



Research article

Impact of etiological factors on citrullination markers and susceptibility of PADI4 allele for CHIKV induced rheumatoid arthritis among South Indian Tamil RA cases

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Abstract: Rheumatoid arthritis (RA) is a multifactorial disease which can be triggered by gene-environment interactions. Numerous risk factors have been acknowledged in varied ethnicities, but their generalizability is vague. Hence, proposed to identify impact of etiology on citrullination and how both interact with peptidyl arginine deiminase 4 (PADI4) polymorphism in RA onset among South Indian Tamil RA cases. Studied 207 RA cases and 186 healthy controls for C-reactive protein (CRP), erythrocyte sedimentation rate (ESR), rheumatoid factor (RF), anti-cyclic citrullinated peptide (CCP), anti-Sa (citrullinated vimentin), anti-citrullinated α -enolase peptide-1 (CEP-1) and diseases activity score-28 (DAS-28). Past exposure to studied etiological risk factors obtained through questionnaire. Family history of RA (FHRA), surgery/injury and chikungunya virus (CHIKV) infection significantly contributed to RA ($p < 0.05$) particularly CHIKV (OR = 6.66, 95% CI 3.92–11.32, $p = 0.001$). Strikingly, 67.1% of surgery/injury and 80% of CHIKV exposed patients had RA onset within a year. RA cases with tooth decay had impact on RF, anti-CCP, anti-Sa frequency and anti-CEP-1 level ($p < 0.05$). Since CHIKV infected cases showed significant anti-Sa ($p = 0.04$) level and frequency ($p = 0.01$), they were genotyped for polymorphism in PADI4_92 (rs874881), 104 (rs1748033) and 94 (rs2240340) by Sanger's sequencing which demonstrated that PADI4 confers risk ($p < 0.05$) for the onset of CHIKV induced RA. This is the initial report that CHIKV may contribute to RA development via vimentin citrullination. FHRA, surgery/injury, CHIKV and smoking posed a key RA risk.

Keywords: anti-CCP; anti-CEP-1; anti-Sa; CHIKV; PADI4; RA

1. Introduction

Rheumatoid arthritis (RA) is a chronic systemic autoimmune inflammatory disease categorized by joint swelling and tenderness leading to destruction of cartilage and bone, resulting in severe physical disability and premature mortality [1,2]. It holds 42nd position in causing physical disability around the World with a stable rise from 3.3 to 4.8 million in a span of 20 years. Globally, the prevalence of RA is believed to range from 0.5% to 1% [3].

Although RA etiology is not fully elucidated, the multifactorial nature reflects “Bermuda triangle” of genetic, environment and autoimmunity robustly persuading the onset and persistence of pathologic condition [4]. Numerous studies have publicized that environmental factors play a substantial role in the development of RA in an appropriate genetic background [5]. FHRA, cigarette smoking, CHIKV, periodontitis [6] and physical trauma [7] has been assumed to be a promising risk factor for RA. Moreover, the recognition of a “preclinical” phase of RA via the evaluation of autoantibodies and other biomarkers proposes that the triggering agents for RA are performing long before the initial clinical indication of inflammatory arthritis.

All these highlight the instigation of new era, which will possibly unravel the complex interplay between gene and environmental factors, and also specific pathologically linked immune reactions that may be initiated in the background of diverse gene to environment combinations. Further, probing of individual risk factors and whether they control specific citrullination between the exposed and non-exposed may provide deeper understanding, which will assist in designing target-oriented drug and modelling of the risk prediction. To this end, consistent result from ethnically diverse population is required, because risk factors recognized in one ethnic population is not replicated in another.

Till date, human leukocyte antigen-DR isotype (HLA-DRB1) gene and smoking has been proved beyond doubt as a RA threat [8], however, other potential targets remain to be elucidated. Convincing case-control study in the context of RA triggering factors such as FHRA, CHIKV, physical trauma/surgery, tooth decay and diet pattern were undertaken. Strikingly, till date, the impact of these triggers on inflammatory mediated citrullination of vimentin and α -enolase is unexplored in the Indian population.

In order for an environmental agent to be established as a disease, biochemical, and immunological event followed by a trigger should act in favour of disease pathology in genetically susceptible individuals. When susceptible loci interact with prospective risk variables, the chances of pathologic signature are higher and more vulnerable. Symptom similarities between RA and CHIKV arthritis was intriguing to investigate whether the immunologic events and genetic susceptibility of RA could also be a mediator of CHIKV induced RA and also whether probable mechanism that remains obscure be elucidated by connecting all the related events.

2. Materials and methods

Blood samples were collected from South Indian Tamil RA patients visiting a Rheumatology clinic in Chennai, Tamil Nadu, India during 2014 to 2018. Patients presenting with at least 1 joint

with definite clinical synovitis or synovitis not clearly explained were subjected to further screening. Details of age, sex, symptoms and duration of disease were obtained and also examined the number of swollen and tender joints. Patients with existing diagnosis of osteoarthritis, and other rheumatic diseases were excluded. We studied 207 South Indian Tamil RA cases satisfying the revised 2010 American College of Rheumatology criteria for the classification of RA. Age and sex matched 186 South Indian Tamil healthy individuals without any other rheumatic diseases were enrolled as healthy control group. To identify the etiologic factors, prepared a well-designed questionnaire incorporating potential risk factors and details obtained from RA cases and healthy controls with their informed consent prior to enrolment into the protocol. Past exposure of CHIKV infection ascertained from their medical records.

2.1. Blood sample collection

Venous blood was collected by standard protocol into redtop vacutainer for serological parameters, blacktop (citrate) for ESR, purple top (ethylene diamine tetra acetic acid (EDTA)) for genetic analysis. Serum and whole blood was stored at -70°C .

2.2. Calculation of disease activity score (DAS)-28

DAS-28 was calculated based on the number of swollen joint count, tender joint count and ESR using online DAS-28 calculator (<https://www.rheumakit.com/en/calculators/das28>). A DAS-28 of less than 3.2 implies low disease activity, 3.2–5.1 moderate disease activity and more than 5.1 active disease.

2.3. Laboratory parameters

2.3.1. Rheumatoid factor (RF)

RF was assayed by Immunoturbidimetry. Values >14 IU/mL were considered positive. Samples above the linearity limit of 130 IU/mL were diluted to get the absolute value.

2.3.2. C-reactive protein (CRP)

CRP was performed by particle enhanced Immunoturbidimetry. CRP of <5.0 mg/L was considered negative and >5.0 mg/L as positive. Samples above the linearity limit of 250 mg/L were diluted to get the exact value.

2.3.3. Anti-cyclic citrullinated peptide (CCP)

Anti-CCP was performed by second generation kit which uses multiple citrullinated peptide by Electrochemiluminescence Immuno Assay (ECLIA). Values >17 U/mL were considered positive. All samples above the linearity limit of 500 U/mL were taken as 501 U/mL.

Bio-Rad Liquicheck Immunology quality control (RF & CRP) and Roche Preci control (anti-CCP) was run daily. IgM RF, CRP and anti-CCP were performed as per the respective pack insert of the manufacturer on automated Roche COBAS 6000 equipment.

2.3.4. Anti-Sa and anti-CEP-1 antibodies

Anti-Sa and anti-CEP-1 antibody were quantified using an enzyme-linked immunosorbent assay (ELISA) (EUROIMMUN) according to the manufacturer's instructions. Values less than 20 RU/mL were considered negative and more than 20 RU/ml positive. Results above 200 RU/mL were diluted to obtain absolute value.

2.3.5. Erythrocyte sedimentation rate (ESR)

ESR was measured by Westergren's method. Reference interval is 5–15 mm/h for male and 5–20 mm/h for female.

2.4. Statistical analysis

Nominal variables were expressed as frequencies and continuous variables distributed normally were expressed as mean \pm standard deviation. Mann–Whitney U-test were used to evaluate the differences between two groups. Odds ratio (OR) and 95% confidence intervals (CI) of cases and controls were examined using bivariate analysis and chi-square test. The Hardy–Weinberg equilibrium (HWE) for the single nucleotide polymorphisms (SNP) was calculated with the chi-square test. Genotype frequency of studied polymorphism between cases and controls were analyzed using an online SNPSTATS program (<https://www.snpstats.net/snpstats/analyser.php>). Statistical analysis was performed using Statistical Package for the Social Sciences (SPSS) version 20.0. p -value < 0.05 was considered as statistically significant.

2.5. Ethics approval of research

The study was approved by Institutional Human Ethics Committee (IHEC), HDC/IHEC/002, Hitech Diagnostic Centre, Chennai.

3. Results

3.1. Assessment of serological and etiological factors in RA cases and healthy controls of South Indian Tamil population

Table 1 summarizes serological and etiological characteristic of 207 South Indian Tamil RA cases and 186 South Indian Tamil healthy controls. Two groups restrain similar ratio of gender, cases enclosed 178 (86.0%) females and 29 (14.0%) males whereas controls constituted 161 (86.6%) females and 25 (13.4%) males. At the time of enrolment, RA had travelled 6.7 ± 4.3 years with moderate disability index (4.5 ± 0.6). Frequency of analysed antibody positivity among RA cases were RF (88.9%), anti-CCP (85.5%), anti-Sa (61.8%) and anti-CEP-1 (68.6%) with mean value of

RF (162.7 ± 235.8), anti-CCP (293.8 ± 195.3), anti-Sa (94.9 ± 129.4), anti-CEP-1 (77.8 ± 82.4), CRP (19.7 ± 33.9) and ESR (54.7 ± 28.6). In controls, studied serological and inflammatory markers were within the respective reference interval.

Table 1. Assessment of serological and etiological factors in RA cases and healthy controls of South Indian Tamil population.

	RA cases (n = 207)	Controls (n = 186)	OR (95% CI)	p value
Age (mean \pm SD)	49.1 \pm 11.8	48.6 \pm 12.5		
Female, n (%)	178 (86.0)	161 (86.6)		
Male, n (%)	29 (14.0)	25 (13.4)		
RA duration (mean \pm SD)	6.7 \pm 4.3	-		
HAQ (mean \pm SD)	1.4 \pm 0.4	-		
DAS-28 (mean \pm SD)	4.5 \pm 0.6	-		
RF positive, n(%)	184 (88.9)	-		
RF, IU/mL (mean \pm SD)	162.7 \pm 235.8	8.1 \pm 3.7		
Anti-CCP positive, n(%)	177 (85.5)	-		
Anti-CCP, U/mL (mean \pm SD)	293.8 \pm 195.3	7.2 \pm 1.4		
Anti-Sa positive, n(%)	128 (61.8)	-		
Anti-Sa, RU/mL (mean \pm SD)	94.9 \pm 129.4	3.3 \pm 2.8		
Anti-CEP-1 positive, n(%)	142 (68.6)	-		
Anti-CEP-1, RU/mL (mean \pm SD)	77.8 \pm 82.4	7.7 \pm 2.8		
CRP, mg/L (mean \pm SD)	19.7 \pm 33.9	3.1 \pm 2.4		
ESR, mm (mean \pm SD)	54.7 \pm 28.6	21.2 \pm 11.6		
Family history of RA	No, n(%)	153 (73.9)	180 (96.8)	1.00
	Yes, n(%)	54 (26.1)	6 (3.2)	10.58 (4.43–25.28)
	First degree relatives, n(%)	39 (72.2)	-	
	Non-first degree relatives, n(%)	15 (27.8)	-	
Surgery/injury	No, n(%)	122 (58.9)	146 (78.5)	1.00
	Yes, n(%)	85 (41.1)	40 (21.5)	2.54 (1.62–3.97)
	Cesarean	53 (62.4)		
	Other surgery/accident/fractures	32 (37.6)		
	RA onset within a year of surgery/injury, n(%)	57 (67.1)		
Chikungunya	No, n(%)	112 (54.1)	165 (88.7)	1.00
	Yes, n(%)	95 (45.9)	21 (11.3)	6.66 (3.92–11.32)
	RA onset within a year of Chikungunya, n(%)	76 (80.0%)		
Smoking	Non-smokers, n(%)	13 (44.8)	20 (80)	1.00
	Smokers, n (%)	16 (55.2)	5 (20)	4.92 (1.44–16.72)
Tooth decay	No, n(%)	64 (30.9)	67 (36.0)	1.00
	Yes, n(%)	143 (69.1)	119 (64.0)	1.25 (0.82–1.91)
Diet pattern	Non-vegetarian, n (%)	174 (84.1)	144 (77.4)	1.00
	Vegetarian, n (%)	33 (15.9)	42 (22.6)	0.65 (0.39–1.07)

RA = rheumatoid arthritis; RF = rheumatoid factor; Anti-CCP = anti-cyclic citrullinated peptide; Anti-Sa = anti-citrullinated vimentin; Anti-CEP-1 = anti-citrullinated α enolase peptide 1; ESR = erythrocyte sedimentation rate; CRP = C reactive protein; DAS-28 = disease activity score-28; OR = odds ratio; CI = confidence interval. Results are presented as mean \pm SD. p value < 0.05 was considered statistically significant.

Genetic and environment factors that are promising trigger for RA initiation were analysed. Among 26.1% of RA cases FHRA was associated with high RA risk ($p = 0.001$) to controls. 72.2% of RA cases disclosed RA affected first degree relatives (FDRs) and 27.8% non-FDRs.

When analysing RA triggers, 41.1% cases who reported surgery/injury were significantly linked with risk ($p = 0.001$) to 21.5% controls. Among surgery/injury experienced RA cases 62.4% had underwent cesarean. Overall 67.1% had RA onset within a year of exposure to surgery/injury especially cesarean delivery (62.4%) to be a potential etiologic threat for RA.

In addition, 45.9% RA cases affected with CHIKV had marked association with RA risk compared with 11.3% controls affected with CHIKV ($p = 0.001$). Shockingly, among 95 CHIKV exposed RA cases, 76 (80%) of cases have developed RA within a year of infection.

Smoking vigorously influenced RA risk among 55.2% of male cases to controls ($p = 0.010$). Both tooth decay history and diet pattern did not demonstrate RA risk among the South Indian Tamil RA cases.

3.2. Comparison of serological parameters between with and without surgery in Tamil RA cases

Analysis performed to recognize whether assessed risk factors have worsening effect on pathological process of citrullination and inflammation in Tamil RA cases. As summarized in Table 2, cases with surgery/injury did not document marked variation in concentration and frequency of studied serological and inflammatory markers ($p > 0.05$).

Table 2. Comparison of serological parameters between with and without surgery in South Indian Tamil RA cases.

		Surgery/injury (n = 85)	Non-surgery/injury (n = 122)	p value
Autoantibody level	RF, IU/mL (mean \pm SD)	163.5 \pm 217.1	162.2 \pm 248.8	0.648
	Anti-CCP, U/mL (mean \pm SD)	284.3 \pm 194.3	300.3 \pm 196.5	0.509
	Anti-Sa, RU/mL (mean \pm SD)	87.8 \pm 124.1	99.9 \pm 133.2	0.780
	Anti-CEP-1, RU/mL (mean \pm SD)	70.5 \pm 72.2	82.8 \pm 88.8	0.399
Inflammatory markers	ESR, mm (mean \pm SD)	53.4 \pm 26.5	55.7 \pm 30.1	0.695
	CRP, mg/L (mean \pm SD)	21.4 \pm 44.8	18.5 \pm 23.7	0.847
Disability index	DAS-28, (mean \pm SD)	4.5 \pm 0.6	4.5 \pm 0.6	0.901
Autoantibody positive	RF positive, n (%)	77 (90.6)	107 (87.7)	0.517
	Anti-CCP positive, n (%)	70 (82.4)	107 (87.7)	0.284
	Anti-Sa positive, n (%)	51 (60)	77 (63.1)	0.650
	Anti-CEP-1 positive, n (%)	58 (68.2)	84 (68.9)	0.925

Results are presented as mean \pm SD. SD = Standard deviation; p value < 0.05 was considered statistically significant.

3.3. Comparison of serological parameters between with and without CHIKV in South Indian Tamil RA cases

Table 3 depicts 95 CHIKV infected RA cases had remarkably higher level of anti-Sa ($p = 0.048$) and ESR ($p = 0.018$) compared with 112 non-CHIKV infected RA cases and no disparity in the level of anti-CCP, anti-CEP-1, CRP and DAS-28 ($p > 0.05$).

Table 3. Comparison of serological parameters between with and without chikungunya in South Indian Tamil RA cases.

		Chikungunya (n = 95)	Non-chikungunya (n = 112)	p value
Autoantibody level	RF, IU/mL (mean ± SD)	160.1 ± 216.3	165.1 ± 252.1	0.400
	Anti-CCP, U/mL (mean ± SD)	297.2 ± 186.5	290.9 ± 203.3	0.920
	Anti-Sa, RU/mL (mean ± SD)	101.1 ± 111.5	89.8 ± 143.1	0.048
	Anti-CEP-1, RU/mL (mean ± SD)	82.1 ± 77.1	74.1 ± 87.1	0.336
Inflammatory markers	ESR, mm (mean ± SD)	60.1 ± 30.9	50.2 ± 25.8	0.018
	CRP, mg/L (mean ± SD)	22.6 ± 40.3	17.2 ± 27.4	0.089
Disability index	DAS28 (mean ± SD)	4.5 ± 0.6	4.4 ± 0.6	0.268
Autoantibody positive	RF positive, n (%)	87 (91.6)	97 (86.6)	0.260
	Anti-CCP positive, n (%)	84 (88.4)	93 (83.0)	0.275
	Anti-Sa positive, n (%)	67 (70.5)	61 (54.5)	0.018
	Anti-CEP-1 positive, n (%)	70 (73.7)	72 (64.3)	0.147

RA = rheumatoid arthritis; RF = rheumatoid factor; Anti-CCP = anti-cyclic citrullinated peptide; Anti-Sa = anti-citrullinated vimentin; Anti-CEP-1 = anti-citrullinated α enolase peptide 1; ESR = erythrocyte sedimentation rate; CRP = C reactive protein; DAS-28 = disease activity score-28. Results are presented as mean ± SD. SD = standard deviation; p value < 0.05 was considered statistically significant.

Probably, CHIKV persuaded RA through vimentin citrullination which is evidenced by increased anti-Sa frequency also ($p = 0.018$). The other antibodies failed to show any key variation ($p > 0.05$).

3.4. Comparison of serological parameters between with and without tooth decay history in South Indian Tamil RA cases

Table 4 illustrates 143 cases reported with preceding history of tooth decay had significantly elevated level of anti-CEP-1 ($p = 0.015$) and inflammatory markers ($p < 0.05$) compared to 64 cases with no history of tooth decay. While RF, anti-CCP and anti-Sa level were insignificant ($p > 0.05$), Tamil RA cases with tooth decay showed a remarkably higher frequency of RF ($p = 0.023$), anti-CCP ($p = 0.047$), anti-Sa ($p = 0.008$) and anti-CEP-1 ($p = 0.026$) to those without tooth decay.

Table 4. Comparison of serological parameters between with and without tooth decay in South Indian Tamil RA cases.

		Tooth decay (n = 143)	Without tooth decay (n = 64)	p value
Autoantibody level	RF, IU/mL (mean ± SD)	169.2 ± 247.2	148.3 ± 209.1	0.187
	Anti-CCP, U/mL (mean ± SD)	302.9 ± 187.9	273.3 ± 211.1	0.350
	Anti-Sa, RU/mL (mean ± SD)	92.1 ± 107.6	101.5 ± 169.1	0.161
	Anti-CEP-1, RU/mL (mean ± SD)	86.8 ± 86.8	57.6 ± 68.1	0.015
Inflammatory markers	ESR, mm (mean ± SD)	58.1 ± 30.3	47.2 ± 22.8	0.026
	CRP, mg/L (mean ± SD)	22.6 ± 39.5	13.1 ± 13.7	0.023
Disability index	DAS28 (mean ± SD)	4.6 ± 0.6	4.3 ± 0.6	0.041
Autoantibody positive	RF positive, n (%)	132 (92.3)	52 (81.2)	0.023
	Anti-CCP positive, n (%)	127 (88.8)	50 (78.1)	0.047
	Anti-Sa positive, n (%)	97 (67.8)	31 (48.4)	0.008
	Anti-CEP-1 positive, n (%)	105 (73.4)	37 (57.8)	0.026

Results are presented as mean ± SD. SD = Standard deviation; p value < 0.05 was considered statistically significant.

Smoking did not influence the serology and inflammatory markers in Tamil RA cases as in Table 5.

Table 5. Comparison of serological parameters between with and without smoking in South Indian Tamil RA cases.

		Smokers (n = 16)	Non-smokers (n = 13)	p value
Autoantibody level	RF, IU/mL (mean ± SD)	439.6 ± 480.5	180.2 ± 268.1	0.072
	Anti-CCP, U/mL (mean ± SD)	318.3 ± 171.8	246.1 ± 189.3	0.253
	Anti-Sa, RU/mL (mean ± SD)	102.6 ± 78.1	134.2 ± 102.5	0.354
	Anti-CEP-1, RU/mL (mean ± SD)	128.3 ± 104.2	106.1 ± 121.7	0.401
Inflammatory markers	ESR, mm (mean ± SD)	66.4 ± 36.8	57.3 ± 33.6	0.562
	CRP, mg/L (mean ± SD)	34.6 ± 33.3	25.1 ± 30.9	0.536
	DAS28 (mean ± SD)	4.8 ± 0.6	4.5 ± 0.5	0.143
Autoantibody positive	RF positive, n (%)	14 (87.5)	12 (92.3)	0.675
	Anti-CCP positive, n (%)	15 (93.8)	11 (84.6)	0.435
	Anti-Sa positive, n (%)	14 (87.5)	11 (84.6)	0.823
	Anti-CEP-1 positive, n (%)	12 (75)	10 (76.9)	0.904

Results are presented as mean ± SD. SD = standard deviation. p value < 0.05 was considered statistically significant.

3.5. PADI4 genotype and allelic frequencies in CHIKV infected RA cases and healthy controls

As represented in Table 6, CHIKV to PADI4 interaction demonstrated CHIKV infected RA cases have significantly higher proportion of susceptible C allele compared to CHIKV infected controls (PADI4_92; p = 0.04, PADI4_104; p = 0.02, PADI4_94; p = 0.02) highlighting “C” as the risk allele playing a central role in CHIKV induced RA.

Table 6. Comparison of genotype and allelic frequencies of PADI4 SNPs (PADI4_92, PADI4_104 and PADI4_94) in CHIKV infected RA cases and healthy controls.

SNPs			RA cases, n = 95 (%)	Controls, n = 21 (%)	OR (95% CI)	p value
PADI4_92, rs874881 (C > G)	Genotype	GG	48 (50.5%)	15 (71.6%)		
		CG	22 (23.2%)	3 (14.2%)		
		CC	25 (26.3%)	3 (14.2%)	2.44 (0.87–6.84)	0.08
	Allele	G	118 (62.1%)	33 (78.6%)		0.04
		C	72 (37.9%)	9 (21.4%)	2.23 (1.01–4.94)	
PADI4_104, rs1748033 (C > T)	Genotype	TT	23 (24.2%)	9 (42.9%)		
		CT	38 (40%)	9 (42.9%)		
		CC	34 (35.8%)	3 (14.2%)	2.34 (0.87–6.27)	0.08
	Allele	T	84 (44.2%)	27 (64.3%)		0.02
		C	106 (55.8%)	15 (35.7%)	2.27 (1.13–4.54)	
PADI4_94, rs2240340 (C > T)	Genotype	TT	29 (30.5%)	8 (38.2%)		
		CT	35 (36.8%)	10 (47.6%)		
		CC	31 (32.7 %)	3 (14.2%)	0.71 (0.26–1.90)	0.50
	Allele	T	93 (48.9%)	26 (61.9%)		0.02
		C	97 (51.1%)	16 (38.1%)	1.69 (0.85–3.36)	

RA = rheumatoid arthritis; PADI4 = peptidylarginine deiminases-4; OR = odds ratio; CI = confidence interval. $P < 0.05$ was considered statistically significant.

Considering genotype frequency, PADI4_92 (OR = 2.44, 95% CI 0.87–6.84; $p = 0.08$) and PADI4_104 (OR = 2.34, 95% CI 0.87–6.27; $p = 0.08$) had trend towards risk for RA as suggested by odds ratio. Nevertheless, genotype of PADI4_94 had no remarkable variation ($p = 0.50$).

4. Discussion

Genomic and pharmacological research in RA clinical areas have shown attractive difference between ethnic groups in disease phenotype, gene to environment interaction and drug efficiency, underscoring the need of specific ethnic studies to gain insights into the linkage between risk factors and RA phenotype. Therefore, the present investigation was undertaken to identify potential risk factors that may drive citrullination mediated antibody generation and assess their interaction with PADI4 loci in South Indian Tamil RA population.

DAS-28 and HAQ were moderately presented in this population, describing both are intertwining in RA pathogenesis. Equivalent pattern documented in previous findings [9,10]. Current South Indian Tamil RA population travelled 6.7 years with RA is well correlated with other studies [11]. Frequency of RF, anti-CCP, anti-Sa and anti-CEP-1 were 88.9%, 85.5%, 61.8% and 68.6% respectively. This outcome is supported by previous publications which documented the incidence of RF [12], anti-CCP [12,13], anti-Sa [14] and anti-CEP-1 [12,15,16].

RA risk increases 1.5 to 3 fold for children of RA women during their lifetime [17], emphasizing the inevitable role of heredity in RA etiology. Present work displayed 10 fold higher risk of RA in patients with FHRA (26.1%). Of them, FDRs constituted 72.2% and non-FDRs comprised 27.8% are almost similar proportion to North Kerala study [6]. FHRA increases the risk of RA, especially those with FDRs had severe diseases activity [18].

It has been acknowledged that noticeable signs of inflammation materialized in synovial joints after few months of physical trauma/injury/fracture [19,20], nonetheless, the commencement of synovial inflammation after surgery in normal healthy individuals is still uncertain. In this study, preceding history of surgery/injury (41.1%) was statistically prominent to controls. Alarming, of them, 67.1% registered RA onset within one year of the incidence. The possible mechanism include ischemic reperfusion, either localized or systemic hypoxia from tissue injury [21], release and accumulation of mitochondrial constituents and nuclear antigens from dying tissue [22], and the collision of cartilaginous particles into synovial tissue, which then provokes the generation of inflammatory cytokines and proteinases by synovial lining [23]. Strikingly, 62.4% surgery reported RA cases had experienced caesarean surgery, strongly emphasize cesarean was independently connected to RA onset in this population. This could be partly explained by the fact that inflammatory process involved in tissue repair after cesarean surgery at some point, activates citrullination of endogenous vimentin that has high affinity to shared epitope (SE) of HLA-DRB1, succeeded by T-cells recognition [24,25] with subsequent antibody generation.

CHIKV shares similar clinical characteristics with RA [26] in particular, both have highly identical peripheral T cell phenotypes. [27]. Earlier reports documented persistent rheumatic manifestations with musculoskeletal tissue devastation in CHIKV infected patients [28,29] and convalescent patients are more likely to experience chronic joint disease because of the immune response to CHIKV [30]. In this study, 45.9% of RA cases reported CHIKV history was remarkably higher than controls, closely resembling previous observations [6,26]. Strikingly, 80% of them had rapid onset within a year is in line with earlier studies [31]. Possibly, post CHIKV polyarthritis would develop if they failed to eliminate viral RNA, and persistent immune response with increased generation of proinflammatory mediators participate in the proliferation of osteoblast and osteoclast cells [32], generating RA favored autoimmune response in genetically predisposed individuals.

Meta-analysis has claimed 40% higher likelihood of RA among ever smokers than never smokers [33]. The present study investigated its impact on male cases only because none of the female participants were in the habit of smoking, the percentage of RA smokers 55.2% was remarkably higher to controls, suggesting smoking has been implicated in the initiation and propagation of RA. The major pathologic route of smoking persuaded RA is reported to be citrullination in lungs and interaction with HLA-DRB1 genes [34].

Porphyromonas gingivalis (*P.gingivalis*) causes periodontal disease, affecting connective tissues that support the survival of teeth and gradually leading to tooth loss/tooth decay. Despite 69.1% of Tamil RA cases had tooth decay/tooth loss, significant association not met for RA risk, but OR of 1.25 denotes it might have indirectly involved in etiology via creating pro-pathologic milieu for RA development. This result is in close agreement with earlier report [35]. Contrasting result observed in Korea National Health and Nutrition Examination Survey [36].

Diet pattern showed no influence on RA cases in the current study. While Mediterranean diet score and healthy diet predominantly of vegetarian source were reported to be inversely related to RA risk in large sample size of Epidemiological Investigation of Rheumatoid Arthritis (EIRA) and Nurses' Health study (NHS) respectively [37,38]. The possible protective nature of vegetarian diet could be drawn from experimental model that described suppression of osteoclastogenesis by downregulating tumour necrosis factor (TNF)- α , nitric oxide synthase and interleukin (IL)-6 [39].

It is essential to look at the connection between the individual risk factors and how well they interact with inflammatory mediated autoimmunity or whether they have impact on antibody level

and specific citrullination between the exposed and non-exposed RA cases. Serological markers of RA cases who underwent surgery were almost similar to those who did not undergo surgery, revealing other factors may be strongly implicated in disease pathogenesis. No statistical significance noticed in smokers relative to non-smokers in this ethnicity probably due to lower number of male cases. An identical pattern documented in previously published South Korean [40] and Swedish population [41].

Although tooth decay did not contribute to RA risk in south Indian Tamil ethnicity an enormous inflammatory and citrullination (vimentin and α -enolase) reaction observed in cases with tooth decay history compared to those without tooth decay among RA cases. Previous studies documented remarkable variation in the concentration of anti-Sa but not anti-CCP and anti-CEP-1 between RA with and without periodontitis subgroups of European descent [42]. The possible explanation of marked inflammation and citrullination observed in cases with tooth decay history could be drawn from periodontitis provoked RA, because hypercitrullination in periodontitis and RA are identical [43]. The probable pathological episode was collectively described as “two-hit” model, first, breaching of tolerance against specific citrullinated proteins produced by *P. gingivalis* at gingival inflammation, following epitope spreading to other host citrullinated proteins/peptides at inflamed synovium, would eventually lead to typical chronic deteriorating inflammation that symbolize RA [44], secondly, bacterial α -enolase could also citrullinate at internal arginines through human PADI enzymes present at inflammatory site is another possible way for triggering autoantibodies [16]. Entire stretch of human enolase-1 and *P. gingivalis* enolase has been reported to share 51.4% homology with 82% sequence similarity at CEP-1 region, with corresponding similarity in antibody response too. Interestingly, cross reaction of human anti-CEP-1 with *P. gingivalis* enolase has been illustrated in vitro model portraying molecular mimicry [16] and the cumulative effect of PPAD from *P. gingivalis* and other etiopathogenic and genetic interaction could be a rationale for increased anti-CEP-1 levels in RA cases with tooth decay history.

In the current examination 45.9% of Tamil ethnic RA cases reported previous CHIKV history had significantly high magnitude to progress into RA, shockingly, RA began within one year among 80% of them. Further, stratification of citrullination markers of CHIKV infected RA cases illustrated significantly higher frequency and level of anti-Sa in CHIKV exposed RA cases. This speculation may not be replicated in all races infected with CHIKV, because genetic background can modify the inflammatory signalling pathways within same ethnic population. Indeed, citrullination is a cornerstone of RA and PADI4 genetic variants are consistently proved to be a prominent RA risk allele among Asians, instigated to investigate whether PADI4 polymorphism could be a trigger of CHIKV induced RA.

Genotype and allelic frequency analysis performed between CHIKV infected RA cases and CHIKV infected healthy Tamil non RA controls, revealed “C” allele of all the three studied PADI4_92 (rs874881), PADI4_104 (rs1748033) and PADI4_94 (rs2240340) conferred susceptibility, highlighting the central role played by PADI4 genetic polymorphism in CHIKV induced RA cases.

It has been reported that intact structure of vimentin is indispensable for initial entry of CHIKV into targeted host cells and these filaments begin to lose its original structures at the time of viral replication and eventually lead to retraction from cell membrane. Moreover, vimentin connection to CHIKV replication by two-dimensional gel electrophoresis, evidences differential regulation of host proteins principally vimentin, a cytoskeletal protein facilitating a supporting network for viral

replication [45]. Both innate and adaptive immune response arise with subsequent large cytokine production after CHIKV infection [46] promoting osteoclast activity and osteoclastogenesis [47].

It can be theorized sequential event right from CD4+ T cells apoptosis in an early phase of onset [48] followed by increased activation of immune system [49] that constantly secretes pro-inflammatory molecules at CHIKV replication site [32], amid calcium influx from dying macrophages in synovium enhances the intracellular vimentin citrullination catalyzed by PADI4 enzymes [50]. Non clearance of citrullinated proteins by the immune system results in loss of tolerance presenting them to the antigenic site of major histocompatibility complex (MHC) II alleles. The aforementioned biological event is further triggered in individuals possessing PADI4_92, 94 and 104 risk loci, ultimately generating anti-Sa and anti-citrullinated protein antibodies (ACPAs) and synovial inflammation. Probable proposed mechanism for CHIKV induced RA via vimentin citrullination and PADI-4 polymorphism in Tamil RA cases is depicted in Figure 1.

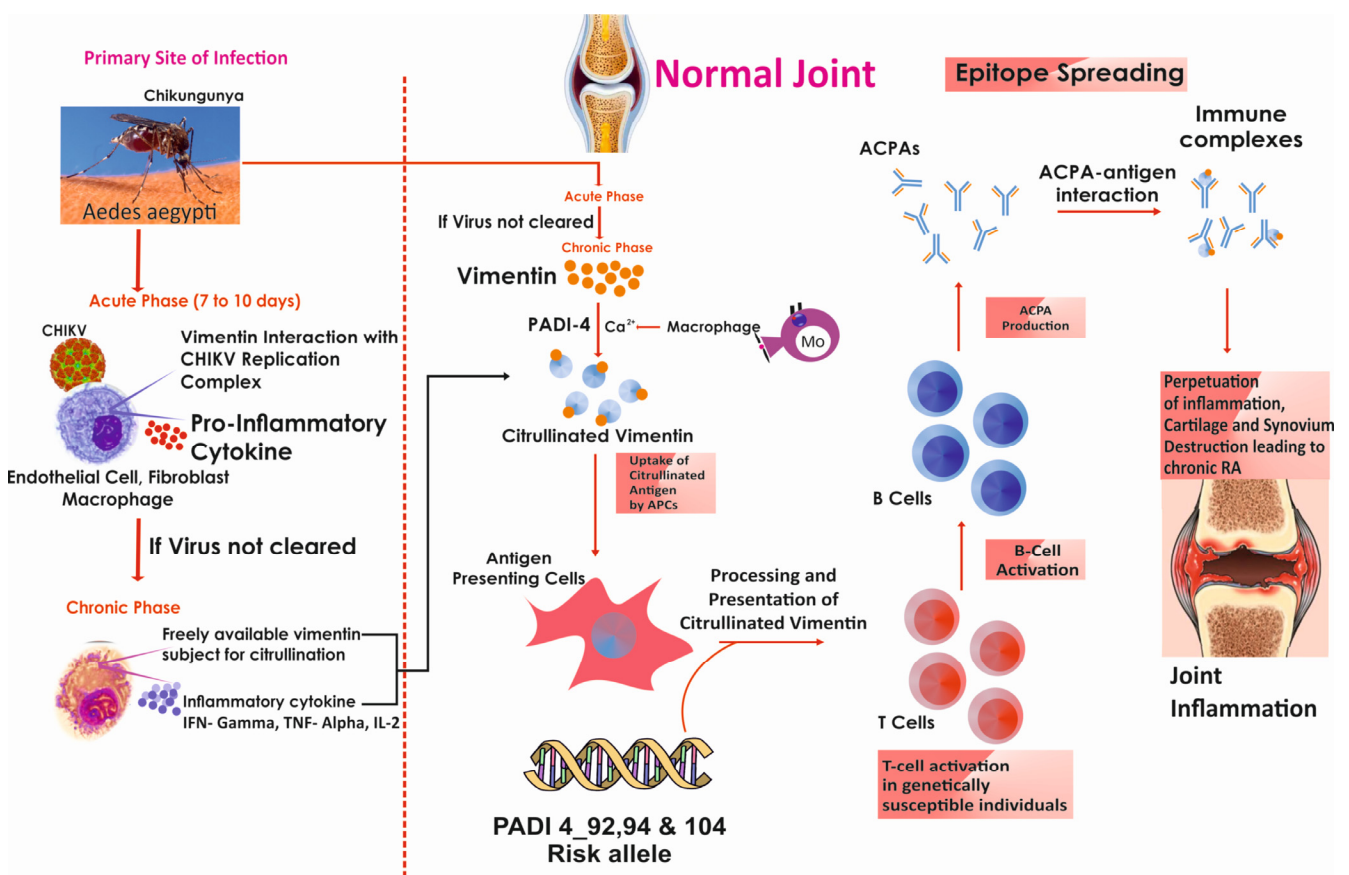


Figure 1. Schematic illustration of probable proposed mechanism for CHIKV induced RA via vimentin citrullination and PADI-4 polymorphism in South Indian Tamil RA cases. Vimentin, an intermediate filament protein is indispensable for CHIKV entry and replication. Replication takes place into the various sites of host cells and synovium simultaneously. Non clearance of virus within 7–10 days reaches a chronic phase. Due to persistent CHIKV replication, freely available disintegrated vimentin is subjected to citrullination by calcium influx from dying macrophages in synovium in the presence of PADI4. If citrullinated vimentin is not cleared by the immune system and results in loss of tolerance and increased activation of the T cells that present citrullinated vimentin at

the antigenic site of the MHC II alleles leads to epitope spreading. Subsequently, B cells produce antibodies specific to citrullinated vimentin and the formation of immune complexes. The aforementioned biological event is further triggered in individuals possessing PADI4_92, 94 and 104 risk loci, ultimately leading to perpetuation of the inflammatory process eventually causing chronic RA.

Collectively, indispensability of vimentin for CHIKV entry and replication into various sites of host cells, intracellular and extracellular vimentin presence within the synovium together with polymorphism in PADI4 transcribed stable mRNA that prolongs the PADI4 enzyme activity with sustained production of citrulline from vimentin. Further CHIKV persistence might have enhanced the adaptive immune system that mistakenly attack the endogenous citrullinated self antigens particularly vimentin and inflammatory cascade in the synovium, strongly postulating vimentin citrullination might have played a central role in post-CHIKV RA onset in PADI4 predisposed individuals.

5. Conclusions

In conclusion, FHRA, smoking, CHIKV, and surgery were truly linked to RA at high magnitude is the first report in South Indian Tamil RA population. This is the first evidence that CHIKV may contribute to RA development via citrullination of vimentin in PADI4_92, 94 and 104 predisposed individuals. PADI4 poses risk loci for the initiation and exacerbation of CHIKV induced RA in Tamil population. Overall, yet again it highlights the significance of recording thorough history encompassing all the likely threats and confounding factors of RA at first visit to the clinic. Optimistically, this study would contribute towards unravelling complex etiopathogenesis and also design tailored comprehensive therapeutic interventions through companion diagnostics to alleviate disease severity and improve quality of life of patients afflicted with RA.

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Conflict of interest

All authors declare no conflicts of interest in this paper.

References

1. McInnes IB, Schett G (2011) The pathogenesis of rheumatoid arthritis. *N Engl J Med* 365: 2205–2219. <https://doi.org/10.1056/NEJMra1004965>

2. Tobón GJ, Youinou P, Saraux A (2010) The environment, geo-epidemiology, and autoimmune disease: Rheumatoid arthritis. *J Autoimmun* 35: 10–14. <https://doi.org/10.1016/j.jaut.2009.12.009>
3. Cross M, Smith E, Hoy D, et al. (2014) The global burden of rheumatoid arthritis: estimates from the global burden of disease 2010 study. *Ann Rheum Dis* 73: 1316–1322. <https://doi.org/10.1136/annrheumdis-2013-204627>
4. van der Woude D, Alemayehu WG, Verduijn W, et al. (2010) Gene-environment interaction influences the reactivity of autoantibodies to citrullinated antigens in rheumatoid arthritis. *Nat Genet* 42: 814–816. <https://doi.org/10.1038/ng1010-814>
5. Deane KD, Demoruelle MK, Kelmenson LB, et al. (2017) Genetic and environmental risk factors for rheumatoid arthritis. *Best Pract Res Clin Rheumatol* 31: 3–18. <https://doi.org/10.1016/j.berh.2017.08.003>
6. Paul B, Pariyapurath R (2018) Risk factor assessment of rheumatoid arthritis in North Kerala. *Eur J Rheumatol* 5: 184–190. <https://doi.org/10.5152/eurjrheum.2018.17111>
7. Brawer AE, Goel N (2016) The onset of rheumatoid arthritis following trauma. *Open Access Rheumatol Res Rev* 8: 77–80. <https://doi.org/10.2147/OARRR.S110560>
8. Hedström AK, Rönnelid J, Klareskog L, et al. (2019) Complex relationships of smoking, HLA-DRB1 genes, and serologic profiles in patients with early rheumatoid arthritis: update from a Swedish population-based case-control study. *Arthritis Rheumatol* 71: 1504–1511. <https://doi.org/10.1002/art.40852>
9. Sunar I, Garip Y, Yilmaz Ö, et al. (2015) Disease activity (rheumatoid arthritis disease activity index-5) in patients with rheumatoid arthritis and its association with quality of life, pain, fatigue, and functional and psychological status. *Arch Rheumatol* 30: 144–149. <https://doi.org/10.5606/ArchRheumatol.2015.5122>
10. Corbacho MI, Dapuetto JJ (2010) Assessing the functional status and quality of life of patients with rheumatoid arthritis. *Rev Bras Reumatol* 50: 31–43. <https://doi.org/10.1590/S0482-50042010000100004>
11. Ghosh SK, Bandyopadhyay D, Biswas SK, et al. (2017) Mucocutaneous manifestations in patients with rheumatoid arthritis: A cross-sectional study from Eastern India. *Indian J Dermatol* 62: 411–417. https://doi.org/10.4103/ijd.IJD_260_17
12. Meyer PW, Ally MT, Hodgkinson B, et al. (2018) Comparison of the diagnostic potential of three anti-citrullinated protein antibodies as adjuncts to rheumatoid factor and CCP in a cohort of South African rheumatoid arthritis patients. *Rheumatol Int* 38: 993–1001. <https://doi.org/10.1007/s00296-018-4036-y>
13. Montes A, Dieguez-Gonzalez R, Perez-Pampin E, et al. (2011) Particular association of clinical and genetic features with autoimmunity to citrullinated α -enolase in rheumatoid arthritis. *Arthritis Rheum* 63: 654–661. <https://doi.org/10.1002/art.30186>
14. Prasanth G, Padmaraj SR, Mathew R, et al. (2013) Anti-Sa antibody, cyclic citrullinated peptide antibody and rheumatoid factor as diagnostic markers of rheumatoid arthritis. *Age* 212: 249–264.
15. Mahdi H, Fisher BA, Källberg H, et al. (2009) Specific interaction between genotype, smoking and autoimmunity to citrullinated α -enolase in the etiology of rheumatoid arthritis. *Nat Genet* 41: 1319–1324. <https://doi.org/10.1038/ng.480>

16. Lundberg K, Kinloch A, Fisher BA, et al. (2008) Antibodies to citrullinated α -enolase peptide 1 are specific for rheumatoid arthritis and cross-react with bacterial enolase. *Arthritis Rheum* 58: 3009–3019. <https://doi.org/10.1002/art.23936>
17. Clowse ME, Chakravarty E, Costenbader KH, et al. (2012) Effects of infertility, pregnancy loss, and patient concerns on family size of women with rheumatoid arthritis and systemic lupus erythematosus. *Arthritis Care Res* 64: 668–674. <https://doi.org/10.1002/acr.21593>
18. Ramos-Remus C, Castillo-Ortiz JD, Aguilar-Lozano L, et al. (2015) Autoantibodies in prediction of the development of rheumatoid arthritis among healthy relatives of patients with the disease. *Arthritis Rheumatol* 67: 2837–2844. <https://doi.org/10.1002/art.39297>
19. Furman BD, Kimmerling KA, Zura RD, et al. (2015) Brief report: articular ankle fracture results in increased synovitis, synovial macrophage infiltration, and synovial fluid concentrations of inflammatory cytokines and chemokines. *Arthritis Rheumatol* 67: 1234–1239. <https://doi.org/10.1002/art.39064>
20. Struglics A, Larsson S, Kumahashi N, et al. (2015) Changes in cytokines and aggrecan ARGS neoepitope in synovial fluid and serum and in C-terminal crosslinking telopeptide of type II collagen and N-terminal crosslinking telopeptide of type I collagen in urine over five years after anterior cruciate ligament rupture: an exploratory analysis in the knee anterior cruciate ligament, nonsurgical versus surgical treatment trial. *Arthritis Rheumatol* 67: 1816–1825. <https://doi.org/10.1002/art.39146>
21. Eltzschig HK, Carmeliet P (2011) Hypoxia and inflammation. *N Engl J Med* 364: 656–665. <https://doi.org/10.1056/NEJMra0910283>
22. Manfredi AA, Rovere-Querini P (2010) The mitochondrion—a Trojan horse that kicks off inflammation. *N Engl J Med* 362: 2132–2134. <https://doi.org/10.1056/NEJMcibr1003521>
23. Swärd P, Frobell R, Englund M, et al. (2012) Cartilage and bone markers and inflammatory cytokines are increased in synovial fluid in the acute phase of knee injury (hemarthrosis)—a cross-sectional analysis. *Osteoarthr Cartilage* 20: 1302–1308. <https://doi.org/10.1016/j.joca.2012.07.021>
24. Snir O, Rieck M, Gebe JA, et al. (2011) Identification and functional characterization of T cells reactive to citrullinated vimentin in HLA-DRB1* 0401-positive humanized mice and rheumatoid arthritis patients. *Arthritis Rheum* 63: 2873–2883. <https://doi.org/10.1002/art.30445>
25. Hill JA, Southwood S, Sette A, et al. (2003) Cutting edge: the conversion of arginine to citrulline allows for a high-affinity peptide interaction with the rheumatoid arthritis-associated HLA-DRB1* 0401 MHC class II molecule. *J Immunol* 171: 538–541. <https://doi.org/10.4049/jimmunol.171.2.538>
26. Manimunda SP, Vijayachari P, Uppoor R, et al. (2010) Clinical progression of chikungunya fever during acute and chronic arthritic stages and the changes in joint morphology as revealed by imaging. *T Roy Soc Trop Med H* 104: 392–399. <https://doi.org/10.1016/j.trstmh.2010.01.011>
27. Miner JJ, Aw Yeang HX, Fox JM, et al. (2015) Brief report: chikungunya viral arthritis in the United States: a mimic of seronegative rheumatoid arthritis. *Arthritis Rheumatol* 67: 1214–1220. <https://doi.org/10.1002/art.39027>
28. Schilte C, Staikovskiy F, Couderc T, et al. (2013) Chikungunya virus-associated long-term arthralgia: a 36-month prospective longitudinal study. *PLoS Negl Trop Dis* 7: e2137. <https://doi.org/10.1371/journal.pntd.0002137>

29. Chopra A, Anuradha V, Ghorpade R, et al. (2012) Acute chikungunya and persistent musculoskeletal pain following the 2006 Indian epidemic: a 2-year prospective rural community study. *Epidemiol Infect* 140: 842–850. <https://doi.org/10.1017/S0950268811001300>
30. Maek-a-Nantawat W, Silachamroon U (2009) Presence of autoimmune antibody in chikungunya infection. *Case Rep Med* 2009: 1–4. <https://doi.org/10.1155/2009/840183>
31. Gauri LA, Thaned A, Fatima Q, et al. (2016) Clinical spectrum of chikungunya in Bikaner (North Western India) in 2006 and follow up of patients for five years. *J Assoc Physicians India* 64: 22–25.
32. Chen W, Foo SS, Sims NA, et al. (2015) Arthritogenic alpha viruses: new insights into arthritis and bone pathology. *Trends Microbiol* 23: 35–43. <https://doi.org/10.1016/j.tim.2014.09.005>
33. Sugiyama D, Nishimura K, Tamaki K, et al. (2010) Impact of smoking as a risk factor for developing rheumatoid arthritis: a meta-analysis of observational studies. *Ann Rheum Dis* 69: 70–81. <https://doi.org/10.1136/ard.2008.096487>
34. Hedström AK, Rönnelid J, Klareskog L, et al. (2019) Complex relationships of smoking, HLA-DRB1 genes, and serologic profiles in patients with early rheumatoid arthritis: update from a Swedish population-based case–control study. *Arthritis Rheumatol* 71: 1504–1511. <https://doi.org/10.1002/art.40852>
35. Roopa DA, Agrawal N, Johari S, et al. (2015) Prevalence of periodontitis among rheumatoid arthritis patients: An epidemiological study. *Rama Univ J Dent Sci* 2: 2–8.
36. Kim JW, Park JB, Yim HW, et al. (2018) Rheumatoid arthritis is associated with early tooth loss: results from Korea National Health and Nutrition Examination Survey V to VI. *Korean J Intern Med* 34: 1381–1391. <https://doi.org/10.3904/kjim.2018.093>
37. Johansson K, Askling J, Alfredsson L, et al. (2018) Mediterranean diet and risk of rheumatoid arthritis: a population-based case-control study. *Arthritis Res Ther* 20: 1–8. <https://doi.org/10.1186/s13075-018-1680-2>
38. Hu Y, Sparks JA, Malspeis S, et al. (2017) Long-term dietary quality and risk of developing rheumatoid arthritis in women. *Ann Rheum Dis* 76: 1357–1364. <https://doi.org/10.1136/annrheumdis-2016-210431>
39. Tsubaki M, Takeda T, Kino T, et al. (2015) Mangiferin suppresses CIA by suppressing the expression of TNF- α , IL-6, IL-1 β , and RANKL through inhibiting the activation of NF- κ B and ERK1/2. *Am J Transl Res* 7: 1371–1381.
40. Bang SY, Han TU, Choi CB, et al. (2010) Peptidyl arginine deiminase type IV (PADI4) haplotypes interact with shared epitope regardless of anti-cyclic citrullinated peptide antibody or erosive joint status in rheumatoid arthritis: a case control study. *Arthritis Res Ther* 12: 1–9. <https://doi.org/10.1186/ar3051>
41. Hedström AK, Stawiarz L, Klareskog L, et al. (2018) Smoking and susceptibility to rheumatoid arthritis in a Swedish population-based case-control study. *Eur J Epidemiol* 33: 415–423. <https://doi.org/10.1007/s10654-018-0360-5>
42. Laugisch O, Wong A, Sroka A, et al. (2016) Citrullination in the periodontium—a possible link between periodontitis and rheumatoid arthritis. *Clin Oral Investig* 20: 675–683. <https://doi.org/10.1007/s00784-015-1556-7>
43. König MF, Abusleme L, Reinholdt J, et al. (2016) *Aggregatibacter actinomycetemcomitans*-induced hypercitrullination links periodontal infection to autoimmunity in rheumatoid arthritis. *Sci Transl Med* 8: 369ra176. <https://doi.org/10.1126/scitranslmed.aaj1921>

44. Wegner N, Wait R, Sroka A, et al. (2010) Peptidyl arginine deiminase from *Porphyromonas gingivalis* citrullinates human fibrinogen and α -enolase: Implications for autoimmunity in rheumatoid arthritis. *Arthritis Rheum* 62: 2662–2672. <https://doi.org/10.1002/art.27552>
45. Issac THK, Tan EL, Chu JJH (2014) Proteomic profiling of chikungunya virus-infected human muscle cells: reveal the role of cytoskeleton network in CHIKV replication. *J Proteomics* 108: 445–464. <https://doi.org/10.1016/j.jprot.2014.06.003>
46. Fox JM, Diamond MS (2016) Immune-mediated protection and pathogenesis of chikungunya virus. *J Immunol* 197: 4210–4218. <https://doi.org/10.4049/jimmunol.1601426>
47. Phuklia W, Kasisith J, Modhiran N, et al. (2013) Osteoclastogenesis induced by CHIKV-infected fibroblast-like synoviocytes: a possible interplay between synoviocytes and monocytes/macrophages in CHIKV-induced arthralgia/arthritis. *Virus Res* 177: 179–188. <https://doi.org/10.1016/j.virusres.2013.08.011>
48. Wauquier N, Becquart P, Nkoghe D, et al. (2010) The acute phase of Chikungunya virus infection in humans is associated with strong innate immunity and T CD8 cell activation. *J Infect Dis* 204: 115–123. <https://doi.org/10.1093/infdis/jiq006>
49. Hoarau JJ, Bandjee MCJ, Trotot PK, et al. (2010) Persistent chronic inflammation and infection by chikungunya arthritogenic alphavirus in spite of a robust host immune response. *J Immunol* 184: 5914–5927. <https://doi.org/10.4049/jimmunol.0900255>
50. Vossenaar ER, Després N, Lapointe E, et al. (2004) Rheumatoid arthritis specific anti-Sa antibodies target citrullinated vimentin. *Arthritis Res Ther* 6: R142–R150. <https://doi.org/10.1186/ar1057>



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