



Editorial

Antimicrobial agents and microbial ecology

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Abstract: Antimicrobials are therapeutic substances used to prevent or treat infections. Disinfectants are antimicrobial agents applied to non-living surfaces. Every year, several thousand tonnes of antimicrobials and their by-products are released into the environment and in particular into the aquatic environment. This type of xenobiotic has ecological consequences in the natural environment but also in technological environments such as wastewater treatment plants and methane fermentation sewage sludge treatment plants. The constant exposure of microbial communities not only to high concentrations but also to sub-inhibitory concentrations of antibiotics is a key element in the development of antibiotic resistance in aquatic environments and in soils. The future of antimicrobials lies in the development of biosourced or bioinspired molecules. The observation and deciphering of interactions between living organisms is the key to this development.

Keywords: Antimicrobial; antibiotic; disinfectant; resistance; ecology; biosourced; bioinspired

Antimicrobials are therapeutic substances used to prevent or treat infections. They include antiseptics, antibiotics, antivirals, antifungals and antiparasitics. Disinfectants are antimicrobial agents applied to non-living surfaces. Antimicrobials can kill microorganisms and/or prevent their growth by targeting key steps in cellular metabolism such as the synthesis of biological macromolecules, the activity of cellular enzymes, or cellular structures such as the cell wall, cell membranes [1–4]. The presence of antimicrobial agents in an ecosystem, whether natural or man-made, always has an ecological impact [5].

Every year, several thousand tonnes of antimicrobials and their by-products are released into the environment and in particular into the aquatic environment [6]. In addition, some antimicrobials are particularly persistent in the environment, which facilitates their diffusion and accumulation in

different compartments. This type of xenobiotic has ecological consequences in the natural environment but also in technological environments such as wastewater treatment plants and methane fermentation sewage sludge treatment plants [7–9]. The constant exposure of microbial communities not only to high concentrations but also to sub-inhibitory concentrations of antibiotics is a key element in the development of antibiotic resistance in aquatic environments and in soils [5,10]. This leads to the development of antibiotic-resistant bacteria and to the spread of genetic determinants of resistance. In this context, environmental biofilms and their associated virome serve as reservoirs for antimicrobial resistance [10]. The development of nanoparticles with antibacterial activity is one of the solutions for combating bacteria that are multi-resistant to antibiotics [11].

Chlorine-based treatments are widely used during drinking water production and distribution. The post-chlorine treatment survival and regrowth of microorganisms, due to microbial chlorine resistance and to microbial chlorine tolerance, can conduct to an increase of human exposure to waterborne pathogens [12]. Resistance has a genetic origin and is transmissible, whereas tolerance is linked to transient phenotypic adaptations, i.e., extracellular polymeric substances production and biofilm formation [13,14]. Microbial resistance to chlorine also poses problems for wastewater treatment [15].

Antimicrobials have been present in nature for much longer than their use by humans. The study of interactions between different microorganisms, whether prokaryotes or eukaryotes, is an important source of new antimicrobial discovery [16–20]. Ecological knowledge of antimicrobial production conditions, functions and roles in an ecosystem is necessary for the discovery of new antimicrobials and their safe and effective use. This approach can also be used for man-made environments like fermented artisanal food products [21].

The microbial ecology of the human mouth and gastrointestinal tract is very complex. The use of broad-spectrum antimicrobial agents can lead to (i) proliferation of previously minor components of the microbiota, (ii) colonisation by saprophytic organisms, (iii) colonisation by antimicrobial resistant pathogens, which may increase the risk of disease. [22–24]. Cranberry can help prevent urinary tract infections and avoid the use of antibiotics [25,26]. In contrast to antibiotic treatment, dietary cranberry supplementation does not appear to affect the faecal concentrations of thermotolerant coliforms, *Enterococci* spp. and *Lactobacilli* spp. in Wistar rats [27]. Thus, dietary use of cranberry would not disturb the microbial ecology of the intestinal tract.

Antimicrobials are micropollutants that have a strong impact on different ecosystems. Studies are needed to understand their ecological impact and learn how to control them. The future of antimicrobials lies in the development of biosourced or bioinspired molecules. The observation and deciphering of interactions between living organisms is the key to this development.

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Conflict of interest

The author declares no conflicts of interest in this article.

References

1. Pursell E (2020) Antimicrobials. In: Hood, P., Khan, E., eds., *Understanding Pharmacology in Nursing Practice*, Springer Nature, Switzerland, 147–165. https://doi.org/10.1007/978-3-030-32004-1_6.
2. Reygaert WC (2018) An overview of the antimicrobial resistance mechanisms of bacteria. *AIMS Microbiol* 4: 482–501. <https://doi.org/10.3934/microbiol.2018.3.482>.
3. Di Martino P (2021) Ways to improve biocides for metalworking fluid. *AIMS Microbiol* 7: 13–27. <https://doi.org/10.3934/microbiol.2021002>.
4. Romani M, Warscheid T, Nicole L, et al. (2022) Current and future chemical treatments to fight biodeterioration of building materials and associated biofilms: moving away from ecotoxic and towards efficient, sustainable solutions. *Sci Total Environ* 802: 149846. <https://doi.org/10.1016/j.scitotenv.2021.149846>.
5. Grenni P, Ancona V, Caracciolo AB (2018) Ecological effects of antibiotics on natural ecosystems: A review. *Microchem J* 136: 25–39. <https://doi.org/10.1016/j.microc.2017.02.006>.
6. Felis E, Kalka J, Sochacki A, et al. (2020) Antimicrobial pharmaceuticals in the aquatic environment - occurrence and environmental implications. *Eur J Pharmacol* 866: 172813. <https://doi.org/10.1016/j.ejphar.2019.172813>.
7. Varela AR, Andr es, Nunes OC, et al. (2014) Insights into the relationship between antimicrobial residues and bacterial populations in a hospital-urban wastewater treatment plant system. *Water Res* 54: 327–336. <https://doi.org/10.1016/j.watres.2014.02.003>.
8. Du B, Wang Q, Yang Q, et al. (2021) Responses of bacterial and bacteriophage communities to long-term exposure to antimicrobial agents in wastewater treatment systems. *J Hazard Mater* 414: 125486. <https://doi.org/10.1016/j.jhazmat.2021.125486>.
9. Czatzkowska M, Harnisz M, Korzeniewska E, et al. (2021) The impact of antimicrobials on the efficiency of methane fermentation of sewage sludge, changes in microbial biodiversity and the spread of antibiotic resistance. *J Hazard Mater* 416: 125773. <https://doi.org/10.1016/j.jhazmat.2021.125773>.
10. Flores-Vargas G, Bergsveinson J, Lawrence JR, et al. (2021) Environmental biofilms as reservoirs for antimicrobial resistance. *Front Microbiol* 12: 3880. <https://doi.org/10.3389/fmicb.2021.766242>.
11. Hashem NM, Hosny A, Abdelrahman AA, et al. (2021) Antimicrobial activities encountered by sulfur nanoparticles combating Staphylococcal species harboring *scmecA* recovered from acne vulgaris. *AIMS Microbiol* 7: 481–498. <https://doi.org/10.3934/microbiol.2021029>.
12. Ekundayo TC, Igwaran A, Oluwafemi YD, et al. (2021) Global bibliometric meta-analytic assessment of research trends on microbial chlorine resistance in drinking water/water treatment systems. *J Environ Manage* 278: 111641. <https://doi.org/10.1016/j.jenvman.2020.111641>.
13. Xu ZS, Yang X, G anzle MG (2021) Resistance of biofilm- and pellicle-embedded strains of *Escherichia coli* encoding the transmissible locus of stress tolerance (tLST) to oxidative sanitation chemicals. *Int J Food Microbiol* 359: 109425. <https://doi.org/10.1016/j.ijfoodmicro.2021.109425>.
14. Rajeev M, Sushmitha TJ, Prasath KG, et al. (2020) Systematic assessment of chlorine tolerance mechanism in a potent biofilm-forming marine bacterium *Halomonas boliviensis*. *Int Biodeterior Biodegrad* 151: 104967. <https://doi.org/10.1016/j.ibiod.2020.104967>.

15. Wang YH, Wu YH, Tong X, et al. (2019) Chlorine disinfection significantly aggravated the biofouling of reverse osmosis membrane used for municipal wastewater reclamation. *Water Res* 154: 246–257. <https://doi.org/10.1016/j.watres.2019.02.008>.
16. Molloy EM, Hertweck C (2017) Antimicrobial discovery inspired by ecological interactions. *Curr Opin Microbiol* 39: 121–127. <https://doi.org/10.1016/j.mib.2017.09.006>.
17. Pishchany G (2020) Applying microbial ecology to antimicrobial discovery. *Curr Opin Microbiol* 57: 7–12. <https://doi.org/10.1016/j.mib.2020.03.007>.
18. Far BE, Ragheb M, Rahbar R, et al. (2021) Cloning and expression of *Staphylococcus simulans* lysostaphin enzyme gene in *Bacillus subtilis* WB600. *AIMS Microbiol* 7: 271–283. <https://doi.org/10.3934/microbiol.2021017>.
19. Abdel Fattah R, Fathy F, Mohamed T, et al. (2021) Effect of chitosan nanoparticles on quorum sensing-controlled virulence factors and expression of *LasI* and *RhlI* genes among *Pseudomonas aeruginosa* clinical isolates. *AIMS Microbiol* 7: 415–430. <https://doi.org/10.3934/microbiol.2021025>.
20. Pringgenies D, Setyati WA (2021) Antifungal strains and gene mapping of secondary metabolites in mangrove sediments from Semarang city and Karimunjawa islands, Indonesia. *AIMS Microbiol* 7: 499–512. <https://doi.org/10.3934/microbiol.2021030>.
21. Dal Bello B, Rantsiou K, Bellio A, et al. (2010) Microbial ecology of artisanal products from North West of Italy and antimicrobial activity of the autochthonous populations. *LWT* 43: 1151–1159. <https://doi.org/10.1016/j.lwt.2010.03.008>.
22. Marsh PD (2010) Controlling the oral biofilm with antimicrobials. *J Dent* 38: S11–S15. [https://doi.org/10.1016/S0300-5712\(10\)70005-1](https://doi.org/10.1016/S0300-5712(10)70005-1).
23. Perez F, Pultz MJ, Endimiani A, et al. (2011) Effect of antibiotic treatment on establishment and elimination of intestinal colonization by KPC-producing *Klebsiella pneumoniae* in mice. *Antimicrob Agents Chemother* 55: 2585–2589. <https://doi.org/10.1128/AAC.00891-10>.
24. Yang JJ, Wang JT, Cheng A, et al. (2018) Impact of broad-spectrum antimicrobial treatment on the ecology of intestinal flora. *J Microbiol Immunol Infect* 51: 681–687. <https://doi.org/10.1016/j.jmii.2016.12.009>.
25. Griffiths P (2003) The role of Cranberry juice in the treatment of urinary tract infections. *Br J Commun Nurs* 557–561. <https://doi.org/10.12968/bjcn.2003.8.12.11853>.
26. Nowack R, Schmitt W (2008) Cranberry juice for prophylaxis of urinary tract infections—Conclusions from clinical experience and research. *Phytomedicine* 15: 653–667. <https://doi.org/10.1016/j.phymed.2008.07.009>.
27. Chettaoui R, Mayot G, De Almeida L, et al. (2021) Cranberry (*Vaccinium macrocarpon*) dietary supplementation and fecal microbiota of Wistar rats. *AIMS Microbiol* 7: 257–270. <https://doi.org/10.3934/microbiol.2021016>.

