# G1/S TRANSITION AND CELL POPULATION DYNAMICS

#### Fadia Bekkal-Brikci

Institut de Recherche pour le Développement 32 avenue Henri Varagnat 93143 Bondy Cedex, France

### GIOVANNA CHIORINO

Laboratory of Cancer Pharmacogenomics, Fondo Edo Tempia via Malta 3, 13900 Biella, Italy

### Khalid Boushaba

Iowa State University, Department of Mathematics 482 Carver Hall Ames, IA 50011 USA

(Communicated by Luigi Preziosi)

ABSTRACT. In this paper we present a model connecting the state of molecular components during the cell cycle at the individual level to the population dynamic. The complexes  $Cyclin\ E/CDK2$  are good markers of the cell state in its cycle. In this paper we focus on the first transition phase of the cell cycle  $(S - G_2 - M)$  where the complexe Cyclin E/CDK2 has a key role in this transition. We give a simple system of differential equations to represent the dynamic of the Cyclin E/CDK2 amount during the cell cycle, and couple it with a cell population dynamic in such way our cell population model is structured by cell age and the amount of Cyclin E/CDK2 with two compartments: cells in the G1 phase and cells in the remainder of the cell cycle  $(S - G_2 - M)$ . A cell transits from the G1 phase to the S phase when Cyclin E/CDK2 reaches a threshold, which allow us to take into account the variability in the timing of G1/S transition. Then the cell passes through  $S - G_2 - M$  phases and divides with the assumption of unequal division among daughter cells of the final Cyclin E/CDK2 amount. The existence and the asymptotic behavior of the solution of the model is analyzed.

1. **Introduction.** Since the 80's, cell population dynamics has received much attention from the mathematical modeling community. The first models were related to the proliferation of cell populations. The interest focused not only on the evolution in time of the cell population densities, but also on some other structure variable characterizing individual cells (age, size, *DNA* or *RNA* content, cell maturation). Many differential equations models were built according to the balance between proliferation and cell loss and structured by these variables which influence growth, stabilization, oscillation or other behavior of the population [27] [20] [24] and [51].

Moreover, cells with the same size may have very different age and vice versa, so variability in the cell size and in the cell cycle duration implies that both size and age

<sup>2000</sup> Mathematics Subject Classification. 92D25, 92C40, 58K55.

Key words and phrases. cell cycle,  $G_1/S$  transition, Cyclin E/CDK2, cell population dynamics, unequal cell division, semi group.

K.B is supported by NSF grant DMS-0756669.

structure variables do not give exact information on the state of the cell in its cell cycle. The cell cycle can be split into four phases:  $G_1$ , S,  $G_2$  and M. In mammalian cells, the duration of the  $G_1$  phase is variable whereas the length of S,  $G_2$  and Mphases is relatively constant. The origin of the variability of the  $G_1$  phase duration lies in the amounts and the interactions of the proteins driving the cell from birth to the S phase. Experiments in [12] showed that the S phase cannot be started if the concentration of some proteins which are expressed at the  $G_1$  phase is not high enough. The  $G_1$  phase has been the object of several investigations, especially in relation to the S phase entry (called the  $G_1/S$  transition). The  $G_1/S$  transition is a critical event in the cell cycle progression, and the molecular interactions involved in this transition have a great complexity and require many different families of proteins [23]. Hartwell [16] indicated that cyclins are the most determinant control proteins for phase transitions. These cyclins make complexes with CDK kinases, enzymes which are needed for the cyclin activation, and these complexes are able to phosphorylate other proteins which are important for the cell cycle progression. The more important ones for the  $G_1/S$  transition are the complex formed by Cyclin E and CDK2 kinase (called  $Cyclin\ E/CDK2$ ), the retinoblastoma protein (pRb), the growth factor E2F and the cdk inhibitor  $p27^{KIP1}$  [38, 21]. Under a specific assumptions, several authors [33, 32, 34, 17, 42, 30, 35, 41, 31] have described and simulated part of these complicated reactions regulating the timing of the S phase entry. They have modeled the cell cycle as limit cycles [15, 17, 30, 33], bistable systems [31, 47, 42, 41] and transient processes [1, 23]. The simulations give some insight into the proteins behavior and help us in the choice of the most relevant components influencing the variability of  $G_1$  phase duration [9]. Moreover, even if the cell cycle control is central for cell progression and cell division most of the previous works focus on the simulation of the cell cycle from a molecular standpoint. The connection between the cell population and the cell cycle progression was hardly considered. Our goal in this paper is to present a model taking into account the interdependence between the cell number regulation and the cell cycle progression [7]. Among the few previous works, we cite the paper of Val and Tyson [48]: In their paper, they assumed that molecular mechanisms required in the cell cycle are cell size dependent and then they coupled this cell cycle model with a structured population of budding yeast cells. Actually, they considered three cell cycle components with the cell mass obeying a system of differential equations describing the cell cycle at the molecular level of individual cell and used these four variables as structures of a cell population model. However in this paper, we propose on one hand to connect the cell age with an event implicating a cell component such as a cyclin E/CDK2 complex which is a key entity for the cell cycle control and use this component together with the age as structure variables for the cell population. On the other hand, we take into account the variability of cell cycle duration giving an hypothesis on the amount of cyclins and we also consider an unequal division.

We focus on the critical event: the  $G_1/S$  transition. In section 2, we give a simplified description of the molecular interactions for the  $G_1/S$  transition using the results given in [9, 45, 23]. Afterwards in section 3.1, we present a simple model describing the evolution of only two cell components regulating the  $G_1/S$  transition. In section 3.2 we discuss the variability of the G1 phase duration. Then, in section 3.3 we introduce a linear formulation of these two components in the remain of the cell cycle (S, G2 and M phases). Finally, in section 4 we connect the  $G_1/S$  model with a cell population model with two compartments, which the

first one corresponds to the  $G_1$  phase, with variable duration, and the second to the remainder part of the cell cycle, with constant duration. Then we analyze the behavior of the solution of the cell population model.

2. A quick description of molecular interactions. In this paper, we focus on the  $G_1/S$  transition. The  $Cyclin\ E/CDK2$  complex is the cyclin/CDK complex which has a key role at late  $G_1$  phase for beginning the S phase. Koff et al. [21] observed that the  $Cyclin\ E/CDK2$  increases during  $G_1$  and is maximal in  $G_1$  cells. Afterwards, it decays as cells progress through the remainder of the the cell cycle [37]. The experiments in [45, 9] showed the existence of a threshold value of  $Cyclin\ E$  just before the S phase. Indeed, only cells with  $Cyclin\ E$  amount above the threshold are found in early S phase, while  $G_1$  cells had a very variable content, mostly below the threshold [45].

Hence, we are interested in the molecular interactions that are related to the activity of  $Cyclin\ E/CDK2$  and the fundamental assumption we take into account is that the S phase begins when the level of  $Cyclin\ E/CDK2$  complex reaches a threshold.

Using Kohn's model, Chiorino and Lupi [9], have discussed the molecular network model for the  $G_1$  phase proposed in [23]. They choose this approach because it considers single molecular species and it possible to include all the possible interactions among them. But, of course, the more reactions we add to the system, the more complex it becomes and harder to manage, especially if we want to connect it with a cell population model. Their simulations give the duration of the  $G_1$  phase and its variability as a function of  $G_1$ -related molecular components which have a significative influence on this duration and thus they showed that the higher the pRb level, the longer the  $G_1$  phase, while high initial values of Cyclin E/CDK2 correlate with short  $G_1$  phase. They also remark that the presence of the inhibitor p27 can slow down the  $G_1$  phase, but, after an initial delay, the activity of Cyclin E/CDK2 complex is stronger for a small initial amount of p27. Based on these results, we select the complex Cyclin E/CDK2 and the pRb as the determinant variable of our  $G_1/S$  transition model. Here we don't consider the p27 since we assume that it has a low level at the G1 phase in normal cells.

At the beginning of the cell cycle, pRb is bound to the transcription factor E2F then the complex  $Cyclin\ D/CDK4,6$  phosphorylates partially the pRb but remains bound to the E2F before the so called the restriction point R. Afterwards the activated  $Cyclin\ E/CDK2$  complex achieves the pRb phosphorylation and release E2F. The newly released E2F activates some genes which are important to initiate the S phase (see Figure 1). One of the targets of E2F is to promote the synthesis of  $Cyclin\ E\ [13]$ , [14] (see Figure 1). Then pRb binding to E2F prevent  $Cyclin\ E$  synthesis, which suggests that pRb is responsible for the low expression of  $Cyclin\ E$  gene in early and mid  $C_1$ . Moreover, as well as  $Cyclin\ E$  is synthetisized it binds to the CDK2 kinase and forms the complexe  $Cyclin\ E/CDK2$ . At the  $C_1/S$  transition  $C_1/S$  prevent  $C_2/S$  is completely hyperphosphorylated (inactive) and cannot bind  $C_1/S$  transition hypophosphorylated (active) form in the mitotic phase [29]. Then, we assume that the activity of  $Cyclin\ E/CDK2$  is strongly related to the  $Cyclin\ E/CDK2$  is phosphorylation.

3. Mathematical model. Our aim is not to give an elaborate model of the molecular mechanisms of the  $G_1$  phase but to give a simple system representing the

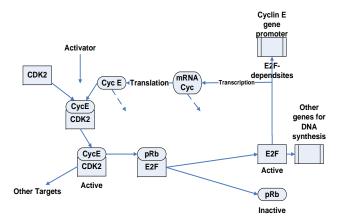


FIGURE 1. : simple scheme of the molecular interactions in the late  $G_1$  phase

progression of the  $Cyclin\ E/CDK2$  which is a key component for the  $G_1/S$  transition and the  $G_1$  phase duration in order to use the  $Cyclin\ E/CDK2$  as a structure variable of the cell population evolving during the cell cycle.

3.1. The  $G_1/S$  transition model: pRb - CycE/CDK2. Based on the molecular mechanisms given in section 2, we give a simple model to describe the activity of  $Cyclin\ E/CDK2$  and pRb phosphorylation in the  $G_1$  phase. We consider that the transition  $G_1/S$  occurs when the level of  $Cyclin\ E/CDK2$  reaches a threshold (denoted  $x_{thr}$ ).

Let x be the amount of  $Cyclin\ E/CDK2$  and y the concentration of hypophosphorylated pRb. Then, we consider x and y as regulating variables in a simple nonlinear system of differential equations with respect to age a.

The  $Cyclin\ E/CDK2$  is regulated by pRb/E2F pathway inducing the pRb phosphorylation. The synthesis of x occurs at the rate k and depends on the concentration (R-y) of the hyper phosphorylated pRb and its degradation occurs at a rate  $d_1$ . Moreover, the total concentration of pRb ( hypo as well as hyper phosphorylated pRb), denoted R, remains constant throughout the cell cycle and we assume that y is chiefly phosphorylated during the  $G_1$  phase by the action of the complex  $Cyclin\ E/CDK2$ , at a rate  $d_2$ . Thus, we give the following system with only two variables  $(Cyclin\ E/CDK2,\ pRb)$  with respect to age a:

$$\begin{cases}
\frac{dx}{da} = k(R - y) - d_1(x - x_{\min}), \\
\frac{dy}{da} = -d_2xy, \\
x(0) = x_i, \quad 0 < x_{\min} \le x_i \le x_{thr}, \\
y(0) = R.
\end{cases}$$
(1)

Where all the parameters are positive  $(k > 0, d_i > 0 \ i = 1, 2, R > 0)$ . We assume that at the beginning of the cell cycle, pRb is totally hypo phosphorylated. Thus the initial concentration of pRb is at its maximum R; whereas the amount of Cyclin E/CDK2 is variable for different cells; bigger than the minimal value  $x_{\min}$  and

smaller than a threshold value denoted  $x_{thr}$ . Now, we investigate the behavior of the solutions of the model described in (1) and we state some results which are useful for further purposes.

**Proposition 1.** System (1) has two steady states  $\left(x_{\min} + \frac{k}{d_1}R, 0\right)$  which is a sink, and  $\left(0, \frac{d_1}{k}x_{\min} + R\right)$  which is a saddle point.

For the global behavior of the solutions of system (1), we state the following proposition.

**Proposition 2.** For any initial conditions  $(x_i, R)$ , such that  $x_i \in (x_{min}, x_{min} + \frac{k}{d_1}R)$ , y always decreases, whereas x can decrease initially, never reaching the minimal value  $x_{min}$  (unless when  $x_i = x_{min}$ ), and then increases towards the asymptotic value  $x_{min} + \frac{k}{d_1}R$ .

proof. We assume that there exists an  $\hat{a} > 0$  such that  $x(\hat{a}) = x_{min}$  then the left derivative of x calculated at  $\hat{a}$  would be less than or equal to zero but from system (1) we remark that whenever  $x = x_{\min}$  ( $\hat{a}$  is the smallest one), the derivative of x is strictly positive, which gives a contradiction. After this initial decrease (if it occurs), x increases towards the asymptotic value:  $x_{\min} + \frac{k}{d_1}R$ . Indeed, x is bounded and if we look at the expression giving the second derivative (x' is the derivative of x and x'' is the second derivative):

$$x'' = -ky' - d_1x',$$

we see that whenever  $\exists \tilde{a} > 0$  such that  $x'(\tilde{a}) = 0$ , x(a) is increasing  $\forall a \geq \tilde{a}$  (the second derivative  $x''(\tilde{a})$  is strictly positive). Moreover, we have

$$y(a) = R e^{-d_2 \int_0^a x(\alpha) d\alpha}$$
 (2)

then  $0 \le y(a) \le R \ \forall a \ge 0$  and y decreases exponentially to 0. Indeed, assuming that

$$\lim_{a \to +\infty} y\left(a\right) > 0,$$

yields  $\int_0^{+\infty} x\left(\alpha\right) d\alpha < +\infty$  then  $\lim_{a \to +\infty} x\left(\alpha\right) = 0$ , which provides a contradiction with  $x\left(a\right) > x_{\min} \ \forall a > 0$ .

Moreover when the initial value x(0) is equal to  $x_{\min}$ , then x always increases.  $\square$ 

**Remark 1.** As observed experimentally [16, 21, 45], we consider that the Cyclin E/CDK2 increases at least in the late  $G_1$  phase until it reaches its maximum at the  $G_1/S$  transition. Moreover, pRb represented by the variable y goes to zero at the end of the  $G_1$  phase which coincides with the fact that the pRb becomes totally hyper phosphorylated at the end of the  $G_1$  phase.

3.2. The G1 phase duration. The main assumption we consider here is that the transition  $G_1/S$  occurs when  $Cyclin\ E/CDK2$  reaches the threshold  $x_{thr}$  provided a minimum time  $T_{\min}$  passed from the beginning of the cell cycle (see [11] for an experimental support of this matter). Therefore we have a variable  $G_1$  phase duration which depends on the initial state of  $Cyclin\ E/CDK2$  [39]. To ensure that x reaches the threshold  $x_{thr}$  we consider the following condition:  $x_{min} < x_{thr} \le x_{\min} + \frac{k}{d_1}R$ . Then, we can state some hypotheses and results on the system (1) which give us some properties on the  $G_1$  phase duration and the cell age at the transition  $G_1/S$ .

**Proposition 3.** For any two initial conditions  $(x_1(0), R)$ ,  $(x_2(0), R)$  of (1) with  $x_1(0) < x_2(0)$  then  $x_1(a) < x_2(a)$ ,  $\forall a > 0$ .

*proof.* Indeed, if there exists an  $a^* > 0$  ( $a^*$  is the smallest one), such that  $x_1(a^*) = x_2(a^*)$  then we have:

$$x_1(a) < x_2(a), 0 < a < a^* \ x_1(a^*) = x_2(a^*)$$
  $\Longrightarrow x'_1(a^*) \ge x'_2(a^*),$ 

but from (1) we have  $y_1(a) = Re^{-d_2 \int_0^a x_1(s)ds}$  and  $y_2(a) = Re^{-d_2 \int_0^a x_2(s)ds}$ . Which implies that  $y_2(a^*) < y_1(a^*)$  and  $x_1'(a^*) < x_2'(a^*)$ , which gives a contradiction.  $\square$ 

**Remark 2.** This proposition can ensure that the higher the initial  $Cyclin\ E/CDK2$  level, the faster  $Cyclin\ E/CDK2$  reaches the threshold and thus the cell passes.

**Lemma 3.1.** if  $x_{thr} < \frac{kR}{d_1}(1 - e^{-d_2T_{min}x_{min}}) + x_{min}$  then there exists  $\bar{a} \geq T_{min}$  such that  $x(\bar{a}) = x_{thr}$ 

*proof.* Since  $y(T_{\min}) \leq R \cdot e^{-d_2 T_{\min} x_{\min}}$  and since we have  $x(a) \leq x_{thr}$  (else the lemma is proved) then

$$\frac{dx}{da} \ge k (R - y) - d_1 (x_{thr} - x_{min})$$

$$\ge kR \left(1 - e^{-d_2 T_{\min} x_{\min}}\right) - d_1 (x_{thr} - x_{\min})$$

which gives the following necssary condition:

if 
$$\frac{kR}{d_1} \left( 1 - e^{-d_2 T_{\min} x_{\min}} \right) > x_{thr} - x_{\min}$$
 then  $\frac{dx}{da} > 0$  for  $a \ge T_{\min}$ .

**Remark 3.** We assume that the degradation rate of the  $Cyclin\ E/CDK2$  in the  $G_1$  phase is low  $(d_1 << 1)$  whereas the degradation of the pRb is high  $(d_2)$  which ensure that the condition in Lemma 3.1 is not a strong constraint.

Then from theses results we can give some conclusions on the  $G_1$  phase duration. So, let  $\varphi_1(a,x_i)$  be the first component of the flow associated to the solution of x in (1) with initial condition  $x_i$  (the  $Cyclin\ E/CDK2$  amount at age a of a cell beginning the cell cycle with the amount  $x_i$ ) and let  $x^*$  be the  $Cyclin\ E/CDK2$  amount such that  $\varphi_1(T_{\min},x^*)=x_{thr}$ . That means an initial amount smaller than  $x^*$  leads to a transition age bigger than  $T_{\min}$  while an initial condition larger than  $x^*$  leads the cell to remain in the  $G_1$  phase until it reaches the minimal age  $T_{\min}$  (see Figure 2). Thus in the first case, the duration of  $G_1$  phase changes and the amount of the  $Cyclin\ E/CDK2$  at the end of  $G_1$  is equal to the threshold value:

$$\varphi_1\left(T_1\left(x_i\right), x_i\right) = x_{thr}.\tag{3}$$

In the second case, the duration of  $G_1$  is the same  $T_{min}$ , but the amount of the  $Cyclin\ E/CDK2$  at the transition is an increasing function of  $x_i$ .

**Remark 4.** The longest duration (denoted T) that the cell can spend in the  $G_1$  phase is associated to the initial value equal to  $x_{min}$ :

$$T_1(x_{min}) = \bar{T} \ge T_1(x_i) \quad \forall x_i \in [x_{min}, x_{thr}], \tag{4}$$

 $\varphi_1(\bar{T}, x_{\min}) = x_{thr}$ , and the largest Cyclin E/CDK2 amount (denoted  $\bar{x}$ ) reached at the end of the  $G_1$  phase is associated to the initial value  $x_{thr}$ :

$$T_1(x_{thr}) = T_{min}, \quad \varphi_1(T_{\min}, x_{thr}) = \bar{x}. \tag{5}$$

From above, we summarize that at the  $G_1/S$  transition we have the cell age  $T_1(x_i)$ , the amount x of Cyclin E/CDK2 and the concentration y of pRb are such that:

$$T_{1}(x_{i}) = \begin{cases} a & \text{with } \varphi_{1}(a, x_{i}) = x_{thr}, & \text{if } x_{min} \leq x_{i} \leq x^{*} \\ T_{\min}, & \text{if } x^{*} \leq x_{i} \leq x_{thr}, \end{cases}$$

$$x = x_{G_{1}}(x_{i}) = \begin{cases} x_{thr} & \text{if } x_{min} \leq x_{i} \leq x^{*} \\ \varphi_{1}(T_{min}, x_{i}) & \text{if } x^{*} \leq x_{i} \leq x_{thr} \end{cases}$$

$$y = y_{G_{1}}(x_{i}) = \begin{cases} R e^{-d_{2} \int_{0}^{T_{1}(x_{i})} \varphi_{1}(\alpha, x_{i}) d\alpha} & \text{if } x_{\min} \leq x_{i} \leq x^{*} \\ R e^{-d_{2} \int_{0}^{T_{\min}} \varphi_{1}(\alpha, x_{i}) d\alpha} & \text{if } x^{*} \leq x_{i} \leq x_{thr} \end{cases}$$

$$(6)$$

**Lemma 3.2.** The function  $T_1(x_i)$  defined in (6) is a decreasing function.

*proof.* From Proposition 3 and Lemma 3.1,  $\varphi_1$  is increasing in  $x_i$  and increasing in a close to  $T_1(x_i)$ . Then, differentiating the function given in (3) with respect to  $x_i$  we obtain:

$$\frac{\partial \varphi_1}{\partial a}_{|a=T_1(x_i)} T_1'(x_i) + \frac{\partial \varphi_1}{\partial x_i} = 0, \tag{7}$$

then

$$T_1'(x_i) < 0, \quad \forall x_i \in [x_{\min}, x_{thr}].$$

That means the required cell age  $T_1(x_i)$  to pass to the S phase is a decreasing function of the initial value of the Cyclin E/CDK2 (see Figure 2 and Figure 3).

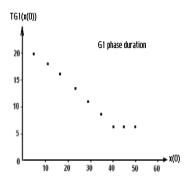


FIGURE 2. This graphic represents the simulation of the  $G_1$  phase duration  $(TG_1)$  as a function of the initial value associated to the variable x and under the assumption that the S phase begins as soon as x reachs a threshold value  $x_{thr}$ , provided a minimum age  $T_{min}$ . The resulting empirical function  $(TG_1)$  has an interval of constancy corresponding to the initial values  $(x^*, x_{thr})$ .

3.3. The S- $G_2$ -M phase formulation. The unequal distribution of some chemicals as cyclins during cell division certainly affects subsequent cell cycles. Therefore, a way to connect cell cycle and proliferation could be that of describing the evolution

of the cyclins until the end of the cell cycle with the assumption of unequal subdivision among daughter cells of the final cyclin content. Then, we have to describe the  $Cyclin\ E/CDK2$  evolution in this remainder of the cell cycle  $(S-G_2-M)$ . In the previous section, the  $G_1$  duration function  $T_1(x_i)$  depends on the initial amount of  $Cyclin\ E/CDK2$  whereas we assume that  $S,\ G_2$  and M phases altogether have fixed duration equal to  $T_2$ , with  $T_2+T_{min}>\bar{T}$ . We assume that the variable x follows (1) until  $\bar{T}$  then decreases linearly with velocity  $\frac{\varphi_1(\bar{T},x_i)-dx_{min}}{T_2}$  whereas y increases linearly to the final value R. That means we describe the evolution of the  $Cyclin\ E/CDK2$  complex and the pRb during the  $S,\ G_2$  and M as follows: for  $T_1(x_i) < a < \bar{T}$ 

$$x(a) = \varphi_1(a, x_i)$$

$$y(a) = Re^{-d_2 \int_0^a \varphi_1(\alpha, x_i) d\alpha}$$
(8)

and for  $\bar{T} < a < T_1(x_i) + T_2$ 

$$x(a) = \varphi_1(\bar{T}, x_i) - \frac{\varphi_1(\bar{T}, x_i) - dx_{\min}}{T_2} (a - \bar{T})$$

$$y(a) = \frac{R - \varphi_2(\bar{T}, x_i)}{T_1(x_i) + T_2 - \bar{T}} (a - \bar{T}) + \varphi_2(\bar{T}, x_i)$$

$$(9)$$

where  $\varphi_2(\bar{T}, x_i) = R e^{-d_2 \int_0^{\bar{T}} \varphi_1(\alpha, x_i) d\alpha}$ 

**Remark 5.** Since x(a) decreases at worst to  $dx_{\min}$  at the end of the cell cycle and have to divide at least with a value bigger than  $x_m in$  for each cell daughter then we assume that d > 2.

Moreover, the system (9) agree with the fact that the  $Cyclin\ E/CDK2$  complex is degraded as cells progress through S phase [40] and that pRb is completely hypophosphorylated at the end of the cell cycle [18], although we do not know whether the dephosphorylation process is gradual or if it occurs only at mitosis. The system (9) can be written under the form of an ODE's system, in the following way:

$$\begin{cases}
\frac{dx}{da} = -\frac{\varphi_1(\bar{T}, x_i) - dx_{min}}{T_2} \\
\frac{dy}{da} = \frac{R - \varphi_2(\bar{T}, x_i)}{T_1(x_i) + T_2 - \bar{T}} \\
x(0) = \varphi_1(\bar{T}, x_i) \\
y(0) = R e^{-d_2 \int_0^{\bar{T}} \varphi_1(\alpha, x_i) d\alpha}
\end{cases}$$
(10)

**Remark 6.** The following condition on the parameters allows us to ensure that, at  $T_1(x_i) + T_2$ , x(a) belongs to the interval  $(dx_{\min}, 2x_{thr})$ :

$$\bar{x} - \frac{\bar{x} - dx_{\min}}{T_2} \left( T_2 + T_{\min} - \bar{T} \right) < 2x_{thr}.$$
 (11)

The conditions given in (5) and (6) ensure that the amount of Cyclin E/CDK2 shared between the two daughters cell belongs to  $(x_{min}, x_{thr})$  in each one.

**Proposition 4.** Let  $X_f$  be the function such that:

$$\begin{array}{cccc} X_f: & (x_{min}, x_{thr}) & \longrightarrow & (dx_{min}, \widehat{x}) \\ & x_i & \longrightarrow & X_f\left(x_i\right) = x_f \end{array}$$

where

$$x_{f} = \varphi_{1}\left(\bar{T}, x_{i}\right) - \frac{\varphi_{1}\left(\bar{T}, x_{i}\right) - dx_{\min}}{T_{2}}\left[T_{1}\left(x_{i}\right) + T_{2} - \bar{T}\right].$$

Then  $X_f$  is invertible.

*proof.* We will prove the above proposition in the case corresponding to a variable duration of the  $G_1$  phase, the other situation being easier to treat (i.e. when  $T_1(x_i) = T_{\min}$ ).

From the definition of  $X_f$  we have:

$$X_{f}(x_{i}) = \varphi_{1}\left(\bar{T}, x_{i}\right) - \frac{\varphi_{1}\left(\bar{T}, x_{i}\right) - dx_{\min}}{T_{2}} \left[T_{1}\left(x_{i}\right) + T_{2} - \bar{T}\right].$$

That means the final value  $x_f$  depends on the initial value  $x_i$ . The derivative of this function is:

$$X_f'\left(x_i\right) = \frac{\partial \varphi_1}{\partial x_i} \left(\bar{T}, x_i\right) \left[\frac{\bar{T} - T_1\left(x_i\right)}{T_2}\right] - \frac{\varphi_1\left(\bar{T}, x_i\right) - dx_{\min}}{T_2} T_1'\left(x_i\right).$$

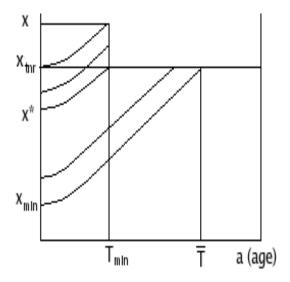
Then we are interested in its sign. We have  $T_1(x_i) < 0$  and  $T_1(x_i) \le \bar{T}$ . Moreover  $\varphi_1(\bar{T}, x_i) \ge dx_{min}$ . Then  $X_f'(x_i) \ge 0$  which implies that  $X_f$  is invertible in  $(x_{min}, x_{thr})$  and thus  $x_i$  is a function of  $x_f$ .

**Remark 7.** This proposition means that from any final value  $x_f = X_f(x_i)$  of  $Cyclin\ E/CDK2$  associated to mother cell at mitosis it is possible to go back to the initial content  $x_i$  which has generated the final state  $x_f$ .

- 4. Connection with the population level. In a population of normal cells there is a balance between proliferation, quiescence and cell death [44]. This balance is maintained by a system of closely interconnected inhibitory and stimulatory signals and ensure tissue homeostasis, but embryonic cells or cancer cell lines growing under optimal conditions present different features avoiding cell quiescence, cell loss and the strong dependence on growth factor stimulation. Our attention was focused on cancer cell lines growing in suspension. Obviously, immortal cell lines growing in vitro do not give a correct idea of the complexity of the environment where either normal or tumor cells grow. Nevertheless, cell lines are a very important tool for the study of cell cycle regulation and cell proliferation, and of cell kinetic parameters, such as cell cycle duration and phase transition times.
- 4.1. Formulation of the evolution problem. In the previous section, we considered the evolution of  $Cyclin\ E/CDK2$  inside a single cell and the way to regulates the timing of cell cycle. We now aim to take into account a cell population structured by the time spent inside the cell cycle since birth (variable a) and the amount of  $Cyclin\ E/CDK2$  complex (variable x). The evolution of x is described by the following equations:

$$\begin{cases} \frac{dx}{da} = \Gamma(a, x) \\ x(0) = x_i \end{cases}$$

where



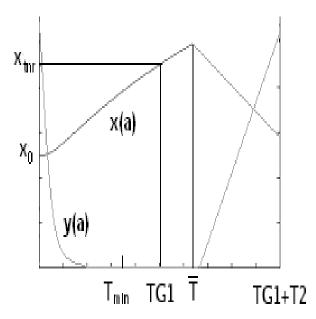


FIGURE 3. The first graphic shows the evolution of the complex CyclinE/CDK2 (x) during the  $G_1$  phase. For the initial conditions bigger than  $x^*$  we have  $x(TG_1) = x(T_{min})$  whereas for the initial conditions smaller than  $x^*$  we have  $x(TG_1) = x_{thr}$  where  $TG_1 > T_{min}$ . The second graphic shows the evolution of the complex CyclinE/CDK2 (x) and the pRb (y) during the cell cycle.

$$\Gamma(a,x) = \left\{ \begin{array}{ll} \Gamma_1(a,x) & for \ 0 < a < T_1(x_i), \\ \Gamma_2(a,x) & for \ T_1(x_i) < a < T_1(x_i) + T_2, \end{array} \right.$$

$$\Gamma_{1}\left(a,x\right) = kR\left(1 - e^{-d_{1}\int_{0}^{a}\varphi_{1}\left(\alpha,x_{i}\right)d\alpha}\right) - c\left(x - x_{\min}\right),\,$$

and

$$\Gamma_2(a,x) = \begin{cases} k R(1 - e^{-d_1 \int_0^a \varphi_1(\alpha, x_i) d\alpha}) - \\ c(x - x_{\min}) & \text{if } T_1(x_i) < a < \bar{T} \\ -\frac{\varphi_1(\bar{T}, x_i) - dx_{\min}}{T_2} & \text{if } \bar{T} < a < T_1(x_i) + T_2 \end{cases}$$

where  $\varphi_1(a, x_i)$  is the solution of the first equation associated to the initial condition  $x_i$  at the age a. We denote  $P_1(t, a, x)$  and  $P_2(t, a, x)$  the densities, with respect to a and x, of cells that, at time t, are in the  $G_1$  phase or in the remainder part of the cell cycle, respectively. Then, we can write the following partial differential equations associated to  $P_i$ , i=1,2:

$$\begin{cases}
\frac{\partial P_1}{\partial t} + \frac{\partial P_1}{\partial a} + \frac{\partial}{\partial x} [\Gamma(a, x) P_1(t, a, x)] = 0 \\
\frac{\partial P_2}{\partial t} + \frac{\partial P_2}{\partial a} + \frac{\partial}{\partial x} [\Gamma(a, x) P_2(t, a, x)] = -F(x) P_2(t, a, x)
\end{cases}$$
(12)

The left sides of the equations represent the variation of the subpopulation  $P_1$  and  $P_2$ with respect to time, age and Cyclin E/CDK2 amount where  $\gamma$  represents the speed of x with respect to the physiological age a. Moreover, we assume that no mortality occurs during cell cycle progression (as we have discussed before this assumption is true as far as we deal with cell cultures growing under optimal conditions) and finally the term  $F(x)P_2(t,a,x)$  is the cell subpopulation density which have the Cyclin E/CDK2 amount equal to x when they leave the cell cycle.

The boundary conditions describing the flow of cells from one compartment  $(G_1)$ to the other (S, G2, M) are the following:

- $\begin{array}{ll} \bullet \ \ ({\rm a}) \ \ P_2(t,a,x_{thr}) = P_1(t,a,x_{thr}), & T_{\min} < a < \bar{T}; \\ \bullet \ \ ({\rm b}) \ \ P_2(t,T_{\min},x) = P_1(t,T_{\min},x), & x_{thr} < x < \overline{x}. \end{array}$

Now, we have to consider the boundary condition that determines the cell division event at the end of the second compartment and the birth of two new cells which enter the cell cycle at the beginning of the  $G_1$  phase (a = 0) but no such condition is needed at x = 0, since cyclin level x = 0 is never reached (2).

4.2. Unequal division. The distribution of the molecular materials between daughter cells such as cyclins is assumed to be unequal. That means the amount xof the Cyclin E/CDK2 inherited by a newborn cell from its mother follows a conditional density f(x,x'), x' being the mother state at the end of its cell cycle [20]. In [2, 3, 4, 5] such condition is given for a model describing the evolution of cells classified according to a constituent which, after division (by mitosis), is divided in a certain stochastic way amongst the two daughter cells. Then the fraction F(x)of cells which have x Cyclin E/CDK2 content leave the cell cycle to undergo cell division - disappearing and being replaced by two daughter cells - is defined by:

$$F(x) = \int_{x_{min}}^{x_{thr}} f(x', x) dx'$$
 (13)

Therefore, we have the following condition at the boundary a=0 which corresponds to the new borns density:

$$P_1(t,0,x) = 2\int_0^{+\infty} f(x,x')P_2[t,T_2 + T_1(X_f^{-1}(x')),x']dx'$$
(14)

where  $X_f^{-1}$  is the inverse of the function  $X_f$  that supplies the value of x at mitosis, given  $x_i$  at the beginning of the cell cycle. From the above definition of the function f, we can state the following properties:

- 1. The daughter amount of cyclins cannot be bigger than the mother one: f(x, x') = 0 if x > x'.
- 2. The amount x' of the Cyclin E/CDK2 of the mother cell is exactly conserved and shared by the two daughters: f(x, x') = f(x' x, x').
- 3. The amount x of the Cyclin E/CDK2 of the daughter cells entering the  $G_1$  phase belongs to the interval  $[x_{\min}, x_{thr}]$ : f(x, x') = 0 if  $x \notin [x_{\min}, x_{thr}]$ , which yields

$$\int_{x_{-in}}^{x_{thr}} f(x, x') dx = 1.$$
 (15)

4. There exists a positive value  $x_1$  such that  $x_{\min} < x_1 < x^* < x_{thr}$  ans a, b such that 0 < a < b, with  $aX_f(x_1) = x_1$  and  $bX_f(x_{thr}) = x_{thr}$ ,  $aX_f(x) > x$  for  $x > x_1$ ,  $bX_f(x) > x$  for  $x > x_{thr}$ ,  $aX_f(x) < x$  for  $x < x_1$ , and  $ax_f(x) < x$  for  $ax_f(x) < x$  for

Supp 
$$2f(x, X_f(y)) = [aX_f(y), bX_f(y)].$$

This property allows us to show that if P is a solution of (27), whose support is initially contained in the interval  $(x_{\min}, x_{thr})$ , then P(t, x) = 0 for t > 0, and  $x < x_{\min}$  or  $x > x_{thr}$ . Moreover, the x-support shrinks asymptotically to the interval  $[x_1, x_{thr}]$ .

- 5.  $f(x, X_f(y))$  is bounded with respect to both variables x and y.
- 6.  $2f(x, X_f(y))$  is nonnegative, continuous and  $2\int_0^\infty f(x, X_f(y))dx > 1$ .

4.3. The cell cycle duration  $T_c$ . Here, we take into account the variability of the cell cycle duration. We denote  $T_c$  the function of the total cell cycle duration defined as follows:

$$T_c(y) = T_1(y) + T_2, \quad \forall y \in [x_{\min}, x_{thr}].$$
 (16)

where  $T_1(y)$  corresponds to the G1 phase duration and  $T_2$  to the duration of the remainder of the cell cycle. Using (6) and Lemma 3.2 we obtain the following results:

1.  $T_c$  is continuous, nondecreasing such that

$$0 < T_{\min} + T_2 < T_c(y) < \bar{T} + T_2 < +\infty. \tag{17}$$

and

$$T_c(y) = T_{\min} + T_2 \quad \forall y \in [x^*, x_{thr}]. \tag{18}$$

2. There exists a constant  $\theta' > 0$ , such that

$$|T_c'(y)| \ge \theta' \quad \forall y \in [x_{\min}, x^*] \tag{19}$$

Inequality (19) can be assured by proposition 3 (see also the graphic representation of  $T_1$  shown in Figure 2).

4.4. **Analysis and qualitative behavior.** We now perform the analysis of the model developed above. We use for this purpose the method based on the theory of semigroups following the approach in [2].

Using the characteristic lines corresponding to (12) we obtain:

$$\begin{cases} P_1(t(s), a(s), x(s)) = P_1(t_0, a_0, x_0)e^{-\int_0^s \frac{\partial \Gamma_1}{\partial x}(\tau, \varphi_1(\tau, x_i))d\tau}, \\ P_2(t(s), a(s), x(s)) = \\ P_2(t_0, a_0, x_0)e^{-\int_0^s \left(\frac{\partial \Gamma_2}{\partial x}(\tau, \varphi_1(\tau, x_i)) + F(\varphi_1(\tau, x_i)))\right)d\tau}. \end{cases}$$

Therefore at the  $G_1/S$  transition we have:

$$\begin{cases}
P_{1}(t, T_{1}(X_{f}^{-1}(x)), x_{thr}) = \\
P_{1}(t - T_{1}(X_{f}^{-1}(x)), 0, X_{f}^{-1}(x))e^{-r_{1}(x)} \quad for \ x \in [x_{min}, x^{*}] \\
P_{1}(t, T_{min}, \varphi_{1}(T_{min}, X_{f}^{-1}(x))) = \\
P_{1}(t - T_{min}, 0, X_{f}^{-1}(x))e^{-r_{2}(x)} \quad for \ x \in [x^{*}, x_{thr}]
\end{cases} (20)$$

where

$$r_1(x) = \int_0^{T_1(X_f^{-1}(x))} \frac{\partial \Gamma_1}{\partial x} (s, \varphi_1(s, X_f^{-1}(x))) ds$$
 (21)

and

$$r_2(x) = \int_0^{T_{min}} \frac{\partial \Gamma_1}{\partial x} (s, \varphi_1(s, X_f^{-1}(x))) ds$$
 (22)

and at the age of the division, we have:

$$\begin{cases}
P_{2}(t, T_{1}(X_{f}^{-1}(x)) + T_{2}, x) = \\
P_{2}(t - T_{2}, T_{1}(X_{f}^{-1}(x)), x_{thr})e^{-r_{3}(x)} & for \ x \in [x_{min}, x^{*}] \\
P_{2}(t, T_{min} + T_{2}, x) = \\
P_{2}(t - T_{2}, T_{min}, \varphi_{1}(T_{min}, X_{f}^{-1}(x)))e^{-r_{4}(x)} & for \ x \in [x^{*}, x_{thr}]
\end{cases} (23)$$

where

$$r_{3}(x) = \int_{T_{1}(X_{f}^{-1}(x))}^{\bar{T}} \frac{\partial \Gamma_{2}}{\partial x} (s, \varphi_{1}(s, X_{f}^{-1}(x))) ds + \int_{T_{1}(X_{f}^{-1}(x))}^{T_{1}(X_{f}^{-1}(x))} F(\varphi_{1}(s, X_{f}^{-1}(x))) ds$$

$$(24)$$

and

$$r_{4}(x) = \int_{T_{min}}^{\bar{T}} \frac{\partial \Gamma_{2}}{\partial x} (s, \varphi_{1}(s, X_{f}^{-1}(x))) ds + \int_{T_{min}}^{T_{min} + T_{2}} F(\varphi_{1}(s, X_{f}^{-1}(x))) ds.$$
(25)

Using the boundary condition (14) and the system (23), we obtain:

$$P_{1}(t,0,x) = 2 \int_{0}^{+\infty} f(x,x') P_{2}(t,T_{1}(X_{f}^{-1}(x')) + T_{2},x') dx'$$

$$= 2 \int_{X_{f}(x_{\min},x^{*})} f(x,x') P_{2}(t-T_{2},T_{1}(X_{f}^{-1}(x')),x_{thr}) e^{-r_{3}(x')} dx' +$$

$$2 \int_{X_{f}(x^{*},x_{thr})} f(x,x') P_{2}(t-T_{2},T_{\min},\varphi_{1}(T_{\min},X_{f}^{-1}(x))) e^{-r_{4}(x')} dx'.$$
(26)

Moreover, from the system (20) and the conditions (a) and (b), we obtain:

$$\begin{split} &P_1(t,0,x) = \\ &2 \int_{X_f(x_{\min},x^*)} \\ &f(x,x') P_1(t-T_1(X_f^{-1}(x'))-T_2,0,X_f^{-1}(x')) e^{-(r_1(x')+r_3(x'))} dx' + \\ &2 \int_{X_f(x^*,x_{thr})} \\ &f(x,x') P_1(t-T_{\min}-T_2,0,X_f^{-1}(x')) e^{-\left(r_2(x')+r_4(x')\right)} dx', \end{split}$$

Now, let P(t,x) be such that  $P(t,x) = P_1(t,0,x)$  and  $y = (X_f)^{-1}(x')$ . Then the integral equation (26) becomes:

$$P(t,x) = 2 \int_{x_{min}}^{x^*} f(x, X_f(y)) P(t - T_1(y) - T_2, y) e^{-R_1(y)} dy + 2 \int_{x^*}^{x_{thr}} f(x, X_f(y)) P(t - T_{min} - T_2, y) e^{-R_2(y)} dy$$
(27)

where

$$R_{1}(y) = \int_{0}^{T_{1}(y)} \frac{\partial \Gamma_{1}}{\partial x}(s, \varphi_{1}(s, y))ds + \int_{T_{1}(y)}^{T} \frac{\partial \Gamma_{2}}{\partial x}(s, \varphi_{1}(s, y))ds + \int_{T_{1}(y)}^{T_{1}(y)+T_{2}} F(\varphi_{1}(s, y))ds$$
(28)

and

$$R_{2}(y) = \int_{0_{-}}^{T_{min}} \frac{\partial \Gamma_{1}}{\partial x}(s, \varphi_{1}(s, y))ds + \int_{T_{min}}^{T} \frac{\partial \Gamma_{2}}{\partial x}(s, \varphi_{1}(s, y))ds + \int_{T_{min}}^{T_{min} + T_{2}} F(\varphi_{1}(s, y))ds.$$

$$(29)$$

The equation (27) is a delay integral equation involving only the population of  $G_1$  phase cells which are about to start a new cell cycle. The delay term corresponds to the duration of the cell cycle which from (6) it is constant and equal to  $(T_{\min} + T_2)$  if the initial value associated to y belongs to the interval  $(x^*, x_{thr})$  otherwise it is bigger than  $T_{\min} + T_2$ . The maximal cell cycle duration corresponds to  $\bar{T} + T_2$ . We denote  $\rho = \bar{T} + T_2$  the maximal delay.

If we consider the time interval  $[-\rho, 0]$ , we will see that the equation (27) is associated to a dynamical system in a class of functions on  $(-\rho, 0) \times (x_{min}, x_{thr})$ .

We choose the positive cone of the Lebesgue space

$$L^1_+((-\rho,0)\times(x_{min},x_{thr}))=X,$$

as the space of the initial data. Let  $P_t$  be the trajectory defined as follows:

$$P_t(\theta, x) = P(t + \theta, x). \tag{30}$$

**Proposition 5.** Let  $P_0$  be in X (or in  $X^{\infty}$ ), and let P be defined on  $(-\rho, T_2 + T_{\min}) \times (x_{\min}, x_{thr})$  by  $P = P_0$  on  $(-\rho, 0) \times (x_{\min}, x_{thr})$  and by

$$P(t,x) = 2 \int_{x_{min}}^{x^*} f(x, X_f(y)) e^{-R_1(y)} P_0[t - T_2 - T_1(y), y] dy + 2 \int_{x^*}^{x_{thr}} f(x, X_f(y)) e^{-R_2(y)} P_0(t - T_2 - T_{min}, y) dy,$$

on  $(0, T_2 + T_{min}) \times (x_{min}, x_{thr})$ . Then for each t in  $(0, T_2 + T_{min})$ ,  $P_t$  is in X (or in  $X^{\infty}$ ) and  $||P_t||_X \leq C ||P_0||_X$ ,  $0 < t < T_2 + T_{min}$ , where C is independent of  $P_0$ .

proof. Since the minimum delay is  $T_{\min} + T_2$ , then the solution is automatically known on the interval  $[0, T_{\min} + T_2]$  by means of simple integration, and therefore we have just to check that this first portion of the solution is in X (or in  $X^{\infty}$ ), given  $P_0$  in X (or in  $X^{\infty}$ ). To do that, we consider

$$\int_{0}^{T_{2}+T_{\min}} \int_{x}^{x_{thr}} |P(t,x)| dxdt$$

and we integrate it first in the variable x (using the probability property (15)) and then in t. This gives:

$$\begin{split} & \int_{0}^{T_{2}+T_{\min}} \int_{x_{\min}}^{x_{thr}} |P\left(t,x\right)| \, dxdt \\ & \leq 2 \int_{0}^{T_{2}+T_{\min}} \left\{ \int_{x_{\min}}^{x^{*}} |P\left(t-T_{2}-T_{1}\left(y\right),y\right)| \, dy \right\} dt + \\ & 2 \int_{0}^{T_{2}+T_{\min}} \left\{ \int_{x^{*}}^{x_{thr}} |P\left(t-T_{2}-T_{\min},y\right)| \, dy \right\} dt \\ & \leq 2 \int_{-\rho}^{0} \left\{ \int_{x_{\min}}^{x^{*}} |P\left(\theta,y\right)| \, dy \right\} d\theta \\ & + 2 \int_{-\rho}^{0} \left\{ \int_{x^{*}}^{x_{thr}} |P\left(\theta,y\right)| \, dy \right\} d\theta \\ & \leq C \int_{-\rho}^{0} \int_{x_{\min}}^{x_{thr}} |P\left(\theta,y\right)| \, dy d\theta. \end{split}$$

We have therefore come back to the interval  $(-\rho,0) \times (x_{\min}, x_{thr})$ . So, this shows that, starting from any element in X, the solution remains in X, at least for all  $t \in [0, T_{\min} + T_2]$ . For initial values in  $X^{\infty}$ , we just have to notice that f being a distribution probability in the x, the right hand side of equation (27) is uniformly bounded on the domain  $[0, T_{\min} + T_2] \times (x_{\min}, x_{thr})$ .

Characterization of the solution Using the approach developed in [2], we can prove the following theorem which gives the asymptotic behavior of the solution of 27.

**Theorem 4.1.** Let us assume that all the properties defined above for the functions  $T_c$  and f are verified. Then, there exist a positive number  $\lambda^*$  and a continuous function  $\mu^*$  such that:

$$\mu^*(x) > 0, \quad x_1 < x < x_{thr},$$
  
 $\mu^*(x) = 0, \quad x \le x_1 \quad or \quad x \ge x_{thr},$ 

and, if P is a solution of (27) for  $t \ge t_0$  with support  $P_t(\theta, \cdot)$  in  $[x_{\min}, x_{thr}]$ , then there exists a constant C' with the property that:

$$P(t,x) - C' \exp(\lambda^* t) \mu^*(x) = o \left[ \exp(\lambda^* t) \right], \qquad t \to \infty$$

If P > 0, then C' > 0.

First of all, we have to establish the existence of the stationary solution. If we substitute the expression  $\exp(\lambda^*t)\mu^*(x) + o\left[\exp(\lambda^*t)\right]$ , for P in (27), we see that the pair  $(\lambda^*, \mu^*)$  is a solution of the following equation:

$$\mu(x) = 2 \int_{x_{\min}}^{x_{thr}} f(x, X_f(y)) e^{-\lambda T_2 - \lambda T_1(y)} \mu(y) \, dy = L_{\lambda}(\mu)(x), \tag{31}$$

where  $T_1(y) = T_{\min}$ ,  $\forall y \in (x^*, x_{thr})$ .

For each  $\lambda,\,L_\lambda$  is an integral operator with a nonnegative continuous kernel:

$$k_{\lambda}(x,y) = 2e^{-\lambda T_2 - \lambda T_1(y)} f(x, X_f(y)).$$

Therefore,  $L_{\lambda}$  is a positive compact operator on each class  $L^{p}\left(x^{*}, x_{thr}\right), p \geq 1$ . We will use the adjoint  $L_{\lambda}^{*}\nu$  of  $L_{\lambda}$  with respect to the scalar product in  $L^{2}\left(x_{\min}, x_{thr}\right)$ :

$$L_{\lambda}^{*}\nu = \int_{x_{\min}}^{x_{thr}} k_{\lambda}(x, y) \nu(x) dx.$$

We have to find a nontrivial fixed point of  $L_{\lambda}$  in the cone of the nonnegative functions.

**Theorem 4.2.** Suppose that property 4 for the function f be verified. Then the set  $\Lambda$  of the (real and complex) numbers such that  $(\lambda, \mu)$  is a solution of (31) for some  $\mu \neq 0$  in  $L^2(x_{\min}, x_{thr})$  is discrete. Set  $\Lambda$  has a unique element  $\lambda^*$  with the greatest real part:  $\lambda^*$  is real and the space of the functions  $\mu$  such that  $(\lambda^*, \mu^*)$  is a solution of (31), is one-dimensional, generated by a nonnegative function  $\mu^*$ . Moreover, the one-dimensional space of the functions  $\nu$  in  $L^2(x_{\min}, x_{thr})$  satisfying  $L^*_{\lambda^*}\nu = \nu$  is generated by a strictly positive function  $\nu^*$ .

Theorem 4.2 has been proved in [2] under analogous hypotheses. To prove the existence of a pair  $(\lambda^*, \mu^*)$  the authors used a result by Bonsall. Then they proved the discreteness of  $\Lambda$ . An important role in the demonstration of the last part of theorem 4.2 is played by the above mentioned property 4 of the function f, since it allows to show that the support of the iterates of  $L^*_{\lambda^*}$  fills asymptotically the interval  $[x_1, x_{thr}]$ .

Semigroup of the operator associated to (27) and its infinitesimal generator. Now, we associate to (27) the family of operators defined by:

$$T(t) P_0 = P_t. (32)$$

As a consequence, using the fact that the translations are continuous in  $L^1$ , we can assert that  $t \longrightarrow P_t$  is continuous from  $\mathbb{R}^+$  into X.

**Proposition 6.** The family of operators T(t) defined by (32) sends X into itself and constitutes on X a strongly continuous semigroup of operators.

Now, we will describe the infinitesimal generator of the semigroup  $T\left(t\right)$  and examine its spectral properties. We know that it is a closed operator with a dense domain and it is given by:

$$A\chi = \lim_{t \to 0^{+}} \frac{\left[T(t)\chi - \chi\right]}{t}$$

for all the  $\chi$ 's for which the limit above exists in X. Formally, the generator is defined by

$$(A\chi)(s,x) = \frac{\partial \chi(s,x)}{\partial s}, \quad -\rho < s < 0, \quad x_{\min} < x < x_{thr},$$

with  $\frac{\partial \chi}{\partial s}$  in X. This means that  $\chi$  is absolutely continuous from  $[-\rho, 0]$  into  $L^1(]x_{\min}, x_{thr}[)$  and so  $\chi(s, \cdot)$  is well defined for each s in  $[-\rho, 0]$  and continuous. At s = 0, it coincides with  $\lim_{t \to 0^+} T(t) \chi$ , yielding:

$$\chi(0,x) = 2 \int_{x_{\min}}^{x_{thr}} \tilde{f}(x,y) \chi(-T_1(y) - T_2, y) dy,$$
 (33)

where

$$\tilde{f}(x,y) = \begin{cases} f(x, X_f(y))e^{-R_1(y)} & if \ y \in (x_{min}, x^*) \\ f(x, X_f(y))e^{-R_1(y)} & if \ y \in (x^*, x_{thr}) \end{cases}$$

Conversely, one can check (we skip this) that any  $\chi$  in X such that  $\chi$  is absolutely continuous as a function from  $[-\rho,0]$  into  $L^1(]x_{\min},x_{thr}[)$  and  $\chi$  satisfies the boundary condition (33) is in the domain of A. Therefore, we have established that the set D(A) of all the  $\chi \in X$  such that  $\chi$  is absolutely continuous from  $[-\rho,0]$  into  $L^1(]x_{\min},x_{thr}[)$  satisfies (33) and, for  $\chi$  in D(A), we have  $A\chi = \frac{\partial \chi}{\partial s}$ . Now, we consider the resolvent equation,

$$A\chi - \lambda\chi = \xi, \quad \xi \in X; \tag{34}$$

The equation (34) can be integrated in the form of

$$\chi(s,x) = \chi(0,x) \exp(\lambda s) + \int_0^s \exp[\lambda(s-u)] \,\xi(u,x) \,du. \tag{35}$$

Assuming that  $\chi(0,x)$  is well defined and belongs to  $L^1(]x_{\min},x_{thr}[)$ , it is straightforward to see that the right hand side of formula (35) is indeed absolutely continuous as a function of s into  $L^1$ . Substituting this term for  $\chi(s,x)$  in (33), we obtain the following equation:

$$\chi(0,x) = 2 \int_{x_{\min}}^{x_{thr}} \tilde{f}(x,y)\chi(0,y) \exp(-\lambda(T_1(y) + T_2))dy + 2 \int_{x_{\min}}^{x_{thr}} \tilde{f}(x,y) 
\left( \int_{0}^{-T_1(y) - T_2} \exp\left[-\lambda(T_1(y) + T_2) - u\right] \xi(u,y)du \right) dy.$$
(36)

where

$$T_1(y) = T_{\min} \text{ if } y \in (x^*, x_{thr}),$$

The linear operator in equation (36) is similar to the operator  $L_{\lambda}$  defined in equation (31). It has the same fixed points with the same multiplicity. We denote by  $S_{\lambda}$  the operator acting on  $\xi$ .

Compactness of  $L_{\lambda}$  yields that existence of a solution to equation (36) for all  $\xi$  is equivalent to  $(I-L_{\lambda})$  being invertible, which itself is equivalent to  $(I-L_{\lambda})$  being one-to-one. So the spectrum of the generator is precisely the set of  $\lambda's$  for which the null set of  $(I-L_{\lambda})$  is  $\neq 0$ . Now, for  $\lambda$  in the resolvent set, the solution associated to each  $\xi$  reads in terms of  $\xi$  as

$$\chi(s,x) = \exp(\lambda s) \left(I - L_{\lambda}\right)^{-1} S_{\lambda} \xi + \int_{0}^{s} \exp\left[\lambda \left(s - u\right)\right] \xi\left(u, x\right) du. \tag{37}$$

Now, it is easy to see that  $S_{\lambda}$  is compact, as it is essentially an integral operator in both variables and on bounded domains. So, formula (37) defines a decomposition of the resolvent of  $(\lambda - A)^{-1}$  into the sum of a compact part which contains information about the spectrum and a noncompact part which seems to be neutral in this respect.

It can be proved (see [2]) that the infinitesimal generator A enjoys then the following properties:

- 1. The spectrum of A,  $\sigma(A)$ , is a pure point spectrum;
- 2.  $\sigma(A) = \Lambda$ ;
- 3. for each  $\lambda$ ,  $Im(A \lambda I)$  is closed in X with  $Ker(A \lambda I) = e^{\lambda s} \otimes Ker(I L_{\lambda})$ ;
- 4. for  $\lambda = \lambda^*$ ,  $S = Im(A \lambda^*I)$  has codimension 1 in X and so  $X = N \oplus S$ .

On S,  $(A - \lambda^* I)$  is a bijection from  $S \cap D(A)$  onto S:  $(\sigma(A_{|S})) \subseteq \{Re\lambda \leq \lambda_1\}$  for some  $\lambda_1 \leq \lambda^*$ . A generates a semigroup on S,  $T_S(t)$ , which can be completed into a semigroup on X,  $\widetilde{T}_S$ , such that

$$\widetilde{T}_S(t)\left(e^{\lambda^*}\mu^*\right) = e^{\lambda^*(t+s)}\mu^*.$$

The infinitesimal generator of  $\widetilde{T}_S$  is A, so  $\widetilde{T}_S = T$  and  $T_{|S} = T_S$ . Therefore S is invariant with respect to T(t).

However, since the semigroup T(t) is not compact, we are not allowed to extend the decomposition of the generator to a decomposition of the semigroup. Nor we are able to assert in general that the spectrum of the semigroup restricted to S,  $\sigma\left[T(t)_{|S}\right]$  is contained in a complex ball  $\left\{z:|z|\leq e^{\lambda_1 t}\right\}$  for some  $\lambda_1<\lambda^*$ , which would immediately prove theorem 4.1.

Properties of the semigroup T(t). Our aim is to show that, for t large enough, the semigroup T(t) is "quasi compact", that is, it can be decomposed into the sum of two operators, one of which is compact, while the other is "small", in the sense that its growth rate is dominated by the growth rate of the compact part. This way, we will be able to prove that the solution asymptotically reaches asynchronous exponential growth.

Let us introduce the following notations:

$$(KP)(t,x) = 2 \int_{x_{\min}}^{x^*} \widetilde{f}(x,y) P[t - T_2 - T_1(y), y] dy, \quad P \in X.$$
 (38)

where  $K: X \longrightarrow X$ .

Iterating the operator K yields:

$$(K^{2}P)(t,x) = 2 \int_{x_{\min}}^{x^{*}} \widetilde{f}(x,y) (KP) [t - T_{2} - T_{1}(y), y] dy$$

$$= 4 \int_{x_{\min}}^{x^{*}} \widetilde{f}(x,y)$$

$$\int_{x_{\min}}^{x^{*}} \widetilde{f}(y,z) \{P[t - 2T_{2} - T_{1}(y) - T_{1}(z), z] dz\} dy.$$

**Proposition 7.** Under the assumptions that property 5. for function  $\widetilde{f}$  be verified and that

$$|T_1'|_{\infty} + \left| \left( T_1^{-1} \right)' \right|_{\infty} < +\infty,$$

then the operator  $K^2$  sends functions of  $L^1$  into bounded functions:

$$K^2: L^1 \longrightarrow L^\infty$$
.

proof. We have:

$$\left| \left( K^{2}P\right) (t,x) \right| \leq 4 \left| \widetilde{f} \right|_{\infty}^{2} \int_{x_{min}}^{x^{*}} \int_{x_{min}}^{x^{*}} \left| P\left[ t - 2T_{2} - T_{1}\left( y \right) - T_{1}\left( z \right), z \right] \right| dz dy.$$

Now, the following change of variable  $\tau = T_1(y)$ , gives:

$$\begin{split} &\left|\left(K^{2}P\right)\left(t,x\right)\right| \\ &\leq 4\left|\widetilde{f}\right|_{\infty}^{2} \\ &\int_{x_{\min}}^{x^{*}} \left\{\int_{T_{1}\left(x^{*}\right)}^{T_{1}\left(x_{\min}\right)}\left|P\left[t-2T_{2}-\tau-T_{1}\left(z\right),z\right]\right|\left|\left(T_{1}^{-1}\right)'\left(\tau\right)\right|d\tau\right\}dz \\ &\leq \gamma\left(t\right)\left|P_{0}\right|_{L^{1}} \Longrightarrow \forall t>0:K^{2}P\in L^{\infty}. \end{split}$$

**Proposition 8.** The operator  $K^2$  is compact from  $L^{\infty}$  to  $L^1$ .

proof. We now introduce another change of variables:

$$\begin{cases} \tau = t - 2T_2 - T_1(y) - T_1(z) \\ z = z \end{cases}$$

and write the operator  $K^2$  as follows:

$$\begin{split} \left(K^{2}P\right)(t,x) &= 4 \int_{x_{\min}}^{x^{*}} \int_{t-2T_{2}-T_{1}(z)-T_{1}(x_{\min})}^{t-2T_{2}-T_{1}(z)-T_{1}(x_{\min})} \\ g\left(x,T_{1}^{-1}\left[t-2T_{2}-T_{1}(z)-\tau\right],z\right) \\ P\left(\tau,z\right) \left|\left(T_{1}^{-1}\right)'\left[t-2T_{2}-T_{1}(z)-\tau\right]\right| d\tau dz. \end{split}$$

Since the function  $g\left(x,T_{1}^{-1}\left[t-2T_{2}-T_{1}\left(z\right)-\tau\right],z\right)$  is in  $L^{1}$  of the four variables (t,x,y,z), it can be approximated by a sequence  $\{g_{n}\left(t,x,y,z\right)\}_{n\in\mathbb{N}}$  of functions,

continuous in those variables. Now, we denote  $G_n$  the operator associated to one of such continuous functions:

$$(G_n P)(t, x) = 4 \int_{x_{\min}}^{x^*} \int_{t-2T_2-T_1(z)-T_1(x^*)}^{t-2T_2-T_1(z)-T_1(x^*)} g_n(x, T_1^{-1}[t-2T_2-T_1(z)-\tau], z)$$

$$P(\tau, z) \left| \left(T_1^{-1}\right)'[t-2T_2-T_1(z)-\tau] \right| d\tau dz.$$

The following lemma can then be proved:

**Lemma 4.3.** The operator  $G_n$  is compact from  $L^{\infty}$  to  $\mathcal{C}([2\rho, T] \times [x_{\min}, x^*])$ ,  $T \geq 2\rho$ .

proof. Since  $g_n$  is continuous, bounded sets of  $L^{\infty}$  are mapped by  $G_n$  into uniformly bounded and equicontinuous sets of the space  $\mathcal{C}\left([A,B]\times[x_{\min},x^*]\right)$ . Therefore, by applying the Arzelà-Ascoli theorem, we conclude that bounded sets of  $L^{\infty}$  are mapped by  $G_n$  into relatively compact sets of  $\mathcal{C}\left([A,B]\times[x_{\min},x^*]\right)$ , which proves that  $G_n$  is compact. The interval [A,B] has to be chosen in a way such that t is big enough to bypass any delay, that is:

$$A = 2\rho$$
$$B \ge 2\rho.$$

Corollary 1. The operator  $G_n$  is compact from  $L^{\infty}$  to  $L^1$ .

Now, if we let n go to infinity, we conclude that the sequence  $\{G_n\}_{n\in\mathbb{N}}$  of operators converges to  $K^2$  in the operator norm:

$$\|K^2 - G_n\|_{\mathcal{L}(L^1, L^\infty)} \le \|g - g_n\|_{L^1} \underset{n \to \infty}{\longrightarrow} 0.$$

Therefore,  $K^2$  is compact from  $L^{\infty}$  to  $L^1$ .

Let us now consider  $K^4$ , which acts on  $L^1$ . We know that  $K^2$  maps  $L^1$  into  $L^{\infty}$  and another application of  $K^2$  maps  $L^{\infty}$  into  $L^1$  compactly. Then we can state the following result:

**Proposition 9.** the operator  $K^4$ , defined as:

$$\left(K^{4}P\right)\left(t,x\right) = 4\int_{x_{\min}}^{x^{*}} \widetilde{f}\left(x,y\right)$$
$$\int_{x_{\min}}^{x^{*}} \widetilde{f}\left(y,z\right) \left\{K^{2}P\left[t - 2T_{2} - T_{1}\left(y\right) - T_{1}\left(z\right),z\right]dz\right\}dy$$

is compact from  $L^1$  to  $L^1$ .

Now, for  $t \geq 2\rho$ , we can write P(t,x) in the following way:

$$\begin{split} P\left(t,x\right) &= 4 \int_{x_{\min}}^{x^*} \widetilde{f}\left(x,y\right) \\ &\int_{x_{\min}}^{x_{thr}} \widetilde{f}\left(y,z\right) \left\{ P\left[t - 2T_2 - T_1\left(y\right) - T_1\left(z\right),z\right] dz \right\} dy + \\ &4 \int_{x^*}^{x_{thr}} \widetilde{f}\left(x,y\right) \\ &\int_{x_{\min}}^{x_{thr}} \widetilde{f}\left(y,z\right) \left\{ P\left[t - 2T_2 - T_{\min} - T_1\left(z\right),z\right] dz \right\} dy. \end{split}$$

Let us decompose  $(x_{\min}, x_{thr})$  into the union of  $(x_{\min}, x^*)$  and  $(x^*, x_{thr})$  in the second integral of the second term on the right hand side. We will obtain again a compact part, the one corresponding to the integration interval  $(x_{\min}, x^*)$ , and a noncompact part.

Now, for  $t \geq 5\rho + n (T_2 + T_{\min})$ , we obtain:

$$P(t,x) = \overline{P}(t,x) + \widetilde{P}(t,x),$$

where the operator  $P_0 \longrightarrow \overline{P}$  is compact and  $\widetilde{P}(t,x)$  is given by:

$$\begin{split} \widetilde{P}\left(t,x\right) &= 2^{n} \int_{\left(x^{*},x_{thr}\right)^{n} \times \left(x_{\min},x_{thr}\right)} \\ \widetilde{f}\left(x,y\right) \widetilde{f}\left(y,z_{1}\right) ... \widetilde{f}\left(z_{n-1},z_{n}\right) \\ P\left(t-2nT_{2}-2\left(n-1\right)T_{\min}-T_{1}\left(z_{n}\right),z_{n}\right) dz_{n} ... dz_{1} dy. \end{split}$$

If we define:

$$(G(t) P_0)(s,x) = \overline{P}(t+s,x)$$
  
 $(L(t) P_0)(s,x) = \widetilde{P}(t+s,x)$ 

then we can prove that  $G(t): X \longrightarrow X$  is compact and that  $\exists C, \varepsilon > 0: ||L(t)|| \le C \exp[(\lambda^* - \varepsilon)t]$ . This results in the following

**Proposition 10.** Assume that properties 2. for the function  $T_c$  and 7. for the function f are verified. Then, there exist  $\varepsilon > 0$ ,  $C \ge 0$  such that, for t large enough, T(t) = G(t) + L(t) where G(t) is compact and  $||L(t)|| \le C \exp[(\lambda^* - \varepsilon)t]$ .

*proof.* The proof is obtained by comparing operators  $L_{\lambda}$  (defined in (31)) and L(t). We introduce the operator  $\mathcal{L}$  defined by

$$\mathcal{L}\mu\left(x\right) = 2\int_{\left(x^{*}, x_{thr}\right)} \widetilde{f}\left(x, y\right) \mu\left(y\right) dy,$$

on the space  $L^1(]x^*, x_{thr}[)$ .

In terms of  $\mathcal{L}$  formula (39) reads:

$$\widetilde{P}\left(t,x\right) = \\ \mathcal{L}^{n} \int_{x_{\min}}^{x_{thr}} \widetilde{f}\left(x,z\right) P\left(t - 2nT_{2} - 2\left(n - 1\right)T_{\min} - T_{1}\left(z\right),z\right) dz,$$

from which we deduce that

$$\|L(t)\varphi\|_{X} \le C \|\mathcal{L}^{n}\|_{L^{1}(]x^{*},x_{thr}[)} \|\varphi\|_{X}$$

$$\tag{39}$$

for  $4\rho + n(T_{\min} + T_2) \le t \le 5\rho + n(T_{\min} + T_2)$ .

To estimate the quantity  $\|\mathcal{L}^n\|_{L^1(]x^*,x_{thr}[)}$  amounts to estimating the spectral radius r of the operator  $\mathcal{L}$ . It can be shown (see [6]) that, under the hypothesis that  $\widetilde{f}(x,y)$  is positive at each point (x,y) in  $[x^*,x_{thr}]\times[x^*,x_{thr}]$ , the following inequality holds:

$$r < \exp\left(\left(T_{\min} + T_2\right)\lambda^*\right). \tag{40}$$

Now, inequality (39) holds when t is of the order of  $n(T_{\min} + T_2)$ , for t large. Therefore,  $\|L(t)\|^{\frac{1}{t}}$  is equivalent to  $\|L(t)\|^{\frac{1}{n(T_{\min}+T_2)}}$ . If we apply the same reasoning to  $\|\mathcal{L}^n\|_{L^1(]x^*,x_{thr}[)}$ , we obtain, when n goes to infinity, a term which is bounded by  $r^{\frac{1}{(T_{\min}+T_2)}}$ . From (40), it follows that the latter is less than  $\exp \lambda^*$  and, going back to (39), we can conclude that there exists  $\varepsilon > 0$ , such that  $\|L(t)\| \le C \exp[(\lambda^* - \varepsilon) t]$ .

5. Conclusion. A way to connect cell cycle and proliferation could be that of describing the evolution of the content of some selected proteins until the end of the cell cycle, with the assumption of unequal subdivision among daughter cells of the final protein contents. We have considered a linear model connecting the behavior at individual cell level to the behavior at population level. The model describe a cell population structured by the cell age and the amount of Cyclin E/CDK2 with two compartments: cells in the  $G_1$  phase and cells in the remainder of the cell cycle  $(S-G_2-M)$ . We have also taken into account the variability in the timing of  $G_1/S$ transition which dependent on the initial amount of Cyclin E/CDK2. The analysis we have carried out for the model allowed us to make simplifying assumptions, such as the homogeneity of the extra-cellular milieu, and the absence of mortality or quiescence since we have considered cell lines growing in vitro under optimal conditions. We have proved that the population P(t,x) of cells which start a new cell cycle is such that  $P(t,x) - C' \exp(\lambda^* t) \mu^*(x) = o[\exp(\lambda^* t)]$  when  $t \to \infty$ . In this first step, we have used only one cyclin/CDK complex but one can hope to give a model more realistic and take into account the other cyclin/CDK complexes which are important for cell cycle progression. We will keep the  $G_1$  phase and separate the remainder of the cell cycle into the S,  $G_2$  and M phases. We hope also to add drug effects since cell lines are used to test many anti-tumor treatments. The comparison between data related to treated cell populations and the kinetic data of unperturbed cell lines are an important source of information about cytostatic and cytotoxic activity of a drug against cancers and are used for the first tests of new drugs. However, maintaining the proper number of cells in any tissue requires a delicate balance between quiescence, differentiation, cell cycle length, cellular production by division and cell loss by necrosis or apoptosis ([19], [49]) and an imbalance between these processes can result in tumoral tissue. The representation of these effects on the cell cycle using modeling approach with linear or nonlinear parameters can be helpful to understand cell cycle regulation and cell proliferation.

## REFERENCES

- [1] B. D. Aguda and Y. Tang, The kinetic origins of the restriction point in the mammalian cell cycle, Cell Prolif., **32** (1999), 321–335.
- [2] O. Arino and M. Kimmel, Asymptotic analysis of a cell cycle model based on unequal division, Siam J. Appl. Math., 47 (1987), 128–145.
- [3] O. Arino and M. Kimmel, Asymptotic behavior of a nonlinear functional-integral equation of cell kinetics with unequal division, J. Math. Biol., 27 (1989), 341–354.
- [4] O. Arino and M. Kimmel, Asymptotic behavior of nonlinear semigroup describing a model of selective cell growth regulation, J. Math. Biol., 29 (1991), 283–314.

- [5] O. Arino and M. Kimmel, Comparison of approaches to modeling of cell population dynamics, SIAM J. Appl. Math., 53 (1993), 1480–1504.
- [6] O. Arino, Some spectral properties for the asymptotic behavior of semigroups connected to population dynamics, SIAM Review, 34 (1992), 445–476.
- [7] F. Bekkal Brikci, et al., Analysis of a molecular structured population model with possible polynomial growth for the cell division cycle, Mathematical and Computer Modelling, 47, (2008), 699–713.
- [8] G. Chiorino, "Modelisation Mathematique et etude du Couplage Proliferation-Cycle Cellulaire," these de doctorat, Université de Pau et des pays de l'Adour Dec 2000.
- [9] G. Chiorino and M. Lupi, Variability in the timing of G<sub>1</sub>/S transition, Mathematical Biosciences, 177 (2002), 85–101.
- [10] S. Cooper, C. Yu and J. A. Shayman, Revisiting retinoblastoma protein phosphorylation during the mammalian cell cycle, CMLS Cell. Mol. Life Sci., 58 (2001), 580–595.
- [11] Z. Darzynkiewicz, "Metabolic and Kinetic Compartments of the Cell Cycle Distinguished by Multiparameter Flow Cytometry, from Growth, Cancer and the Cell Cycle," Skehan P. and Friedman S.J. eds., Humana Press (1984).
- [12] R. J. Deshaies and T. Jacks, Cell multiplication, Peering in and peering out: Regulation of and by the cell cycle, Curr Opin Cell Biol. Dec, 11 (1999), 705–707.
- [13] D. Donjerkovic and D. W. Scott, Regulation of the G<sub>1</sub> phase of the mammalian cell cycle, Cell Res. Mar., 10 (2000), 1–16.
- [14] Y. Geng, E. N. Eaton, M. Picon, J. M. Roberts, A. S. Lundberg, A. Gifford, C. Sardet and R. A. Weinberg, Regulation of Cyclin E transcription by E2F's and Retinoblastoma protein, Oncogen, 12 (1996), 1173–1180.
- [15] A. Goldbeter, A minimal cascade model for the mitotic oscillator involving cyclin and cdc2 kinase, Proc. Natl. Acad. Sci. U S A., Oct 15, 88 (1991), 9107–11.
- [16] L. H. Hartwell and M. B. Kastan, Cell cycle control and cancer, Science Dec 16, 266 (1994), 1821–1828.
- [17] V. Hatzimanikatis, K. H. Lee and J. E. Bailey, A mathematical description of regulation of the G<sub>1</sub>-S transition of the Mammalian cell cycle, Biotechnol Bioeng. Dec 20, 65 (1999), 631–637.
- [18] G. Juan, L. Xiaoyu and Z. Darzynkiewicz, Phosphorylation of retinoblastoma protein assayed in individual HL-60 cells during their proliferation and differentiation, Exp. Cell. Res. 244 (1998), 83–92.
- [19] K. L. King and J. A. Cidlowski, Cell cycle regulation and apoptosis, Annu. Rev. Physiol., 60 (1998), 601–617.
- [20] M. Kimmel, Z. Darzynkiewicz, O. Arino and F. Traganos, Analysis of a cell cycle model based on unequal division of metabolic constituents to daughter cells during cytokinesis, J. Theor. Biol., 110 (1984), 637–664.
- [21] A. Koff, A. Giordano, D. Desai, K. Yamashita, J. W. Harper, S. Elledge, T. Nishimoto, D. O. Morgan, B. R. Franza and J. M. Roberts, Formation and activation of a Cyclin E-cdk2 complex during the G<sub>1</sub> phase of the human cell cycle, Science, Sep 18, 257 (1992), 1689–1694.
- [22] K. W. Kohn, Functional capabilities of molecular network components controlling the mammalian G<sub>1</sub>/S cell cycle phase transition, Oncogene. Feb 26, 16 (1998), 1065–1075.
- [23] K. W. Kohn, Molecular interaction map of the mammalian cell cycle control and DNA repair systems, Mol Biol Cell, Aug 10, (1999), 2703–2734.
- [24] A. Lasota and M. C. Mackey, Globally asymptotic properties of proliferation cell populations, J. Math. Biol, 19 (1984), 43–62.
- [25] G. Leone, J. DeGregori, L. Jakoi, J. G. Cook and J. R. Nevins, Collaborative role of E2F transcriptional activity and G<sub>1</sub> cyclin-dependent kinase activity in the induction of S phase, Proc. Natl. Acad. Sci. USA, 96 (1999), 6626–6631.
- [26] J. Lukas, T. Herzinger, K. Hansen, M. C. Moroni, D. Resnitzki, K. Helin, S. Reed and J. Bartek, CyclinE induced S phase without activation of the pRb/E2F pathway, Gen. Dev., 11 (1997), 1479–1492.
- [27] J. A. J. Metz and O. Diekmann, "The Dynamics of Physiologically Structured Populations," Lecture Notes in Biomath, 68, Springer-Verlag, Berlin, 1986.
- [28] K. I. Nakayama, S. Hatakeyama and K. Nakayama, Regulation of the cell cycle at the G<sub>1</sub>-S transition by proteolysis of Cyclin E and p27Kip1, Biochem Biophys Res Commun., Apr 13, 282 (2001), 853–860.
- [29] S. Mittnacht, Control of pRb phosphorylation, Curr. Op. Gen. Dev., 8 (1998), 21–27.

- [30] R. Norel and Z. Agur, A model for the adjustment of the mitotic clock by cyclin and MPF levels, Science. Mar 1, 251 (1991), 1076–1078.
- [31] B. Novak, A. Csikasz-Nagy, B. Gyorffy, K. Chen and J. J. Tyson, Mathematical model of the fission yeast cell cycle with checkpoint controls at the G<sub>1</sub>/S, G<sub>2</sub>/M and metaphase/anaphase transitions, Biophys. Chem., 72 (1998), 185–200.
- [32] M. N. Obeyesekere, S. L. Tucker and S. O. Zimmerman, Mathematical models for the cellular concentrations of cyclin and MPF, Biochem Biophys Res Commun., Apr 30, 184 (1992), 782–789.
- [33] M. N. Obeyesekere, J. R. Herbert and S. O. Zimmerman, A model of the G<sub>1</sub> phase of the cell cycle incorporating Cyclin E/CDK2 complex and retinoblastoma protein, Oncogene, Sep 21, 11 (1995), 1199–1205.
- [34] M. N. Obeyesekere and E. S. Tecarro, Analysis of a model of the Mammalian Cell cycle's G<sub>1</sub> phase, Nonlinear Analysis and Applications, Real World Applications, 4 (2003), 87–107.
- [35] Z. Qu, J. N. Weiss and W. R. MacLella, Regulation of the mammalian cell cycle: a model of the G<sub>1</sub>-to-S transition, Am. J. Physiol. Cell. Physiol., 284 (2003), 349–364.
- [36] R. j. Sheaff, M. Groudine, M. Gordon, J. M. Roberts and B. E. Clurman, Cyclin E-CDK2 is a regulator of p27<sup>KIP1</sup>, Genes & Development, 11 (1997), 1464–1478.
- [37] C. J. Sherr, Cancer cell cycles, Science Dec 6, 274 (1996), 1672–1677.
- [38] C. J. Sherr and J. M. Roberts, CDK inhibitors: positive and negative regulators of G<sub>1</sub>-phase progression, Genes Dev. Jun 15, 13 (1999), 1501–1512.
- [39] J. A. Simth and L. Martin, Do cells cycle? Proc. Nat. Acad. Sci. USA, 70 (1973), 1263–1267.
- [40] C. H. Spruck, K. A. Won and S. I. Reed, Deregulated Cyclin E induces chromosome instability, Nature, 16 (1999), 297–300.
- [41] M. Swat, A. Kel and H. Herzel, Bifurcation analysis of the regulatory modules of the mammalian G<sub>1</sub>/S transition, Bioinformatics, 20 (2004), 1506–1511.
- [42] C. D. Thron, Mathematical analysis of a model of the mitotic clock, Science, Oct 4, 254 (1991), 122–123.
- [43] J. Slingerland and M. Pagano, Regulation of the cdk inhibitor p27 and its deregulation in cancer, J. Cell Physiol., Apr., 183 (2000), 10–17.
- [44] L. Spinelli, A. Torricelli, P. Ubezio and B. Basse, Modelling the balance between quiescence and cell death in normal and tumour cell populations, submitted, 202 (2006), 349–370.
- [45] D. Tomasoni, M. Lupi, F. Bekkal Brikci and P. Ubezio, Timing the changes of cyclin E cell content in G1 in exponentially growing cells, Experimental Cell Research, 288 (2003), 158–167.
- [46] H. Toyoshima and T. Hunter, A novel inhibitor of G<sub>1</sub> cyclin-Cdk protein kinase activity is related to p21, Cell, Jul 15, 78 (1994), 67–74.
- [47] J. J. Tyson and B. Novak, Regulation of the eukaryotic cell cycle: molecular antagonism, hysteresis, and irreversible transitions, J. Theor. Biol. May 21, 210 (2001), 249–263,
- [48] J. Val and J. Tyson, A purely deterministic model for the population dynamics of budding yeast, Advances in Mathematical Population Dynamics - Molecules, Cells and Man, Arino, O., Axelrod, D. & Kimmel, M. eds., World Scientific, (1997).
- [49] C. J. F. Van Noorden, L. C. Meade-Tollin and F. T. Bosman, *Metastasis*, American Scientist, 86 (1998), 130–141.
- [50] S. Wiggins, "Introduction to Applied Nonlinear Dynamical Systems and Chaos," Texts in Applied Mathematics, Springer-Verlag New York, vol. 2, 1990.
- [51] G. F. Webb, "Theory of nonlinear Age-dependent Population Dynamics," Monographs and textbooks in pure and applied mathematics, Marcel Dekker, New York, 1985.

Received February 2008; revised November 2008.

E-mail address: b\_fadia@yahoo.fr

 $E ext{-}mail\ address: giovanna.chiorino@gmail.com}$ 

 $E ext{-}mail\ address: boushaba@iastate.edu}$