



Review

The divergence between the virus and cellular oxidative stress as separate environmental agents that trigger autoimmunity originates from their different procedural mechanisms of activating the same molecular entity: the transcription factor NF-kappa B

Norbert O. Temajo * and Neville Howard

Institute of Endocrinology and Diabetes, The Children's Hospital at Westmead, Sydney, NSW, Australia

* **Correspondence:** Email: temajo@grapevine.net.au; Tel: +6-126-262-7619; Fax: +6-129-845-3170.

Abstract: To happen, autoimmunity in man requires triggering by environmental factors: the viruses and cellular stress, in genetically primed individuals. The viruses and stress are operatives in this scene as stimuli for the activation of the transcription factor (TF), NF-KB. NF-KB is unusually activated: the viral activation occurs via serine residues-phosphorylation by IKK β and IKK ϵ , while the activation by oxidative stress occurs via tyrosine phosphorylation of IKK α . The phosphorylation of particular amino acid residues of a given protein molecule modulates that protein's polymorphic conformations, appropriately. For a TF, a given conformation influences its choice of cognate DNA sequence recognition as well as its interactions with neighboring molecules. The TF NF-KB performs a battery of regulatory functions. Because it is variously phosphorylated as seen above this implies that NF-KB is capable of assuming a multitude of polymorphic conformations that we refer to as "*derivative isoforms*". Thus the virus activation of NF-KB occurring by phosphorylation at serines S536 and S468 is observed to generate the *isoforms* with the potential to activate the transcription of viral latency genes, hereby installing a latent infection. But oxidative stress activation of NF-KB occurs via phosphorylation of Tyr42 of IKK α and this yields *isoforms* that activate the transcription of replication and transcription activator (RTA), the master lytic switch, which thereby abrogates the latency. The steps involved are that the stress-activated NF-KB and the viral miRNAs conjoin in a regulatory circuitry identified as feedback and feed-forward network motifs that co-accomplish the switching on of RTA which in turn activates the transcription of the immediate early

genes BZLF1 and BRLF1. These two latter genes together mobilize the expression of the set of lytic genes, resulting in a lytic cascade and consequentially set in trend the viral journey to the triggering of autoimmunity (Figure 1).

Keywords: nuclear factor (NF-KB); replication and transcription activator (RTA); autoimmunity; environmental factors (viruses, oxidative stress); viral latency; microRNAs (miRNAs)

1. Introduction

Recent advances in genetic and environmental contributions to autoimmunity suggest that interaction between genetic elements and epigenetic changes caused by environmental agents may be responsible for inducing autoimmune diseases (ADs). Autoimmunity arises when immune responses mounted in the host are directed against self-components as a result of the loss of self-tolerance. ADs may be either tissues-specific where unique tissue-specific antigens are targeted, or systemic in which multiple tissues are affected and a variety of apparently ubiquitously expressed autoantigens are targeted [1,2] (Table 1). The affliction of autoimmunity is believed to develop when genetically

Table 1. Examples of selected human autoimmune diseases (adapted from Ray S et al. [1]).

Organ specific autoimmune diseases		
Organ	Disease(s)	Self-Antigen
Adrenal cells	Addison's disease	Cytochrome P-450 antigens
Red blood cells	Autoimmune haemolytic anaemia	Red blood cell membrane proteins
Platelets	Idiopathic thrombocytopenic purpura	Platelet antigens
Stomach	Pernicious anaemia	Gastric parietal cell antigens
Small bowel	Coeliac sprue (gluten enteropathy)	Transglutaminase
Thyroid	Hashimoto's thyroiditis	Thyroid cell antigens
	Grave's disease	Thyroid-stimulating hormone receptor
Muscle	Myasthenia gravis	Acetylcholine receptors
Pancreatic islets	Type 1 diabetes	Beta cell antigens
Hepatocytes	Autoimmune hepatitis	Hepatocyte antigens (cytochrome P-450 2D6)
Bile duct cells	Primary biliary cirrhosis	Intra-hepatic bile duct
Heart	Rheumatic heart disease	Myocardial antigens
Kidney/lung	Goodpasture's syndrome	Basement membrane antigens
Systemic autoimmune diseases		
	Disease(s)	Self-Antigen
	Ankylosing spondylitis	Vertebrae
	Multiple sclerosis	Brain or white matter
	Rheumatoid arthritis	Connective tissue, IgG
	Systemic lupus erythematosus	DNA nuclear protein RBC and platelet membranes
	Scleroderma	Nuclei, heart, lungs, gastrointestinal tract, kidney
	Sjogren's syndrome	Salivary gland, liver, kidney, thyroid

predisposed individuals encounter environmental agents that trigger it. The environmental agents that are revered in this respect are stress and the viruses [3–15], which are operatives in this scene as stimuli for the activation of the nuclear factor (NF-KB), a transcription factor which specifically binds promoter DNA to activate target gene expression.

1.1. *NF-KB*

The transcription factor NF-KB is involved in the regulation of a large number of genes that control various aspects of the immune and inflammatory responses. According to Zhu et al. [16], NF-KB is a family of structurally related transcription factors. In mammals the NF-KB family consists of five members: NF-KB1 (p105/50), NF-KB2 (p100/52), RelA (p65), RelB, and c-Rel. They all have a structurally conserved N-terminal 300-amino-acid Rel homology domain (RHD), which contains sequences responsible for dimerization, nuclear translocation and DNA binding. Only RelA, RelB and c-Rel have a transactivation domain, which is non-homologous and located at the C-termini of the proteins. The other two NF-KB proteins, NF-KB1 and NF-KB2, lack a transactivation domain but instead contain seven ankyrin repeats, a 33-amino-acid characteristic motif of the inhibitor of NF-KB, (IKB), that mediates protein-protein interactions. The five NF-KB proteins above can form 15 transcription factors through homo- or hetero-dimerization [17]: the main activated forms being RelA: p50 and RelB: p52. These NF-KB family members play critical roles in a wide variety of biological processes, including immune and inflammatory responses, development, cell growth and apoptosis. In unstimulated cells, the NF-KB dimers are sequestered in the cytoplasm by a family of inhibitors called IKBs (IKB α , IKB β , IKB ϵ and IKB γ , IKB δ , IKB ζ and Bcl-3), which are proteins that contain multiple copies of the sequence ankyrin repeats. By virtue of their ankyrin repeat domains, the IKB proteins mask the nuclear localization signals of NF-KB proteins and keep them sequestered in an inactive state in the cytoplasm [18]. The NF-KB dimer becomes activated through degradation of the IKB following phosphorylation of specific serines in the IKB proteins by the IKB- kinase complex (IKK), leading to their ubiquitination and thus allowing the liberation and nuclear translocation of NF-KB dimers to induce gene expression.

1.2. *Activation of NF-KB*

The NF-KB is recognized as a master transcription factor that responds to diverse stimuli by activating the expression of stress-response genes. Multiple signals, including cytokines, growth factors, engagement of the T cell receptor, bacterial and viral products, various forms of radiation and the oxidative stress, induce NF-KB transcriptional activity [19,20,21]. We are particularly interested in, and will limit ourselves to, the mechanism by which NF-KB is activated by oxidative stress and the viruses, the latter being represented by the gamma herpesviruses KSHV and EBV.

1.2.1. Oxidative stress-activation of NF-KB

In respiring cells, a small amount of consumed oxygen is reduced in a specific way, yielding a variety of highly reactive chemical entities collectively called reactive oxygen species (ROS) or reactive oxidative intermediates (ROIs) and include hydrogen peroxide (H₂O₂), nitric oxide radical (NO), superoxide anion (O₂⁻) and hydroxyl radical (OH⁻). A state of moderately increased

levels of intracellular ROS is referred to as cellular oxidative stress [19,22]. Oxidative stress is caused by an imbalance between the production of oxidants and/or decreased ability to detoxify those oxidants. ROS activate NF- κ B by various mechanisms [23,24]. Studies have been made which suggest that I κ B α phosphorylation and degradation might be the step that is responsive to oxidative stress. NF- κ B activation has been suggested to be regulated by the levels of oxidants inside cells [22]. Thus it is observed that H₂O₂, a ROS, regulates NF- κ B activation through alternative phosphorylation of I κ B α . While typically I κ B α is usually phosphorylated on serines 32 and 36, which leads to its ubiquitination and degradation, ROS H₂O₂ affects the phosphorylation of I κ B α on tyrosine Tyr42 or other tyrosine residues, and I κ B α may or may not be degraded as part of the process [25–29]. IKK is not required in this case, and I κ B α phosphorylation may be mediated by casein Kinase II as well as a tyrosine kinase. Degradation of I κ B α may not be necessary in this case because Tyr42-phosphorylated I κ B α is bound by the SH₂ domains of p85 α regulatory subunit of PI3K, thus unmasking NF- κ B and allowing it to translocate to the nucleus [30,31,32]. PI3K as well as c-Src have been implicated in alternative phosphorylation of I κ B α .

1.2.2. Viral activation of NF- κ B, installment of viral latency

NF- κ B activation is imperative for latent infection of gamma herpesviruses. KSHV infection activates IKK β and IKK ϵ to enable host NF- κ B activation and KSHV latent infection. Biochemical and genetic experiments identified RelA as a key player downstream of IKK β and IKK ϵ , i.e., IKK β and IKK ϵ were essential for phosphorylation of serines S536 and S468 of RelA, respectively [33]. That the expression of the phosphorylation-resistant RelA^{S536A} increased KSHV lytic genes expression and impaired latent infection is further seen as completing evidence. NF- κ B activation is coordinated by IKK β and IKK ϵ , which sequentially phosphorylate RelA in a site-specific manner to enable latent infection after KSHV *de novo* infection. As a further evidential support, knockdown of IKK β and IKK ϵ impairs NF- κ B activation and elevates KSHV lytic genes expression resulting in abrogation of the latent infection. Please recall here that RelA is one of the five members of NF- κ B family that is the most abundantly and ubiquitously expressed and is the transcriptionally active subunit of the predominant RelA: p50 dimer. Post translational modifications of the RelA subunit, e.g., phosphorylation and acetylation, are important means to regulate NF- κ B-dependent gene expression [34–37]. Activation of NF- κ B through viral infection results in the transactivation of the NF- κ B site-containing viral promoter, resulting in installment of latency [38].

1.3. Viral latency

In latent infection, the full viral genome is retained in the host, but its expression is dramatically restricted, such that few viral antigens and no viral particles are produced. To qualify as latency, this cryptic form of infection must display two additional properties: persistence and reversibility, i.e., the capacity of the genome, under the appropriate circumstances (e.g., oxidative stress) to be fully reactivated with the production of infectious progeny, (the lytic replication), is the key requirement of latency. It is the property of reversibility that allows latent infections to avoid abortive infection, or a dead end, and instead become an effective mechanism of viral persistence. Efficient establishment of latency allows the viral genome to persist despite the host immune responses to many viral antigens, and in the face of other potentially adverse signals in the microenvironment.

When environmental conditions warrant, appropriate signals can trigger the full repertoire of viral gene expression, allowing viral production and spread to resume.

1.3.1. Maintenance of latency

The herpesviruses, alpha, beta and gamma subfamilies are the viruses capable of true latency as described above. In fact all herpesviruses share a propensity to establish latent infection [39,40]. The herpesviruses HBV and KSHV encode multiple viral miRNAs which mimic cellular miRNAs and modulate cellular and viral gene expression [41–44]. Some of the viral miRNAs have been implicated in maintaining latent infection and altering the balance between latent/lytic infection [45,46]. The first study showing that miRNAs can be encoded in the viral genome was published in 2004 [47]. The unique ability of the herpesviruses to establish long term latent infections means that these viruses need to block protective host innate or adaptive immune responses over the long term while minimizing the expression of potentially antigenic viral proteins, and this is accomplished by the viral miRNAs. Employing quantitative algorithms prediction, the authors, Grey et al. and Lu et al. [48,49] determined that the herpesviruses employ viral miRNAs to suppress expression of their own genes, including their immediate early genes. These authors concluded that these viruses use viral miRNAs-mediated suppression of immediate early genes as part of their strategy to enter and maintain latency. The lytic cascade is reinitiated with the expression of the immediate early genes (RTA, BZLF1 and BRLF1). Multiple viral miRNAs are transcribed during latency. The KSHV encoded miRNAs (miR-K12-9-5p and miR-12-7-5p) have been shown to directly regulate and inhibit the transcription of the master lytic switch protein, the viral immediate early gene known as the replication and transcription activator (RTA), and in this way they contribute to the maintenance of viral latency. They target the expression of RTA at the translational level [50,51,52]. Fang Lu et al. observed that EBV-coded miR-155 attenuates NF-KB signaling and stabilizes latent virus persistence [41]. miRNA-155 is important for the stable maintenance of EBV genomes during latent infection. Several latency genes have been observed to influence the activation of NF-KB which turns out to be important in the maintenance of latency. EBV encoded latency membrane protein 1 (LMP1) is a potent activator of NF-KB signaling pathways and is essential for EBV immortalization of B lymphocytes. Finally, the observations by Lei et al. [53] established that the viral activation of NF-KB is essential for the establishment of latency.

1.3.2. Abrogation of latency (the activation of the lytic, replicative phase of infection)

EBV lytic genes are expressed in a temporally regulated cascade. Following the activation, transcription and expression of the lytic master switch RTA, the switch from the latent to the replicative form of EBV infection is mediated by the two immediate early proteins BZLF1 and BRLF1, which are transcription factors that together activate the entire lytic viral cascade of gene expression, ultimately resulting in the production of infectious viral particles [54]. In natural viral life cycle, the differentiation of the cells triggers the abrogation of latency and entrance into the lytic phase of infection. In the situations leading to virus-induced autoimmunity, however, it is cellular stress, in particular for our purpose, oxidative stress that triggers latency termination [5].

2. The Hypothesis

The switch between latency and lytic replication is regulated by epigenetic factors: (i) hypomethylation of the promoter of replication and transcription activator, RTA (which, as said above, is essential for the lytic switch) leads to viral reactivation; (ii) histone acetylation induces viral replication by influencing protein-protein associations and transcription factor binding [55]; and (iii) it is the general and established situation that post-transcriptional gene regulation of mRNA turnover and translation is mediated by microRNAs and RNA-binding proteins in events identified as network motifs [56–60]. The network motifs in the images of feedback and feed-forward mechanisms or loops [61] are defined as small gene regulating circuits that occur more often than can be expected by chance. Feed-back motifs are prominent throughout biology: they can be identified when protein-protein interactions are combined with transcriptional interactions [62] or when transcriptional interactions and miRNA-mediated post-transcriptional interactions are involved. We hypothesize that this latter scenario and mode of feedback motif occurs with stress-activated NF-KB (a transcription factor) and viral miRNAs: the ensemble being depicted as NF-KB \leftrightarrow miRNA, signifying a reciprocal regulation between the viral miRNA and NF-KB in which NF-KB regulates viral miRNA and is itself regulated by the same miRNA. NF-KB gets directly involved in up-regulation and down-regulation of miRNAs [63]. We suggest that the relationship above induces a down-regulation of the viral miRNA with the consequence that the viral miRNA gets sequestered (titrated out) and neutralized by the RTA site with which it interacts through a process termed “sponging”. Equally interacting with the RTA site is the stress-activated NF-KB. But the down-regulation of the viral miRNA by NF-KB in the feed-back circuit (formula) above accentuates the viral miRNA sponging, making it (the viral miRNA) no longer in excess but reduced to below some minimal proportionate value. This frees the RTA site for the transcription factor NF-KB interaction and it results in the activation and transcription of RTA: we say this event spells switching on of the master lytic switch, if you will!

In the situation of feed-forward circuitry, a regulator X regulates the expression of a target Z via a direct or indirect path through a regulator Y, the whole setting represented as X/Z/Y, which in our particular case is translated as “stress-activated NF-KB/RTA/viral miRNA”. This feed-forward loop reveals a mechanism through which stress-activated NF-KB simultaneously activates RTA transcription and limits its translation through activation of the viral miRNA.

In summary, NF-KB is unusually activated: its viral activation via serine residues phosphorylation by IKK β and IKK ϵ leads to viral latency, while activation by oxidative stress (ROS) occurs via tyrosine phosphorylation of IKK α which then complexes with NF-KB and leads to abrogation of the viral latency. To facilitate understanding of the above we abbreviate that “viral activation of NF-KB leads to viral latency; while stress activation of the same molecular entity, the NF-KB, leads instead to abrogation of the latency”. Thus NF-KB emerges as the converging focus between stress and the viruses in their efforts as separate environmental agents that trigger autoimmunity. The phosphorylation of particular amino acid residues of a protein molecule modulates that protein’s polymorphic conformations, appropriately. For a transcription factor, TF, a given conformation influences its choice of cognate DNA sequence recognition as well as its interaction with a neighboring TF or other proteins and molecules. The transcription factor, NF-KB, in particular is reputed to be involved in a battery of regulatory functions [23,64]. We interpret this to mean that as a TF that is variously and differentially phosphorylated, NF-KB is capable of assuming

or exhibiting a multitude of polymorphic conformations that we choose to call them “*derivative isoforms*”. Thus the virus activation of NF- κ B that occurs via its phosphorylation by IKK β and IKK ϵ at serines S536 and S468 generates the *derivative isoforms* with the potential to activate the transcription of viral latency genes thereby installing and maintaining a latent infection. On the other hand oxidative stress-activation of NF- κ B occurs via complexation with IKB α that is phosphorylated at tyrosine Tyr42 and yields *derivative isoforms* with the attributes that activate the transcription of RTA, the master lytic switch, thereby precipitating abrogation of the viral latency and engendering the lytic cycle phase. The above two alternative involvements of NF- κ B portray its versatility and sophistication and confirms the difference in mechanism between viral- and stress-activations of NF- κ B. The revelation of the feedback and feedforward network motifs cited above represents evidence of regulatory circuits involving stress-activated NF- κ B and viral miRNAs: the feedback mechanism co-accomplishes with the feed-forward loop, in a relation possibly reverberative of a cooperativity, the expression and switching on of RTA, the master lytic switch. This expressed RTA in turn activates the transcription of the immediate early genes BZLF1 and BRLF1: the two transcription factors that mobilize the expression of the whole set of lytic genes, resulting in the procreation of the lytic cascade. Interpret this simply as the activation of NF- κ B by stress materializes in a dual prong: feedback and feedforward loops that cooperate in the switching on of RTA and thus abrogate the viral latency, consequentially setting in trend or motion the journey of the virus to the triggering of autoimmunity (Figure 1) [65].

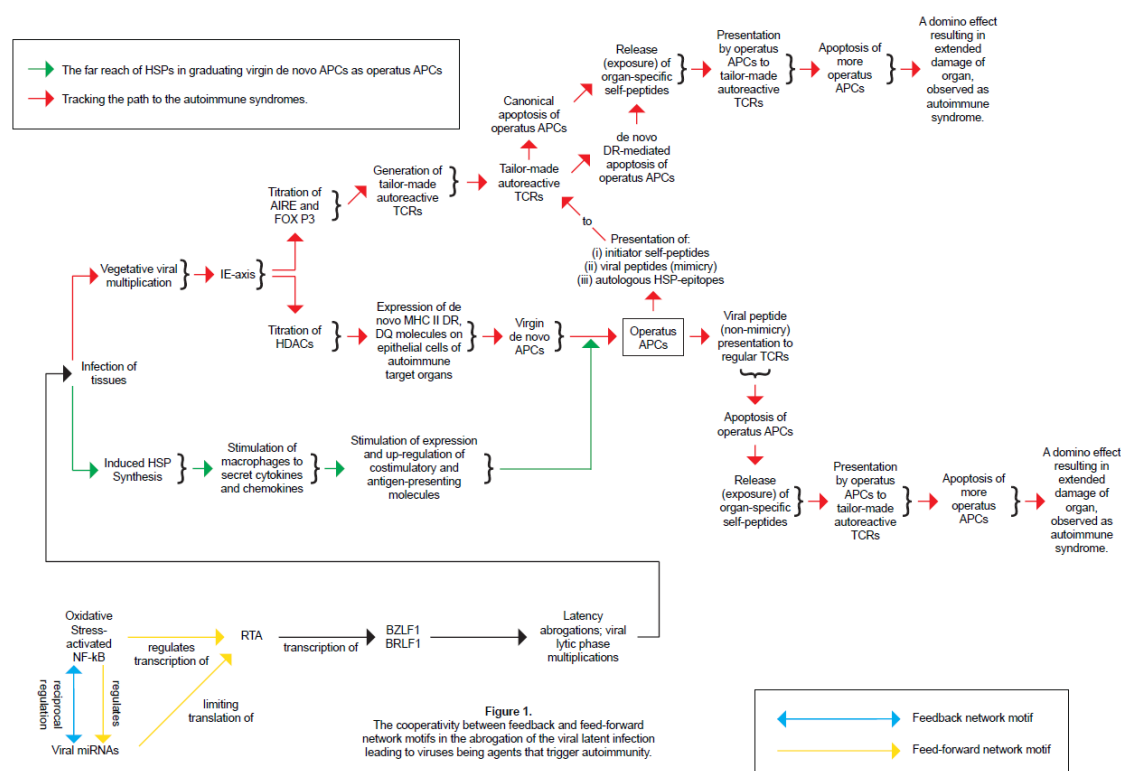


Figure 1. The cooperativity between feedback and feed-forward network motifs in the abrogation of the viral latent infection leading to viruses being agents that trigger autoimmunity.

Take-home Messages

- (1) In man, autoimmunity is an event in which the immune responses mounted in the host are directed against self-components as a result of the loss of self-tolerance. The resulting autoimmune diseases are either tissue-specific when a particular tissue is affected, or systemic when a number of tissues are enrolled.
- (2) To happen, autoimmunity requires triggering by environmental factors: the viruses and oxidative stress, in genetically primed individuals. The viruses and stress feature as stimuli for the activation of the transcription factor NF-KB.
- (3) Post-transcriptional gene regulation of mRNA turnover and translation is mediated by microRNAs and RNA-binding proteins in events identified as network motifs, in the images of feedback and feed-forward mechanisms. The feedback network motif in which transcriptional interactions and miRNA-mediated post-transcriptional interactions are involved occurs with stress-activated NF-KB and viral miRNAs in a reciprocal regulation between NF-KB and viral miRNAs such that NF-KB regulates the viral miRNAs and is itself regulated by the same miRNAs.
- (4) NF-KB and viral miRNAs compete for RTA site with which they interact. The NF-KB regulation of miRNA above is a down-regulation that consequents in miRNA depletion by RTA site in a process called sponging, thus sparing the RTA site for NF-KB; this scenario favors NF-KB-activated transcription and expression of RTA, the lytic master switch, leading to abrogation of the viral latency and to a lytic replication.
- (5) The feed-forward network motif reveals a mechanism through which stress-activated NF-KB simultaneously activates RTA transcription and limits its translation through activation of the viral miRNA.
- (6) Finally, as Figure 1 displays, the feedback and feed-forward mechanisms cooperate in the switching on of RTA which in turn activates the transcription and expression of the immediate early genes BZLF1 and BRLF1. These two immediate early genes mobilize the expression of lytic genes and thus abrogate the viral latency, consequentially setting in trend the journey of the virus to the triggering of autoimmunity.

Acknowledgments

This investigation was supported by grants from the NSW Health Department. The author would like to thank Professor Schlink for providing the facilities for the investigation. Dr Temajo is particularly grateful to his wife Julianne Temajo for her enlightening suggestions in their learned and civilized discourse.

Conflict of Interest

All authors declare no conflicts of interest in this review.

References

1. Ray S, Sonthalia N, Kundu S, et al. (2012) Autoimmune disorders: an overview of molecular and cellular basis in today's perspective. *J Clin Cell Immunol* S10: 003.
2. Von Muhlen CA, Tan EM (1995) Autoantibodies in the diagnosis of systemic rheumatic diseases. *Semin Arthritis Rheum* 24: 323–358.
3. Temajo NO, Howard N (2014) The virus-induced HSPs regulate the apoptosis of *operatus* APCs that results in autoimmunity, not in homeostasis. *Autoimmun Rev* 60: 208–218.
4. Temajo NO, Howard N (2012) The viral enterprises in autoimmunity: conversion of target cells into de novo APCs is the presage to autoimmunity. *Autoimmun Rev* 11: 653–658.
5. Temajo NO, Howard N (2014) The mosaic of environment involvement in autoimmunity: The abrogation of viral latency by stress, a non-infectious environmental agent, is an intrinsic prerequisite prelude before viruses can rank as infectious environmental agents that trigger autoimmune diseases. *Autoimmun Rev* 13: 635–640.
6. Agmon-Levin N, Paz Z, Israeli E, et al. (2009) Vaccines and autoimmunity. *Nat Rev Rheumatol* 5: 648–652.
7. Grossman C, Dovrish Z, Shoenfeld Y, et al. (2010) Do infections facilitate the emergence of systemic sclerosis? *Autoimmun Rev* 10: 244–247.
8. Richer MJ, Horwitz MS (2009) Coxsackievirus infection as environmental factor in the etiology of type 1 diabetes. *Autoimmun Rev* 8: 611–615.
9. Goldberg E, Krause I (2009) Infection and type 1 diabetes mellitus—a two edged sword? *Autoimmun Rev* 8: 682–686.
10. Sansonno L, Tucci FA, Sansonno S, et al. (2009) B cells and HCV: an infection model of autoimmunity. *Autoimmun Rev* 9: 93–94.
11. Aota N, Shiohara T (2009) Viral connection between drug rashes and autoimmune diseases: how autoimmune responses are generated after resolution of drug rashes. *Autoimmun Rev* 8: 488–494.
12. Plot L, Amital H (2009) Infectious association of celiac disease. *Autoimmun Rev* 8: 316–319.
13. Rigopoulou EI, Smyk DS, Matthews CE, et al. (2012) Epstein-barr virus as a trigger of autoimmune liver diseases. *Adv Virol* 2012: 1–12.
14. Toussirot E, Roudier J (2008) Epstein-barr virus in autoimmune diseases. *Best Pract Res Clin Rheumatol* 22: 883–896.
15. Alarconriquelme ME (2007) Recent advances in the genetics of autoimmune diseases. *Ann N Y Acad Sci* 1110: 1–9.
16. Zhu M, Ruddy M, Fu YX (2009) The role of NF-KB in central tolerance, In: Moncef Z, *The Epigenetics of Autoimmune Diseases*, John Wiley & Sons, Ltd, 39–53.
17. Hoffmann A, Baltimore D (2006) Circuitry of nuclear factor kappa B signaling. *Immunol Rev* 210: 171–181.
18. Jacobs MD, Harrison SC (1998) Structure of an IKBalpha/NF-KB complex. *Cell* 95: 749–758.
19. Li N, Karin M (1999) Is NF-KB the sensor of oxidative stress? *FASEB J* 13: 1137–1143.
20. Hayden MS, Ghosh S (2008) Shared principles in NF-KB signalling. *Cell* 132: 344–362.
21. Huxford T, Ghosh G (2009) A structural guide to proteins of the NF-KB signaling module. *CSH Perspect Biol* 1: a000075.
22. Schieber M, Chandel NS (2014) ROS function in redox signaling and oxidative stress. *Curr Biol* 24: R453–462.

23. Morgan MJ, Liu ZG (2011) Crosstalk of reactive oxygen species and NF- κ B signalling. *Cell Res* 21: 103–115.
24. Korn SH, Wouters EF, Vos N, et al. (2001) Cytokine-induced activation of nuclear factor- κ B is inhibited by hydrogen peroxide through oxidative inactivation of IkappaB kinase. *J Biol Chem* 276: 35693–35700.
25. Takada Y, Mukhopadhyay A, Kundu GC, et al. (2003) Hydrogen peroxide activates NF- κ B through tyrosine phosphorylation of I kappa B alpha and serine phosphorylation of p65: evidence for the involvement of I kappa B alpha kinase and Syk protein-tyrosine kinase. *J Biol Chem* 278: 24233–24241.
26. Schieven GL, Kirihara JM, Myers DE, et al. (1993) Reactive oxygen intermediates activate NF- κ B in a tyrosine kinase-dependent mechanism and in combination with vanadate activate the p56lck and p59fyn tyrosine kinases in human lymphocytes. *Blood* 82: 1212–1220.
27. Jr CT, Jr BE, Farr A, et al. (1999) Oxidative stress induces NF- κ B nuclear translocation without degradation of IkappaBalpha. *Circulation* 100: II361–364.
28. Ferreira V, Schoonbroodt S, Best-Belpomme M, et al. (2000) Crucial role of the amino-terminal tyrosine residue 42 and the carboxyl-terminal PEST domain of I kappa B alpha in NF- κ B activation by an oxidative stress. *J Immun* 164: 4292–4300.
29. Volanti C, Matroule JY, Piette J (2002) Involvement of oxidative stress in NF- κ B activation in endothelial cells treated by photodynamic therapy. *Photochem Photobiol* 75: 36–45.
30. Beraud C, Henzel WJ, Baeuerle PA (1999) Involvement of regulatory and catalytic subunits of phosphoinositide 3-kinase in NF- κ B activation. *P Natl Acad Sci USA* 96: 429–434.
31. Fan C, Li Q, Ross D, et al. (2003) Tyrosine phosphorylation of I kappa B activates NF kappa B through a redox-regulated and c-Src-dependent mechanism following hypoxia/reoxygenation. *J Biol Chem* 278: 2072–2080.
32. Lluís JM, Buricchi F, Chiarugi P, et al. (2007) Dual role of mitochondrial reactive oxygen species in hypoxia signalling: activation of nuclear factor- κ B via c-SRC and oxidant-dependent cell death. *Cancer Res* 67: 7368–7377.
33. He Z, Zhao J, Zhang J, et al. (2014) NF- κ B activation coordinated by IKK β and IKK γ enables latent infection of Kaposi's Sarcoma-associated Herpesvirus. *J Virol* 88: 444–455.
34. Zhong H, Voll RE, Ghosh S (1998) Phosphorylation of NF- κ B p65 by PKA stimulates transcriptional activity by promoting a novel bivalent interaction with the coactivator CBP/p300. *Mol Cell* 1: 661–671.
35. Neumann M, Naumann M (2007) Beyond Ikappa Bs: alternative regulation of NF- κ B activity. *FASEB J* 21: 2642–2654.
36. Huang B, Yang XD, Lamb A, et al. (2010) Posttranslational modifications of NF- κ B: another layer of regulation for NF- κ B signalling pathway. *Cell Signal* 22: 1282–1290.
37. Chen LF, Williams SA, Mu Y, et al. (2005) NF- κ B RelA phosphorylation regulates RelA acetylation. *Mol Cell Biol* 25: 7966–7975.
38. Hiscott J, Kwon H, Genin P (2001) Hostile takeovers: viral appropriation of the NF- κ B pathway. *J Clin Invest* 107: 143–151.
39. Zhou FC, Zhang YJ, Deng JH, et al. (2002) Efficient infection by a recombinant Kaposi's sarcoma-associated herpesvirus cloned in a bacterial artificial chromosome: application for genetic analysis. *J Virol* 76: 6185–6196.

40. Speck SH, Ganem D (2010) Viral latency and its regulation: lessons from the gamma-herpesviruses. *Cell Host Microbe* 8: 100–115.
41. Lu F, Weidmer A, Liu CG, et al. (2008) Epstein-barr virus-induced MiR-155 attenuates NF-KB signaling and stabilizes latent virus persistence. *J Virol* 82: 10436–10443.
42. Cullen BR (2006) Viruses and microRNAs. *Nat Genet* 38 (Suppl): S25–30.
43. Grey F, Hook L, Nelson J (2007) The functions of herpesvirus-encoded microRNAs. *Med Microbiol Immun* 197: 261–267.
44. Pfeffer S, Voinnet O (2006) Viruses, microRNAs and cancer. *Oncogene* 25: 6211–6219.
45. Kincaid RP, Sullivan CS (2012) Virus-encoded microRNAs: an overview and a look to the future. *PLoS Pathog* 8: e1003018.
46. Murphy E, Vanicek J, Robins H, et al. (2008) Suppression of immediate-early viral gene expression by herpesvirus-coded microRNAs: implications for latency. *P Natl Acad Sci USA* 105: 5453–5458.
47. Pfeffer S, Zavolan M, Grasser FA, et al. (2004) Identification of virus-encoded microRNAs. *Science* 304:734–736.
48. Grey F, Meyers H, White EA, et al. (2007) A human cytomegalovirus-encoded microRNA regulates expression of multiple viral genes involved in replication. *PLoS Pathog* 3: e163.
49. Lu F, Stedman W, Yousef M, et al. (2010) Epigenetic regulation of Kaposi's sarcoma-associated herpesvirus latency by virus-encoded microRNAs that target RTA and the cellular Rb12-DNMT pathway. *J Virol* 84: 2697–2706.
50. Bellare P, Ganem D (2009) Regulation of KSHV lytic switch protein expression by virus-encoded microRNA: an evolutionary adaptation that fine-tunes lytic reactivation. *Cell Host Microbe* 6: 570–575.
51. Lin X, Liang D, He Z, et al. (2011) miR-K12-7-5p encoded by Kaposi's sarcoma-associated herpesvirus stabilizes the latent state by targeting viral ORF50/RTA. *PLoS One* 6: e16224.
52. Bartel DP (2009) MicroRNAs: target recognition and regulatory functions. *Cell* 136: 215–233.
53. Lei X, Bai Z, Ye F, et al. (2010) Regulation of NF-KB inhibitor IkappaBalpha and viral replication by a KSHV microRNA. *Nat Cell Biol* 12:193–199.
54. Israel BF, Kenney SC (2003) Virally targeted therapies for EBV-associated malignancies. *Oncogene* 22: 5122–5130.
55. Pantry SN, Medvezky PG (2009) Epigenetic regulation of Kaposi's sarcoma-associated herpesvirus replication. *Semin Cancer Biol* 19:153–157.
56. Jens M, Rajewsky N (2014) Competition between target sites of regulators shapes post-transcriptional gene regulation. *Nat Rev Genet* 16: 113–126.
57. Karreth FA, Tay Y, Pandolfi PP (2014) Target competition: transcription factors enter the limelight. *Genome Biol* 15:114–116.
58. Brewster RC, Weinert FM, Garcia HG, et al. (2014) The transcription factor titration effect dictates level of gene expression. *Cell* 156: 1312–1323.
59. Lal A, Mazan-Mamczarz K, Kawai T, et al. (2004) Concurrent versus individual binding of HuR and AUF1 to common labile target mRNAs. *EMBO J* 23: 3092–3102.
60. Barker A, Epis MR, Porter CJ, et al. (2012) Sequence requirements for RNA binding by HuR and AUF1. *J Biochem* 151: 423–437.
61. Martinez NJ, Walhout AJM (2009) The interplay between transcription factors and microRNAs in genome-scale regulatory networks. *Bioessays* 31: 435–445.

62. Yeager-Loetem E, Sattath S, Kashtan N, et al. (2004) Network motifs in integrated cellular networks of transcription-regulation and protein-protein interaction. *P Natl Acad Sci USA* 101: 5934–5939.
63. Vento-Tormo R, Rodriguez-Ubreva J, Lisio LD, et al. (2014) NF- κ B directly mediates epigenetic deregulation of common microRNAs in Epstein-Barr virus-mediated transformation of B-cells and in lymphomas. *Nucleic Acids Res* 42: 11025–11039.
64. Vallabhapurapu S, Karin M (2009) Regulation and function of NF- κ B transcription factors in the immune system. *Annu Rev Immunol* 27: 693–733.
65. Temajo NO, Howard N (2014) The virus-induced HSPs regulate the apoptosis of *operatus* APCs that results in autoimmunity, not in homeostasis. *Autoimmun Rev* 60: 208–218.



AIMS Press

© 2017 Norbert O. Temajo, et al., licensee AIMS Press. This is an open access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>)