockers) up to 4 times standard levels if required. Histamine receptor-2 blockers and anti-leukotrienes drugs can be added, with oral sodium cromoglycate and corticosteroids if patients remain unresponsive. If these combination therapies are ineffective, then omalizumab, the humanized monoclonal antibody that specifically binds to free IgE, can be used and has been shown to diminish the frequency of anaphylactic reactions in a case series [155] ‘Cytoreductive’ therapy can be used in systemic mastocytosis, which now commonly employs one of the tyrosine kinase inhibitors to target the mast cell growth receptor c-KIT. Both midostaurin and avapritinib are currently approved for this treatment and may give prompt resolution of recurrent anaphylaxis although side effects are not infrequent [156] (Gotlib et al, 2021). There are also reports that this treatment reduces splenomegaly and also bone marrow mast cell numbers in cases with advanced systemic mastocytosis [157,158].

7. Conclusions

The mast cell and its role in allergic and indeed non-allergic diseases as well as its wide distribution throughout the body, including the myocardium, central nervous system, and elsewhere continues to expand its importance in a diverse range of human illnesses. Much has been written about the early and rapidly acting mediators released by activated MCs in the context of localized and systemic allergic reactions. However, there is increased appreciation that they synthesize and release a wide range of inflammatory cytokines that can perpetuate inflammation and perhaps also explain the constitutional symptoms seen in association with allergic reactions and help clarify its role in stress

and non-parasitic infections. The data on environmental, menstrual, circadian, and psychological factors influencing the onset and severity of anaphylaxis needs further investigation with its links to mast cell disorders in which normal numbers of MCs appear to be more irritable and prone to activation by life factors and agents that are normally tolerated. The precise role of specific MC receptors such as the MRGPX2 in this irritability needs definition. Consensus on the exact clinical and laboratory features of idiopathic mast cell activation syndrome is urgently needed to avoid this term being used for all illnesses with vague symptoms in which a cause is not apparent. This may open a new chapter in human diseases and mast cell regulation. The increasing incidence of anaphylaxis within a relatively short time frame also needs to be explained in terms of rising stress levels, dysbiosis in the gastrointestinal tract, and increased use of additives and colorings in food. It would certainly be more helpful to delineate the precise mode of interaction between immune genes and the environment.

Conflict of interest

All authors declare no conflicts of interest in this paper.

References

1. Bjornsson HM, Graffeo CS (2010) Improving diagnostic accuracy of anaphylaxis in acute care setting. *West J Emerg Med* 11: 456–461.
2. Tejeclor A, Lonson MA, Moromoro M, et al. (2014) Epidemiology of anaphylaxis. *Clin Exp Allergy* 45: 1027–1039. <https://doi.org/10.1111/cea.12418>
3. Mullins RJ, Wainstein BK, Barnes EH, et al. (2016) Increases in anaphylaxis fatalities in Australia from 1997–2013. *Clin Exp Allergy* 46: 1099–1110. <https://doi.org/10.1111/cea.12748>
4. Sheikh A, Alves B (2001) Age, sex, geographical and socio-economic variations in admissions for anaphylaxis: analysis of four years of English hospital data. *Clin Exp All* 31: 1571–1576. <https://doi.org/10.1046/j.1365-2222.2001.01203.x>
5. Braganza SC, Acworth JP, McKinnon DRL, et al. (2006) Paediatric emergency department anaphylaxis: different patterns from Adults. *Arch Dis Child* 91: 159–163. <https://doi.org/10.1136/adc.2004.069914>
6. Macdougall CF, Cart AJ, Clover AF (2002) How dangerous is food allergy in childhood? The incidence of severe and fatal allergic reactions across UK and Ireland. *Arch Dis Child* 86: 236–237. <https://doi.org/10.1136/adc.86.4.236>
7. Vetander M, Protudjer JLP, Liija G, et al. (2016) Anaphylaxis to foods in a population of adolescents: incidence, characteristics and associated risks. *Clin Exp Allergy* 46: 1575–1587. <https://doi.org/10.1111/cea.12842>
8. Terr AI (1985) Anaphylaxis. *Clin Rev Allergy* 3: 3–23. <https://doi.org/10.1007/BF02993040>
9. Delage C, Irely NS (1972) Anaphylactic deaths: a Clinicopathologic study of 43 cases. *J Forensic Sci* 17: 525–540. <https://doi.org/10.1520/JFS10141J>
10. Greenberger PA, Rotskoff BD, Lifschultz B (2007) Fatal anaphylaxis: postmortem findings and associated comorbid diseases. *Ann Allergy Asthma Immunol* 98: 252–257. [https://doi.org/10.1016/S1081-1206\(10\)60714-4](https://doi.org/10.1016/S1081-1206(10)60714-4)
11. Smith PK, Hourihane JO, Lieberman P (2005) Risk multipliers for severe food anaphylaxis. *World Allergy Organ J* 8: 30. <https://doi.org/10.1186/s40413-015-0081-0>

12. Pumphrey R, Stanworth SJ (1996) The clinical spectrum of anaphylaxis in north-west England. *Clin Exp All* 26: 1364–1370. <https://doi.org/10.1111/j.1365-2222.1996.tb00537.x>
13. Pumphrey RS (2000) Lessons for management of anaphylaxis from a study of fatal reactions. *Clin Exp Allergy* 30: 1144–1150. <https://doi.org/10.1046/j.1365-2222.2000.00864.x>
14. Reber LL, Hernandez JD, Galli SJ (2007) The pathophysiology of anaphylaxis. *J Allergy Clin Immunol* 140: 335–348. <https://doi.org/10.1016/j.jaci.2017.06.003>
15. Gupta R, Sheikh A, Strachan DP (2007) Time trends in allergic disorders in the UK. *Thorax* 62: 91–96. <https://doi.org/10.1136/thx.2004.038844>
16. Berlin MC (2015) Pathogenesis of IgE mediated food allergy. *Clin Exp Allergy* 45: 1483–1496. <https://doi.org/10.1111/cea.12598>
17. Harper NJN, Cook TM, Garcez T, et al. (2018) Anaesthesia, surgery and life-threatening allergic reactions: Management and outcomes in the 6th National Audit Project (NAP6). *Br J Anaesth* 121: 159–171. <https://doi.org/10.1016/j.bja.2018.04.015>
18. Sato T, Morishita S, Horie T, et al. (2019) Involvement of premacular mast cells in the pathogenesis of macular diseases. *PLoS One* 14: e0211438. <https://doi.org/10.1371/journal.pone.0211438>
19. Lauritano D, Mastrangelo F, D'Ovidio C, et al. (2023) Activation of mast cells by neuropeptides: The role of pro-inflammatory and anti-inflammatory cytokines. *Int J Mol Sci* 24: 4811. <https://doi.org/10.3390/ijms24054811>
20. Nishino S, Sakai N, Nishino N, et al. (2022) Brain mast cells in sleep and behavioral regulation. *Curr Top Behav Neurosci* 59: 427–446. https://doi.org/10.1007/7854_2022_359
21. Theoharides TC, Valent P, Akin C (2015) Mast cells, mastocytosis, and related disorders. *N Engl J Med* 373: 163–172. <https://doi.org/10.1056/NEJMra1409760>
22. Leguit RJ, Wang SA, George TI, et al. (2023) The international consensus classification of mastocytosis and related entities. *Virchows Arch* 482: 99–112. <https://doi.org/10.1007/s00428-022-03423-3>
23. Gülen T, Akin C, Bonadonna P, et al. (2021) Selecting the right criteria and proper classification to diagnose mast cell activation syndromes: A critical review. *J Allergy Clin Immunol Pract* 9: 3918–3928. <https://doi.org/10.1016/j.jaip.2021.06.011>
24. Valent P, Hartmann K, Bonadonna P, et al. (2022) Mast cell activation syndromes: Collegium internationale allergologicum update. *Int Arch Allergy Immunol* 183: 693–705. <https://doi.org/10.1159/000524532>
25. Hellman L, Akula S, Fu Z, et al. (2022) Mast cell and basophil granule proteases - *In Vivo* targets and function. *Front Immunol* 13: 918305. <https://doi.org/10.3389/fimmu.2022.918305>
26. Bian G, Gu Y, Xu C, et al. (2021) Early development and functional properties of tryptase/chymase double-positive mast cells from human pluripotent stem cells. *J Mol Cell Biol* 13: 104–115. <https://doi.org/10.1093/jmcb/mjaa059>
27. Komi DEA, Rambasek T, Wöhrl S (2018) Mastocytosis: From a molecular point of view. *Clin Rev Allergy Immunol* 54: 397–411. <https://doi.org/10.1007/s12016-017-8619-2>
28. Irani AA, Schechter NM, Craig SS, et al. (1986) Two types of human mast cells that have distinct neutral protease compositions. *Proc Natl Acad Sci USA* 83: 4464–4468. <https://doi.org/10.1073/pnas.83.12.4464>

29. Siddhuraj P, Clausson CM, Sanden C, et al. (2021) Lung mast cells have a high constitutive expression of carboxypeptidase A3 mRNA that is independent from granule-stored CPA3. *Cells* 10: 309. <https://doi.org/10.3390/cells10020309>
30. Donelan J, Boucher W, Papadopoulou N, et al. (2006) Corticotropin-releasing hormone induces skin vascular permeability through a neurotensin-dependent process. *Proc Natl Acad Sci USA* 103: 7759–7764. <https://doi.org/10.1073/pnas.0602210103>
31. Mustain WC, Rychahou PG, Evers BM (2011) The role of neurotensin in physiologic and pathologic processes. *Curr Opin Endocrinol Diabetes Obes* 18: 75–82. <https://doi.org/10.1097/MED.0b013e3283419052>
32. Gaudenzio N, Sibilano R, Marichal T, et al. (2016) Different activation signals induce distinct mast cell degranulation strategies. *J Clin Invest* 126: 3981–3998. <https://doi.org/10.1172/JCI85538>
33. Cianferoni A (2021) Non-IgE-mediated anaphylaxis. *J Allergy Clin Immunol* 147: 1123–1131. <https://doi.org/10.1016/j.jaci.2021.02.012>
34. Poto R, Quinti I, Marone G, et al. (2022) IgG autoantibodies against IgE from atopic dermatitis can induce the release of cytokines and proinflammatory mediators from basophils and mast cells. *Front Immunol* 13: 880412. <https://doi.org/10.3389/fimmu.2022.880412>
35. Wernersson S, Pejler G (2014) Mast cell secretory granules: armed for battle. *Nat Rev Immunol* 14: 478–494. <https://doi.org/10.1038/nri3690>
36. Lee J, Vadas P (2011) Anaphylaxis: mechanisms and management. *Clin Exp Allergy* 41: 923–938. <https://doi.org/10.1111/j.1365-2222.2011.03779.x>
37. Lundequist A, Pejler G (2011) Biological implications of preformed mast cell mediators. *Cell Mol Life Sci* 68: 965–975. <https://doi.org/10.1007/s00018-010-0587-0>
38. Theoharides TC, Kempuraj D, Tagen M, et al. (2007) Differential release of mast cell mediators and the pathogenesis of inflammation. *Immunol Rev* 217: 65–78. <https://doi.org/10.1111/j.1600-065X.2007.00519.x>
39. Morita H, Nakae S, Saito H, et al. (2017) IL-33 in clinical practice: Size matters? *J Allergy Clin Immunol* 140: 381–383. <https://doi.org/10.1016/j.jaci.2017.03.042>
40. Zhang B, Weng Z, Sismanopoulos N, et al. (2012) Mitochondria distinguish granule-stored from de novo synthesized tumor necrosis factor secretion in human mast cells. *Int Arch Allergy Immunol* 159: 23–32. <https://doi.org/10.1159/000335178>
41. Nakao A, Nakamura Y, Shibata S (2015) The circadian clock functions as a potent regulator of allergic reaction. *Allergy* 70: 467–473. <https://doi.org/10.1111/all.12596>
42. Askenase PW (2005) Mast cells and the mediation of T-cell recruitment in arthritis. *N Engl J Med* 349: 1294. <https://doi.org/10.1056/NEJM200309253491319>
43. Gordon JR, Galli SJ (1990) Mast cells as a source of both preformed and immunologically inducible TNF- α /cachectin. *Nature* 346: 274–276. <https://doi.org/10.1038/346274a0>
44. Zhang B, Alysandratos KD, Angelidou A, et al. (2011) Human mast cell degranulation and preformed TNF secretion require mitochondrial translocation to exocytosis sites: relevance to atopic dermatitis. *J Allergy Clin Immunol* 127: 1522–1531.e8. <https://doi.org/10.1016/j.jaci.2011.02.005>
45. Panula P (2021) Histamine receptors, agonists, and antagonists in health and disease. *Handb Clin Neurol* 180: 377–387. <https://doi.org/10.1016/B978-0-12-820107-7.00023-9>

46. Ahmad S, Varagic J, Groban L, et al. (2014) Angiotensin (1–12); A chymase mediated cellular angiotensin II substrate. *Curr Hypertens Res* 16: 429–437. <https://doi.org/10.1007/s11906-014-0429-9>
47. He S, Walls AF (1998) The induction of a prolonged increase in microvascular permeability by human mast cell chymase. *Eur J Pharmacol* 352: 91–98. [https://doi.org/10.1016/S0014-2999\(98\)00343-4](https://doi.org/10.1016/S0014-2999(98)00343-4)
48. Wong CK, Ng SSM, Lun SWM, et al. (2009) Signaling mechanisms regulating the activation of human eosinophils by mast-cell-derived Chymase; implications for mast-cell-eosinophil interaction in allergic inflammation. *Immunology* 126: 579–587. <https://doi.org/10.1111/j.1365-2567.2008.02916.x>
49. Caughey GH (2007) Mast cell tryptases and chymases in inflammation and host defense. *Immunol Rev* 217: 141–154. <https://doi.org/10.1111/j.1600-065X.2007.00509.x>
50. Saarinen JV, Harvima RJ, Naukkarinen A, et al. (2001) The release of histamine is associated with the inactivation of mast cell chymase during immediate allergic wheal reactions in the skin. *Clin Exp Allergy* 31: 593–601. <https://doi.org/10.1046/j.1365-2222.2001.01030.x>
51. Guilarte M, Sala-Cunill A, Luengo O, et al. (2017) The mast cell contact and coagulation system connection in anaphylaxis. *Front Immunol* 8: 846. <https://doi.org/10.3389/fimmu.2017.00846>
52. Proud D, Togias A, Nacleiro RM, et al. (1983) Kinins are generated in vivo following airway challenge of allergic individuals with allergen. *J Clin Invest* 72: 1678–1685. <https://doi.org/10.1172/JCI111127>
53. Stone SF, Brown SG (2012) Mediators released during human anaphylaxis. *Curr Allergy Asthma Rev* 12: 33–41. <https://doi.org/10.1007/s11882-011-0231-6>
54. Caughey GH (2016) Mast cell proteases as pharmacological targets. *Eur J Pharmacol* 778: 44–55. <https://doi.org/10.1016/j.ejphar.2015.04.045>
55. Imamura T, Dubin A, Moore W, et al. (1996) Induction of vascular permeability enhancement by human tryptase: dependence on activation of prekallikrein and direct release of bradykinin from kininogens. *Lab Invest* 74: 861–870.
56. Vadas P, Gold M, Perelman B, et al. (2008) Platelet-activating factor, PAF acetylhydrolase, and severe anaphylaxis. *N Engl J Med* 358: 28–35. <https://doi.org/10.1056/NEJMoa070030>
57. Fukuda Y, Kawashima H, Saito K, et al. (2000) Effect of human plasma-type platelet activating factor acetylhydrolase in two anaphylactic shock models. *Eur J Pharmacol* 390: 203–207. [https://doi.org/10.1016/S0014-2999\(99\)00920-6](https://doi.org/10.1016/S0014-2999(99)00920-6)
58. Arimura A, Harada M (1991) Differential effect of a PAF antagonist CV- 3988 on active and passive anaphylactic shock in various mouse strains. *Lipids* 26: 1386–1390. <https://doi.org/10.1007/BF02536572>
59. Gill P, Jindal NL, Jagdis A, et al. (2015) Platelets in the immune response; Revisiting platelet activating factor in anaphylaxis. *J Allergy Clin Immunol* 135: 1424–1432. <https://doi.org/10.1016/j.jaci.2015.04.019>
60. Kajiwarra N, Sasaki T, Braddind P, et al. (2010) Activation of human mast cells through the platelet-activating factor receptor. *J Allergy Clin Immunology* 125: 1137–1145. <https://doi.org/10.1016/j.jaci.2010.01.056>
61. Dao VT, Medini S, Bisha M, et al. (2016) Nitric oxide up-regulates endothelial expression of angiotensin II type 2 receptors. *Biochem Pharmacol* 112: 24–36. <https://doi.org/10.1016/j.bcp.2016.05.011>

62. Mombouli JV, Vanhoutte PM (1992) Heterogeneity of endothelium-dependent vasodilator effects of angiotensin converting enzyme inhibitors: role of bradykinin generation during ACE inhibition. *J Cardiovasc Pharmacol* 20: S74–S82. <https://doi.org/10.1097/00005344-199200209-00014>
63. Lowenstein CJ, Michel T (2006) What's in a name? eNOS and anaphylactic shock. *J Clin Invest* 116: 2075–2078. <https://doi.org/10.1172/JCI29406>
64. Kuehn HS, Radinger M, Gilfillan AM (2010) Measuring mast cell mediator release. *Curr Protoc Immunol* 2 Chapter 7: Unit 7.38. <https://doi.org/10.1002/0471142735.im0738s91>
65. Nguyen SMT, Rupprecht CP, Haque A, et al (2021) Mechanisms governing anaphylaxis: inflammatory cells, mediators, endothelial gap junctions and beyond. *Int J Mol Sci* 22: 7785. <https://doi.org/10.3390/ijms22157785>
66. Dileepan KN, Raveendran VV, Sharma R (2023) Mast cell-mediated immune regulation in health and disease. *Front Med (Lausanne)* 10: 1213320. <https://doi.org/10.3389/fmed.2023.1213320>
67. Mukai K, Tsai M, Saito, et al. (2018) Mast cells as sources of cytokines, chemokines, and growth factors. *Immunol Rev* 282: 121–150. <https://doi.org/10.1111/imr.12634>
68. Noordenbos T, Blijdorp I, Chen S, et al. (2016) D. Human mast cells capture, store, and release bioactive, exogenous IL-17A. *J Leukoc Biol* 100: 453–462. <https://doi.org/10.1189/jlb.3HI1215-542R>
69. Moulin D, Donzé O, Talabot-Ayer D, et al. (2007) Interleukin (IL)-33 induces the release of pro-inflammatory mediators by mast cells. *Cytokine* 40: 216–225. <https://doi.org/10.1016/j.cyto.2007.09.013>
70. Saluja R, Ketelaar ME, Hawro T, et al. (2015) The role of the IL-33/IL-1RL1 axis in mast cell and basophil activation in allergic disorders. *Mol Immunol* 63: 80–85. <https://doi.org/10.1016/j.molimm.2014.06.018>
71. Silver MR, Margulis A, Wood N, et al. (2010) IL-33 synergizes with IgE-dependent and IgE-independent agents to promote mast cell and basophil activation. *Inflamm Res* 59: 207–218. <https://doi.org/10.1007/s00011-009-0088-5>
72. Kandere-Grzybowska K, Letourneau R, Kempuraj D, et al. (2003) IL-1 induces vesicular secretion of IL-6 without degranulation from human mast cells. *J Immunol* 171: 4830–4836. <https://doi.org/10.4049/jimmunol.171.9.4830>
73. Theoharides TC, Zhang B, Kempuraj D, et al. (2010) IL-33 augments substance P-induced VEGF secretion from human mast cells and is increased in psoriatic skin. *Proc Natl Acad Sci USA* 107: 4448–4453. <https://doi.org/10.1073/pnas.1000803107>
74. Petra AI, Tsilioni I, Taracanova A, et al. (2018) Interleukin 33 and interleukin 4 regulate interleukin 31 gene expression and secretion from human laboratory of allergic diseases 2 mast cells stimulated by substance P and/or immunoglobulin E. *Allergy Asthma Proc* 39: 153–160. <https://doi.org/10.2500/aap.2018.38.4105>
75. Salamon P, Shoham NG, Gavrieli R, et al. (2005) Human mast cells release Interleukin-8 and induce neutrophil chemotaxis on contact with activated T cells. *Allergy* 60: 1316–1319. <https://doi.org/10.1111/j.1398-9995.2005.00886.x>
76. Sumpter TL, Ho CH, Pleet AR, et al. (2015) Autocrine hemokinin-1 functions as an endogenous adjuvant for IgE-mediated mast cell inflammatory responses. *J Allergy Clin Immunol* 135: 1019–1030.e8. <https://doi.org/10.1016/j.jaci.2014.07.036>
77. Conti P, Caraffa AI, Kritas SK, et al. (2017) Mast cell, pro-inflammatory and anti-inflammatory: Jekyll and Hyde, the story continues. *J Biol Regul Homeost Agents* 31: 263–267.

78. Bruhns P, Chollet-Martin S (2021) Mechanisms of human drug-induced anaphylaxis. *J Allergy Clin Immunol* 147: 1133–1142. <https://doi.org/10.1016/j.jaci.2021.02.013>
79. Zhang B, Li Q, Shi C, et al. (2018) Drug-induced pseudoallergy: A review of the causes and mechanisms. *Pharmacology* 101: 104–110. <https://doi.org/10.1159/000479878>
80. Yu Y, Blokhuis BR, Garssen J, et al. (2016) Non-IgE mediated mast cell activation. *Eur J Pharmacol* 778: 33–43. <https://doi.org/10.1016/j.ejphar.2015.07.017>
81. Cao J, Papadopoulou N, Kempuraj D, et al. (2005) Human mast cells express corticotropin-releasing hormone (CRH) receptors and CRH leads to selective secretion of vascular endothelial growth factor. *J Immunol* 174: 7665–7675. <https://doi.org/10.4049/jimmunol.174.12.7665>
82. Caceda R, Kinkead B, Nemeroff CB (2006) Neurotensin: role in psychiatric and neurological diseases. *Peptides* 27: 2385–2404. <https://doi.org/10.1016/j.peptides.2006.04.024>
83. Lazarus LH, Perrin MH, Brown MR, et al. (1977) Verification of both the sequence and conformational specificity of neurotensin in binding to mast cells. *Biochem Biophys Res Commun* 76: 1079–1085. [https://doi.org/10.1016/0006-291X\(77\)90966-4](https://doi.org/10.1016/0006-291X(77)90966-4)
84. Theoharides TC (2017) Neuroendocrinology of mast cells: challenges and controversies. *Exp Dermatol* 26: 751–759. <https://doi.org/10.1111/exd.13288>
85. Theoharides TC, Cochrane DE (2004) Critical role of mast cells in inflammatory diseases and the effect of acute stress. *J Neuroimmunol* 146: 1–12. <https://doi.org/10.1016/j.jneuroim.2003.10.041>
86. McNeil BD, Pundir P, Meeker S, et al. (2015) Identification of a mast-cell-specific receptor crucial for pseudo-allergic drug reactions. *Nature* 519: 237–241. <https://doi.org/10.1038/nature14022>
87. Kolkhir P, Elieh-Ali-Komi D, Metz M, et al. (2022) Understanding human mast cells: lesson from therapies for allergic and non-allergic diseases. *Nat Rev Immunol* 22: 294–308. <https://doi.org/10.1038/s41577-021-00622-y>
88. Yoo HS, Yang EM, Kim MA, et al. (2014) A case of codeine induced anaphylaxis via oral route. *Allergy Asthma Immunol Res* 6: 95–97. <https://doi.org/10.4168/aair.2014.6.1.95>
89. Casale TB, Bowman S, Kaliner M (1984) Induction of human cutaneous mast cell degranulation by opiates and endogenous opioid peptides: evidence for opiate and nonopiate receptor participation. *J Allergy Clin Immunol* 73: 775–781. [https://doi.org/10.1016/0091-6749\(84\)90447-0](https://doi.org/10.1016/0091-6749(84)90447-0)
90. Logothetidis S (ed). (2014) *Nanomedicine and Nanobiotechnology*. Berlin, Heidelberg: Springer. Available from: <https://doi.org/10.1007/978-3-642-24181-9>.
91. Dézsi L, Fülöp T, Mészáros T, et al. (2014) Features of complement activation-related pseudoallergy to liposomes with different surface charge and PEGylation: comparison of the porcine and rat responses. *J Control Release* 195: 2–10. <https://doi.org/10.1016/j.jconrel.2014.08.009>
92. Drouin SM, Kildsgaard J, Haviland J, et al. (2001) Expression of the complement anaphylatoxin C3a and C5a receptors on bronchial epithelial and smooth muscle cells in models of sepsis and asthma. *J Immunol* 166: 2025–2032. <https://doi.org/10.4049/jimmunol.166.3.2025>
93. Hartmann K, Henz BM, Krüger-Krasagakes S, et al. (1997) C3a and C5a stimulate chemotaxis of human mast cells. *Blood* 89: 2863–2870. <https://doi.org/10.1182/blood.V89.8.2863>
94. Kanagaratham C, El Ansari YS, Lewis OL, et al. (2020) IgE and IgG antibodies as regulators of mast cell and basophil functions in food allergy. *Front Immunol* 11: 603050. <https://doi.org/10.3389/fimmu.2020.603050>
95. Nimmerjahn F, Bruhns P, Horiuchi K, et al. (2005) Fcγ₄R: a novel FcR with distinct IgG subclass specificity. *Immunity* 23: 41–51. <https://doi.org/10.1016/j.immuni.2005.05.010>

96. Bruhns P, Jonsson F (2015) Mouse and human FcR effector functions. *Immunol Rev* 268: 25–51. <https://doi.org/10.1111/imr.12350>
97. Nimmerjahn F, Ravetch JV (2008) Fc gamma receptors as regulators of immune responses. *Nat Rev Immunol* 8: 34–47. <https://doi.org/10.1038/nri2206>
98. Nimmerjahn F, Ravetch JV (2006) Fc gamma receptors: old friends and new family members. *Immunity* 24: 19–28. <https://doi.org/10.1016/j.immuni.2005.11.010>
99. Cocchiara R, Albegiani G, Di Trapani G (1990) Modulation of rat peritoneal mast cell and human basophil histamine release by estrogens. *Int Arch Allergy Appl Immunol* 93: 192–197. <https://doi.org/10.1159/000235300>
100. Nakasato H, Ohru T, Sekizawa K (1999) Prevention of severe premenstrual asthma attacks by leukotriene receptor antagonist. *J Allergy Clin Immunol* 104: 585–588. [https://doi.org/10.1016/S0091-6749\(99\)70327-1](https://doi.org/10.1016/S0091-6749(99)70327-1)
101. Vliagoftis H, Dimitriadou V, Boucher W, et al. (1992) Estradiol augments while tamoxifen inhibits rat mast cell secretion. *Int Arch Allergy Immunol* 98: 398–409. <https://doi.org/10.1159/000236217>
102. Vrieze A, Postma DS, Kerstjens HA (2003) Perimenstrual asthma: a syndrome without known cause or cure. *J Allergy Clin Immunol* 112: 271–282. <https://doi.org/10.1067/mai.2003.1676>
103. Zaitseva M, Narita S, Lambert KC, et al. (2007) Estradiol activates mast cells via a non-genomic estrogen receptor-alpha and calcium influx. *Mol Immunol* 44: 1977–1985. <https://doi.org/10.1016/j.molimm.2006.09.030>
104. Howard PJ, Ambrosius WT, Tewksbury DA, et al. (1998) Serum Angiotensinogen concentration in relation to gonadal hormones, body size and genotype in growing young people. *Hypertension* 32: 875–879. <https://doi.org/10.1161/01.HYP.32.5.875>
105. Giacchetti G, Faloia E, Mariniello B, et al. (2002) Over expression of the renin angiotensin system in human visceral adipose tissue in normal and overweight subjects. *Am J Hypertension* 15: 381–388. [https://doi.org/10.1016/S0895-7061\(02\)02257-4](https://doi.org/10.1016/S0895-7061(02)02257-4)
106. Reinholz M, Ruzicka T, Schaubert J (2012) Vitamin D and its role in allergic disease. *Clin Exp Allergy* 42: 817–826. <https://doi.org/10.1111/j.1365-2222.2011.03923.x>
107. Agrawal T, Gupta GK, Agrawal DK (2013) Vitamin D supplementation reduces airway hyperresponsiveness and allergic airway inflammation in a murine model. *Clin Exp Allergy* 43: 672–683. <https://doi.org/10.1111/cea.12102>
108. Aranow C (2011) Vitamin D and the immune system. *J Investig Med* 59: 881–886. <https://doi.org/10.2310/JIM.0b013e31821b8755>
109. Hewison M (2010) Vitamin D and the immune system: new perspectives on an old theme. *Endocrinol Metab Clin North Am* 39: 365–379. <https://doi.org/10.1016/j.ecl.2010.02.010>
110. Norman AW, Mizwicki MT, Norman DP (2004) Steroid-hormone rapid actions, membrane receptors and a conformational ensemble model. *Nat Rev Drug Discov* 3: 27–41. <https://doi.org/10.1038/nrd1283>
111. Mahon BD, Wittke A, Weaver V, et al. (2003) The targets of vitamin D depend on the differentiation and activation status of CD4 positive T cells. *J Cell Biochem* 89: 922–932. <https://doi.org/10.1002/jcb.10580>
112. Fabri M, Stenger S, Shin DM, et al. (2011) Vitamin D is required for IFN-gamma-mediated antimicrobial activity of human macrophages. *Sci Transl Med* 3: 104ra102. <https://doi.org/10.1126/scitranslmed.3003045>

113. Gupta A, Dimeloe S, Richards DF, et al. (2014) Defective IL-10 expression and in vitro steroid-induced IL-17A in paediatric severe therapy-resistant asthma. *Thorax* 69: 508–515. <https://doi.org/10.1136/thoraxjnl-2013-203421>
114. Herman K, Ring J (1990) Hymenoptera venom anaphylaxis: may decrease levels of angiotensin peptides play a role? *Clin Exp All* 20: 569–570. <https://doi.org/10.1111/j.1365-2222.1990.tb03151.x>
115. Herman K, Ring J (1993) The renin angiotensin system and hymenoptera venom anaphylaxis. *Clin Exp Allergy* 23: 762–769. <https://doi.org/10.1111/j.1365-2222.1993.tb00364.x>
116. Summers CW, Pumphrey RS, Woods CN, et al. (2008) Factors predicting anaphylaxis to peanuts and tree nuts in patients referred to a specialist centre. *J Allergy Clin Immunol* 121: 632–638. <https://doi.org/10.1016/j.jaci.2007.12.003>
117. Slade CA, Douglass JA (2014) Changing practice: no need to stop ACE inhibitors for venom immunotherapy. *Clin Exp Allergy* 44: 617–619. <https://doi.org/10.1111/cea.12295>
118. Varney VA, Warner A, Ghosh A, et al. (2012) IgE mediated anaphylaxis to foods, venom, and drugs; influence of serum angiotensin converting enzyme levels and genotype. *J Allergy (Cairo)* 2012: 258145. <https://doi.org/10.1155/2012/258145>
119. Niedoszytko M, Ratajska M, Jassem E (2007) AGT(M235T), ACE(I/D, I/I, D/D) polymorphism in patients with insect venom allergy preliminary results. *Allergy* 62 (suppl 83): 111.
120. Mueller UR (1990) Clinical presentation and pathogenesis. In: Mueller UR, editor. *Insect sting allergy*, Stuttgart: Gustav Fischer, 33–65.
121. Dimitropoulou C, Chatterjee A, McCloud L (2006) Angiotensin, bradykinin and the endothelium. In: Moncada, S, Higgs, A (eds), *The Vascular Endothelium I. Handbook of Experimental Pharmacology*, Berlin, Heidelberg: Springer, 176: 255–294. https://doi.org/10.1007/3-540-32967-6_8
122. Varney VA, Nicholas A, Warner A, et al. (2019) IgE-mediated systemic anaphylaxis and its association with gene polymorphisms of ACE, angiotensinogen and chymase. *J Asthma Allergy* 12: 343–361. <https://doi.org/10.2147/JAA.S213016>
123. Nisho H, Takai S, Miyazaki M, et al. (2005) Usefulness of serum mast cells specific chymase levels for postmortem diagnosis of anaphylaxis. *Int J Legal Med* 119: 331–334. <https://doi.org/10.1007/s00414-005-0524-1>
124. Zhou X, Whitworth HS, E-Khedr M, et al. (2011) Mast cell chymase: a useful marker in anaphylaxis. *J Allergy Clin Immunol* 127: 990–997. <https://doi.org/10.1016/j.jaci.2011.01.057>
125. Gulen T, Akin C (2021) Idiopathic anaphylaxis: a perplexing diagnostic challenge for allergists. *Curr Allergy Asthma Rep* 21: 11. <https://doi.org/10.1007/s11882-021-00988-y>
126. Akin C (2014) Mast cell activation disorders. *J Allergy Clin Immunol Pract* 2: 252–257. <https://doi.org/10.1016/j.jaip.2014.03.007>
127. Metcalfe DD, Peavy RD, Gilfillan AM (2009) Mechanisms of mast cell signaling in anaphylaxis. *J Allergy Clin Immunol* 124: 639–646. <https://doi.org/10.1016/j.jaci.2009.08.035>
128. Gulen T, Hagglund H, Dahlen SE, et al. (2014) Flushing, fatigue, and recurrent anaphylaxis: a delayed diagnosis of mastocytosis. *Lancet* 383: 1608. [https://doi.org/10.1016/S0140-6736\(14\)60585-7](https://doi.org/10.1016/S0140-6736(14)60585-7)
129. Valent P, Akin C, Arock M, et al. (2012) Definitions, criteria and global classification of mast cell disorders with special reference to mast cell activation syndromes: a consensus proposal. *Int Arch Allergy Immunol* 157: 215–225. <https://doi.org/10.1159/000328760>

130. Nakamura Y, Nakano N, Ishimaru K, et al. (2016) Inhibition of IgE-mediated allergic reactions by pharmacologically targeting the circadian clock. *J Allergy Clin Immunol* 137: 1226–1235. <https://doi.org/10.1016/j.jaci.2015.08.052>
131. Kristensen T, Vestergaard H, Bindslev-Jensen C, et al. (2014) Sensitive KIT D816V mutation analysis of blood as a diagnostic test in mastocytosis. *Am J Hematol* 89: 493–498. <https://doi.org/10.1002/ajh.23672>
132. Bonadonna P, bonifacio M, Lambardero C, et al. (2015) Hymenoptera anaphylaxis and C-KIT Mutations: An unexpected association. *Curr Allergy Asthma Rep* 15: 49. <https://doi.org/10.1007/s11882-015-0550-0>
133. Sonneck K, Florian S, Mullauer L, et al. (2007) Diagnostic and sub-diagnostic accumulation of mast cells in the bone marrow of patients with anaphylaxis: Monoclonal mast cell activation syndrome. *Int Arch Allergy Immunol* 142:158–164. <https://doi.org/10.1159/000096442>
134. Valent P, Akin C, Bonadonna P, et al. (2019) Proposed diagnostic algorithm for patients with suspected mast cell activation syndrome. *J Allergy Clin Immunol Pract* 7: 1125–1133. <https://doi.org/10.1016/j.jaip.2019.01.006>
135. Gulen T, Hagglund H, Dahlen B, et al. Mastocytosis: the puzzling clinical spectrum and challenging diagnostic aspects of an enigmatic disease. *J Intern Med* 279: 211–228. <https://doi.org/10.1111/joim.12410>
136. Valent P, Akin C, Metcalfe DD (2016) Mastocytosis 2016: updated WHO classification and novel emerging treatment concepts. *Blood* 129: 1420–1427. <https://doi.org/10.1182/blood-2016-09-731893>
137. Gotlib J, Kluin-Nelemans HC, George TI, et al. (2016) Efficacy and safety of midostaurin in advanced systemic mastocytosis. *N Engl J Med* 374: 2530–2541. <https://doi.org/10.1056/NEJMoa1513098>
138. Fuchs D, Kilbertus A, Kofler K, et al. (2021) Scoring the risk of having systemic mastocytosis in adult patients with mastocytosis in the skin. *J Allergy Clin Immunol Pract* 9: 1705–1712.e4. <https://doi.org/10.1016/j.jaip.2020.12.022>
139. Kennedy VE, Perkins C, Reiter A, et al. (2023) Mast cell leukemia: clinical and molecular features and survival outcomes of patients in the ECNM Registry. *Blood Adv* 7: 1713–1724. <https://doi.org/10.1182/bloodadvances.2022008292>
140. Gulen T, Ljung C, Nilsson G, et al. (2017) Risk factor analysis of anaphylactic reactions in patients with systemic mastocytosis. *J Allergy Clin Immunol Pract* 5: 1248–1255. <https://doi.org/10.1016/j.jaip.2017.02.008>
141. Gulen T, Hagglund H, Dahlen B, et al. (2014) High prevalence of anaphylaxis in patients with systemic mastocytosis - a single-centre experience. *Clin Exp Allergy* 44: 121–129. <https://doi.org/10.1111/cea.12225>
142. Monaco A, Choi D, Uzun S, et al. (2022) Association of mast-cell-related conditions with hypermobile syndromes: a review of the literature. *Immunol Res* 70: 419–431. <https://doi.org/10.1007/s12026-022-09280-1>
143. Sumantri S, Rengganis I (2023) Immunological dysfunction and mast cell activation syndrome in long COVID. *Asia Pac Allergy* 13: 50–53. <https://doi.org/10.5415/apallergy.0000000000000022>
144. Voelker D, Pongdee T (2014) Urine mast cell mediators in the evaluation and diagnosis of mast cell activation syndrome. *Curr Allergy Asthma Rep* 24: 33–38. <https://doi.org/10.1007/s11882-024-01128-y>

145. Zhu L, Jian X, Zhou B, et al (2024) Gut microbiota facilitate chronic spontaneous urticaria. *Nat Commun* 15: 112. <https://doi.org/10.1038/s41467-023-44373-x>
146. Robey RC, Wilcock A, Bonin H, et al. (2020) Hereditary alpha-tryptasemia: UK prevalence and variability in disease expression. *J Allergy Clin Immunol Pract* 8: 3549–3556. <https://doi.org/10.1016/j.jaip.2020.05.057>
147. Chollet MB, Akin C (2021) Hereditary alpha tryptasemia is not associated with specific clinical phenotypes. *J Allergy Clin Immunol* 148: 889–894. <https://doi.org/10.1016/j.jaci.2021.06.017>
148. Butterfield JH, Ravi A, Pongdee T (2018) Mast cell mediators of significance in clinical practice in mastocytosis. *Immunol Allergy Clin North Am* 38: 397–410. <https://doi.org/10.1016/j.iac.2018.04.011>
149. Lyons JJ, Sun G, Stone KD, et al. (2014) Mendelian inheritance of elevated serum tryptase associated with atopy and connective tissue abnormalities. *J Allergy Clin Immunol* 133: 1471–1474. <https://doi.org/10.1016/j.jaci.2013.11.039>
150. Lyons JJ, Yu X, Hughes JD, et al. (2016) Elevated basal serum tryptase identifies a multisystem disorder associated with increased TPSAB1 copy number. *Nat Genet* 48: 1564–1569. <https://doi.org/10.1038/ng.3696>
151. Simons FE, Arduzzo LR, Bilo MB, et al. (2011) World allergy organization guidelines for the assessment and management of anaphylaxis. *World Allergy Organ J* 4: 13–37. <https://doi.org/10.1097/WOX.0b013e318211496c>
152. Braganza SC, Acworth JP, McKinnon DR, et al. (2006) Paediatric emergency department anaphylaxis: different patterns from adults. *Arch Dis Child* 91: 159–163. <https://doi.org/10.1136/adc.2004.069914>
153. Muraro A, Roberts G, Worm M, et al. (2014) Anaphylaxis: guidelines from the European Academy of Allergy and Clinical Immunology. *Allergy* 69: 1026–1045. <https://doi.org/10.1111/all.12437>
154. Jarkvist J, Salehi C, Akin C, et al. (2020) Venom immunotherapy in patients with clonal mast cell disorders: IgG4 correlates with protection. *Allergy* 75: 169–177. <https://doi.org/10.1111/all.13980>
155. Broesby-Olsen S, Vestergaard H, Mortz CG, et al. (2018) Omalizumab prevents anaphylaxis and improves symptoms in systemic mastocytosis: efficacy and safety observations. *Allergy* 73: 230–238. <https://doi.org/10.1111/all.13237>
156. Gotlib J, Kluin-Nelemans HC, Akin C, et al. (2021) Practical management of adverse events in patients with advanced systemic mastocytosis receiving midostaurin. *Expert Opin Biol Ther* 21: 487–498. <https://doi.org/10.1080/14712598.2021.1837109>
157. Gotlib J, Kluin-Nelemans HC, George TI, et al. (2016) Efficacy and safety of midostaurin in advanced systemic mastocytosis. *N Engl J Med* 374: 2530–2541. <https://doi.org/10.1056/NEJMoa1513098>
158. Kudlaty E, Perez M, Stein BL, et al. (2021) Systemic mastocytosis with an associated hematologic neoplasm complicated by recurrent anaphylaxis: prompt resolution of anaphylaxis with the addition of avapritinib. *J Allergy Clin Immunol Pract* 9: 2534–2536. <https://doi.org/10.1016/j.jaip.2021.02.040>

