



*Research article*

## **The synthesis of conjugated polymers with different length side chains and the effect on their nanoparticles**

**Qiu-bo Wang<sup>1</sup>, Chao Xu<sup>1</sup>, Yi-bao Jiang<sup>1</sup>, Xian Zhang<sup>1\*</sup>, Jin-shui Yao<sup>1</sup>, Cong-de Qiao<sup>1</sup>, Qin-ze Liu<sup>1</sup> and Yuan-hong Zhang<sup>2</sup>**

<sup>1</sup> School of Materials Science and Engineering, Qilu University of Technology (Shandong Academy of Sciences), Jinan, 250353, China

<sup>2</sup> College of Chemistry and Material Structure, Shandong Agricultural University, Taian, 271018, China

\* **Correspondence:** Email: zhangx@qlu.edu.cn; Tel: +8653189631227.

**Abstract:** In recent years, conjugated polymers (CPs) and their nanoparticles (CPNs) were widely concerned and applied in many fields due to their outstanding advantages such as higher absorption coefficients, good photostability, low toxicity and good biocompatibility. However, most reports were focused on their applications, and rare attentions were given on the synthesis of CPs and the effect factors of CPNs' morphology and properties. In this work, four CPs (P1, P2, P3, P4) with different length side chains were synthesized by Wittig–Horner reaction and further used to prepare CPNs by the nano-precipitation method. The CPs and CPNs were characterized and analyzed by Hydrogen Nuclear Magnetic Resonance (<sup>1</sup>H NMR), gel permeation chromatography (GPC), Fourier transform infrared spectroscopy (FTIR), dynamic light scattering (DLS) and transmission electron microscope (TEM). The absorbance and fluorescence emission spectra of CPs in tetrahydrofuran (THF) and CPNs solutions with different length side chains were investigated, and their quantum yields ( $\Phi_s$ ) were calculated by the reference method. The results showed that the synthetic CPs have narrow polydispersity index. Some interesting phenomenon appeared with the increased side chain length. The molecular weights will reduce except P3, and the absorbance and fluorescence emission spectra of CPs and CPNs are red-shifted.  $\Phi_s$  are better and enlarged for CPs and CPNs, and better dispersion and uniform size were founded with longer side chains.

**Keywords:** conjugated polymer; nanoparticles; side chain; length; carbazole; hydroquinone

---

## 1. Introduction

Over the past ten years, conjugated polymer nanoparticles (CPNs) have attracted more attentions because of their merits in comparison to organic fluorescence dyes and quantum dots, such as higher absorption coefficients, good photostability, low toxicity and good biocompatibility [1,2]. These features are widely applied in biological fields including pH sensing, metal ions and temperature sensing [3–6], molecular imaging of tissues and biomarkers of disease [7,8], photodynamic therapy of tumors [9,10], immediate activation of nerve cells, and photothermal therapy of cancer [11,12] and so on. In general, CPNs consist of one or more types of conjugated polymers (CPs), and they supplied photoelectric properties. Therefore, a complete CPNs preparation process is divided into two steps including the synthesis of CPs and the preparation of nanoparticles.

In the polymerization methods of CPs, palladium catalyzed coupling (Suzuki coupling) [13], Sonogashira coupling [14], Heck coupling [15] and Wittig–Horner [16] reactions are widely used in the preparation of PFP, PPE and PPV. CPNs can be obtained by the methods such as nano-precipitation [17,18], microemulsion [19,20] and self-assembly [21,22] based on the synthesized CPs.

As we all know, the different structure and morphology of CPNs will supply diverse performances. The influencing factors include the structure of CPs and the above preparation methods, and the former can be subdivided into the effects of main chains, side chains and substituent groups. A main chain is the most important part of CPs, which determines the basic physical and chemical properties. At present, the designs of the main chain structure are the focus of most work [23]. Side chains play a crucial role in improving the molecular weight, solubility and processability of polymers [24,25]. Therefore, the research showed that flexible side chains other than the main chain have a large influence on the optical properties of CPs, which can change the intermolecular forces and steric hindrance, affecting the effective conjugate chain length and energy band, absorption and emission [26,27]. Some researches about conjugated polymer side chain engineering have been reported [28,29]. The studies were mainly focused on the effect of different alkyl side chains to polymer solubility, the influence of the side chain spacer length to the charge transfer and the tuning of the branch position of the side chain to the properties of CPs [26,30,31]. For all we know, the studies on the effects of different length side chains on the properties of CPs and their nanoparticles are still scarce. Therefore, in this work, we synthesized four hydrophobic CPs by Wittig–Horner reaction and their CPNs by nano-precipitation method with different length side chains, and studied the effects of different length side chains on the optical properties of CPs and their nanoparticles.

## 2. Materials and methods

### 2.1. Materials

Carbazole and *N,N*-dimethyl formamide were purchased from Fuyu fine chemical, Tianjin, China. Hydroquinone and 1-bromo-ethylhexane were purchased from Macklin Biochemical, Shanghai, China. Bromoethane, bromo-*n*-butane, bromo-*n*-octane, bromo-*n*-hexadecane and Polystyrene-maleic anhydride (PSMA) with molecular weight of 18,000 were purchased from Aladdin, Shanghai, China. Other chemicals were AR grade and purchased from the commercial

corporation without further purification.

## 2.2. Methods

$^1\text{H}$  NMR (400 MHz) spectra were determined and recorded using a Bruker Avance spectrometer, and TMS was used as the internal standard. The sample was grinded with KBr to form pellets, then Fourier transform infrared spectroscopy (FTIR) was measured on a Thermo IS10 spectrometer. UV and Fluorescence spectra were measured and recorded on Shimadzu UV-3600 UV spectrophotometer and Hitachi-7000 fluorescence spectrometer, respectively. Transmission Electron Microscope (TEM) images were recorded through JEM-2100 electron microscope. Dynamic light Scattering (DLS) was carried out on a Malvern Zetasizer Nano ZS analyzer.

## 2.3. General procedure for the synthesis of polymers (P1–P4)

9-ethyl-3,6'-diformyl-carbazole (Monomer 1a), 9-butyl-3,6'-diformyl-carbazole (Monomer 1b), 9-octyl-3,6'-diformyl-carbazole (Monomer 1c) and 9-cetyl-3,6'-diformyl-carbazole (Monomer 1d) and 1,4-phosphonic acid diethyl ester-2,5-(1-methyl ether-ethylhexane)-benzene (Monomer 2) were obtained according to the previous methods with carbazole and hydroquinone as raw materials, respectively [32,33].

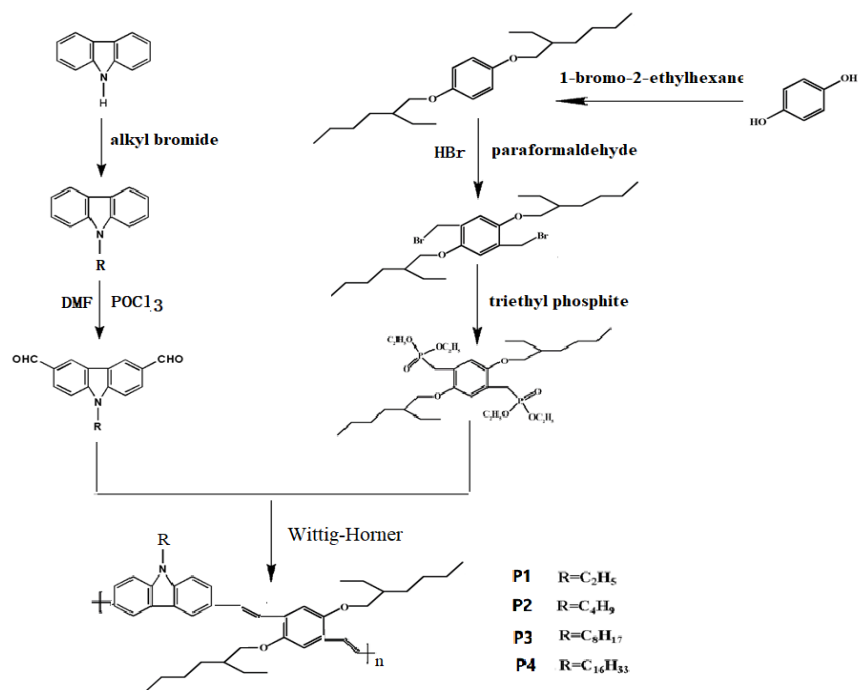
The polymer (P1) was synthesized by Wittig–Horner reaction on the basis of Monomer 1a and Monomer 2 (Scheme 1). The Monomer 2 (0.3 g, 0.45 mmol) was added to the dry tetrahydrofuran (THF) under nitrogen atmosphere and the potassium tertbutoxide (0.2 g, 1.7 mmol) dissolved in THF was added dropwise to the mixture and stirred for 30 minutes in ice–water bath. Then the Monomer 1a (0.15 g, 0.59 mmol) was added at room temperature (r.t.). The reaction mixture was kept stirring for 72 hours at r.t. under nitrogen atmosphere, then poured into the water and extracted with dichloromethane. The solvent was removed by rotary evaporator. The residue was refined by fractional diffusion precipitation with THF and n-hexane. Lastly, the yellow solid was obtained with a yield of 58%.  $^1\text{H}$  NMR ( $\text{CDCl}_3$ , 400 Hz)  $\delta$  (ppm): 8.25 (d, 2H), 7.75 (m, 2H), 7.55 (t, 2H), 7.35 (d, 2H), 7.35 (m, 2H), 7.25 (m, 2H), 4.38 (s, 2H), 4.0 (m, 4H), 1.95–1.75 (m, 2H), 1.75–1.25 (m, 19H), 1.1–0.85 (m, 12H); IR (KBr)  $\nu/\text{cm}^{-1}$ : 1670 (C=C), 1600 (Ar–C), 1096 (C–O), 960 (CH=CH).  $M_n = 2.693 \times 10^4$  ( $\pm 1.448\%$ ),  $M_w = 3.279 \times 10^4$  ( $\pm 1.534\%$ ),  $M_w/M_n = 1.218$  ( $\pm 2.109\%$ ).

The synthetic process of polymers P2, P3 and P4 are similar to that of P1.

**P2:** Yield, 50%.  $^1\text{H}$  NMR ( $\text{CDCl}_3$ , 400 Hz)  $\delta$  (ppm): 8.25 (d, 2H), 7.75 (m, 2H), 7.55 (t, 2H), 7.35 (d, 2H), 7.35 (m, 2H), 7.25 (m, 2H), 4.38 (s, 2H), 4.0 (m, 4H), 1.95–1.75 (m, 2H), 1.75–1.15 (m, 23H), 1.15–0.7 (m, 12H); IR (KBr)  $\nu/\text{cm}^{-1}$ : 1685 (C=C), 1598 (Ar–C), 1033 (C–O), 954 (CH=CH).  $M_n = 2.216 \times 10^4$  ( $\pm 1.016\%$ ),  $M_w = 2.419 \times 10^4$  ( $\pm 1.638\%$ ),  $M_w/M_n = 1.092$  ( $\pm 1.928\%$ ).

**P3:** Yield, 55%.  $^1\text{H}$  NMR ( $\text{CDCl}_3$ , 400 Hz)  $\delta$  (ppm): 8.25 (d, 2H), 7.75 (m, 2H), 7.55 (t, 2H), 7.35 (d, 2H), 7.35 (m, 2H), 7.25 (m, 2H), 4.38 (s, 2H), 4.0 (m, 4H), 1.95–1.75 (m, 2H), 1.75–1.15 (m, 33H), 1.15–0.7 (m, 12H); IR (KBr)  $\nu/\text{cm}^{-1}$ : 1666 (C=C), 1598 (Ar–C), 1039 (C–O), 960 (CH=CH).  $M_n = 3.509 \times 10^4$  ( $\pm 3.887\%$ ),  $M_w = 5.032 \times 10^4$  ( $\pm 6.782\%$ ),  $M_w/M_n = 1.434$  ( $\pm 7.817\%$ ).

**P4:** Yield, 46%.  $^1\text{H}$  NMR ( $\text{CDCl}_3$ , 400 Hz)  $\delta$  (ppm): 8.25 (d, 2H), 7.75 (m, 2H), 7.55 (t, 2H), 7.35 (d, 2H), 7.35 (m, 2H), 7.25 (m, 2H), 4.38 (s, 2H), 4.0 (m, 4H), 1.95–1.80 (m, 2H), 1.80–1.15 (m, 49H), 1.15–0.7 (m, 12H); IR (KBr)  $\nu/\text{cm}^{-1}$ : 1662 (C=C), 1592 (Ar–C), 1031 (C–O), 962 (CH=CH).  $M_n = 2.127 \times 10^4$  ( $\pm 5.248\%$ ),  $M_w = 2.397 \times 10^4$  ( $\pm 6.724\%$ ),  $M_w/M_n = 1.127$  ( $\pm 8.529\%$ ).



#### 2.4. The preparation of CPNs

The preparation of CPNs I–COOH~CPNs IV–COOH: The stock solutions of CPs (P1, P2, P3, P4) and PSMA in THF were prepared with the concentrations of 1 mg/mL and 2 mg/mL, respectively. 0.5 mL of CPs solutions and 0.1 mL of PSMA were added to 10 mL of THF and mixed. After 5 min, the mixture was quickly poured into 20 mL of ultrapure water in a bath sonicator for 5–10 min. Then THF was removed by rotary evaporator, and the solution was concentrated to 10 mL. The solution of carboxyl functional CPNs I–CPNs IV was obtained through a diameter of 0.22  $\mu\text{m}$  micron filter. The same process was carried out without PSMA to get the nanoparticles of CPNs I, CPNs II, CPNs III and CPNs IV, respectively.

### 3. Results and discussion

#### 3.1. The synthetic discussion

Some relative parameters including number-average molecular weight ( $M_n$ ), weight-average molecular weight ( $M_w$ ), polydispersity index ( $\text{PDI} = M_w/M_n$ ) and degrees of polymerization (DP) were summarized in Table 1. One can see that there were narrower PDI (1.092–1.434) by Wittig–Horner reaction. The molecular weights of P1~P4 were found from 21000 to 35000, and the DPs are from 27 to 51. It was found that  $M_n$ ,  $M_w$  and DP reduced with the rising of side chain length. The possible reason may be that the increased side chain length will generate steric hindrance for the reaction [34,35]. However, P3 (octyl) is exceptional, which may be attributed to the octyl side chain supplying a special interaction with 1-bromo-2-ethylhexane to reduce the steric hindrance.

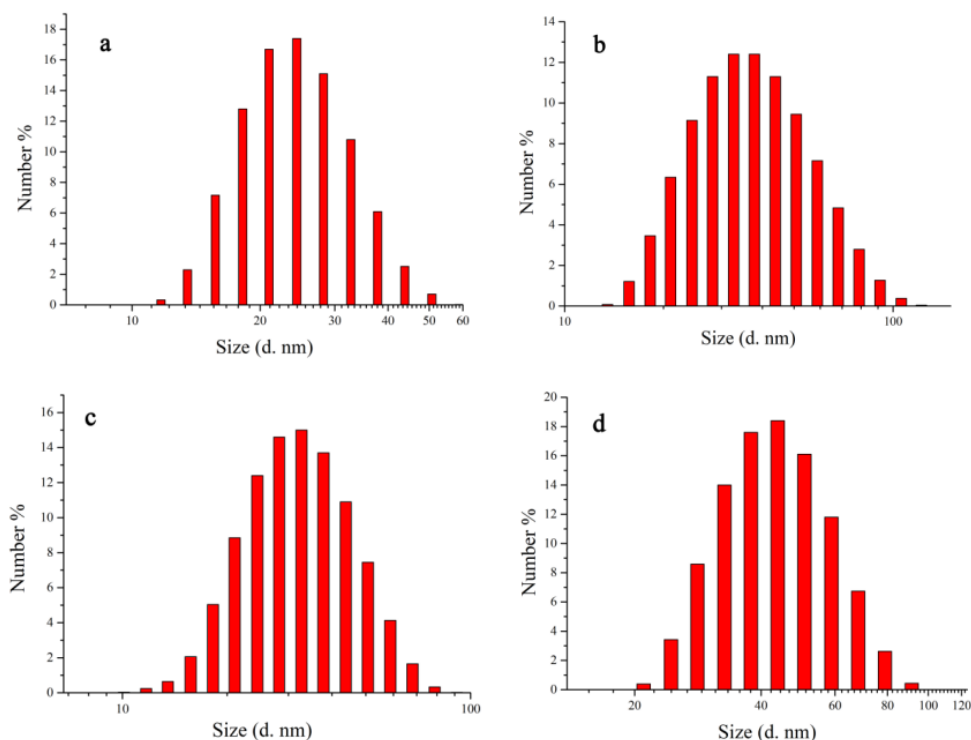
**Table 1.** Some characterization data of conjugate polymers and nanoparticles.

Names	CPNs I	CPNs II	CPNs III	CPNs IV
$M_n$	$2.693 \times 10^4$	$2.216 \times 10^4$	$3.509 \times 10^4$	$2.127 \times 10^4$
$M_w$	$3.279 \times 10^4$	$2.419 \times 10^4$	$5.032 \times 10^4$	$2.397 \times 10^4$
PDI	1.218	1.092	1.434	1.127
Molecular weight of structural units	607	635	689	801
DP	44	35	51	27
Particle size (without PSMA) (nm)	$23.00 \pm 1.00$	$43.95 \pm 2.00$	$32.83 \pm 2.00$	$44.79 \pm 1.00$
Particle size (with PSMA) (nm)	$25.00 \pm 1.00$	$42.15 \pm 2.00$	$28.03 \pm 2.00$	$55.64 \pm 1.00$

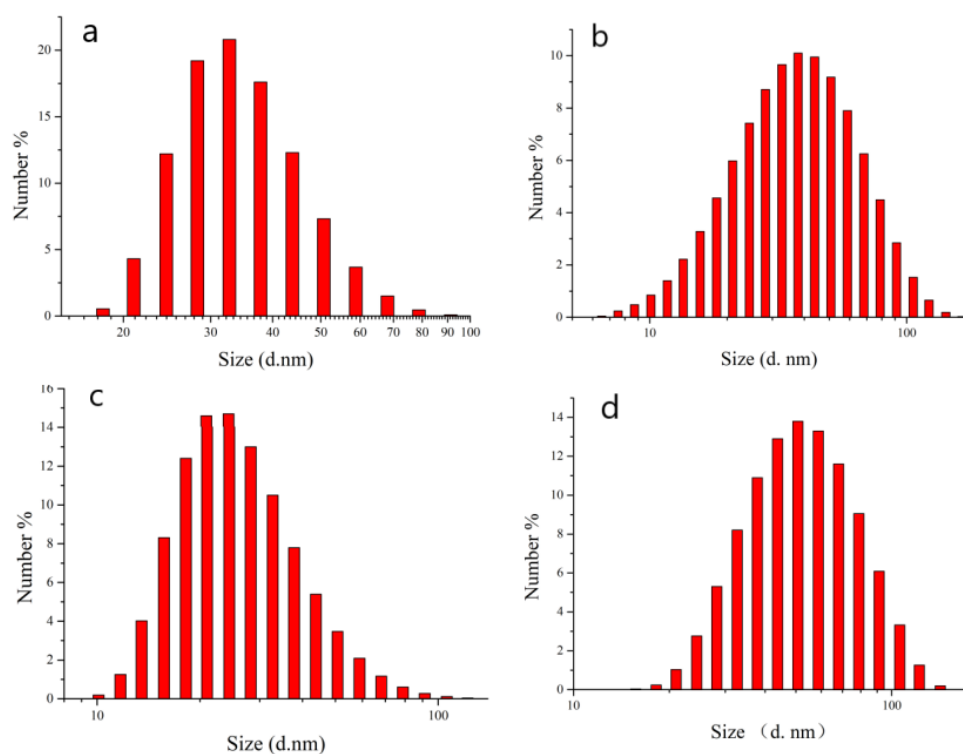
$M_n$ : Number-average molecular weight;  $M_w$ : Weight-average molecular weight; PDI: Polydispersity index ( $M_w/M_n$ ); DP: Degrees of polymerization.

### 3.2. Effects of different side chain length on particle size of nanoparticles

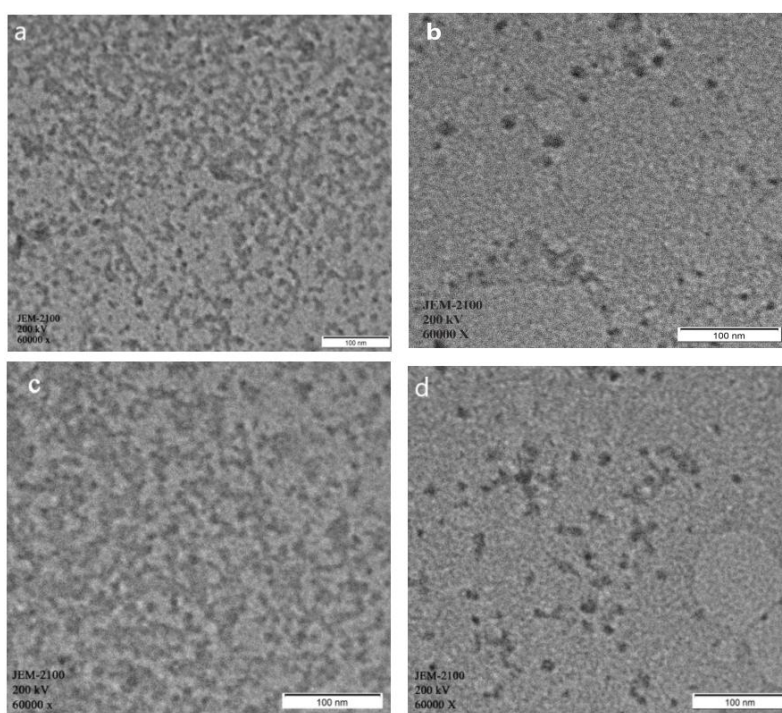
The CPNs were characterized and analyzed by DLS and TEM. The relative data were listed in Table 1. The sizes of CPNs I–IV are 23.00 nm, 43.95 nm, 32.83 nm, and 44.79 nm in Figure 1, respectively. After adding PSMA, their sizes became 25.00 nm, 42.15 nm, 28.03 nm, and 55.64 nm (Figure 2), respectively.



**Figure 1.** The particle size of nanoparticles without PSMA was measured by DLS. (a) The DLS measurement of CPNs I; (b) The DLS measurement of CPNs II; (c) The DLS measurement of CPNs III; (d) The DLS measurement of CPNs IV.



**Figure 2.** The particle size of nanoparticles with PSMA was measured by DLS. (a) The DLS measurement of CPNs I-COOH; (b) The DLS measurement of CPNs II-COOH; (c) The DLS measurement of CPNs III-COOH; (d) The DLS measurement of CPNs IV-COOH.

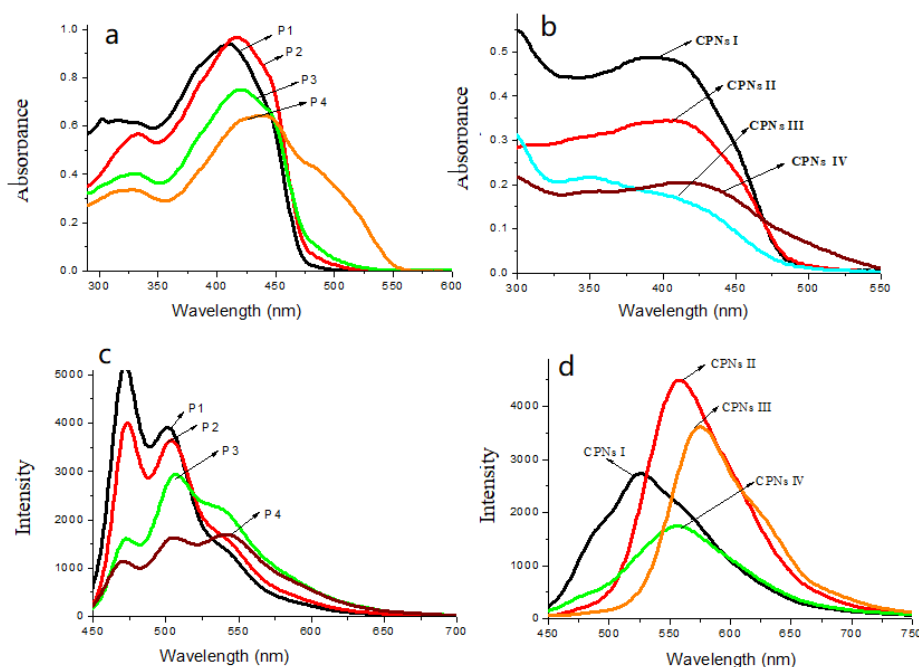


**Figure 3.** TEM images of CPNs I-CPNs IV: (a) CPNs I, (b) CPNs II, (c) CPNs III, (d) CPNs IV.

It is observed that the sizes of CPNs were almost no change before and after adding PSMA. The possible reason is because of the lesser amount of PSMA (CP:PSMA = 5:1). In addition, though DPs are decreasing, the nanoparticles size enhanced with the increased side chain length. As discussed above, the special rule of octyl afforded smaller particles size under a higher molecular weights of P3. Moreover, TEM images in Figure 3 showed that the longer side chain increased the flexibility of polymer, which was benefit to form the better nanoparticles morphology with better dispersion and uniform size.

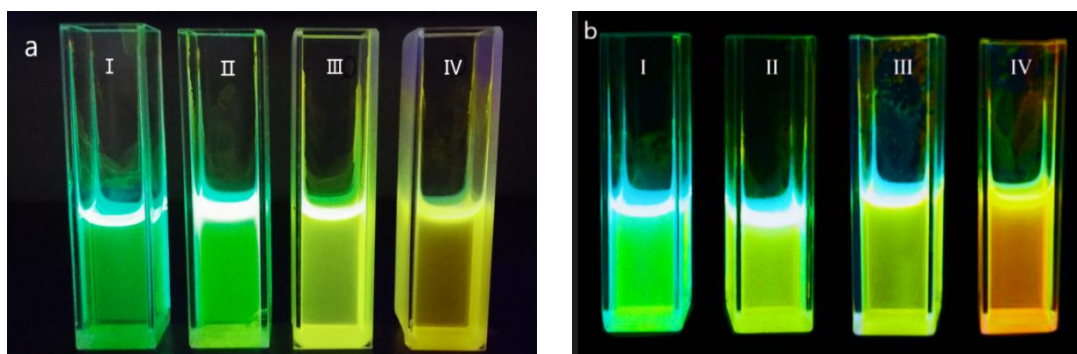
### 3.3. The effect of side chain length on the absorption and fluorescence spectra

The absorption and fluorescence spectra of P1–P4 ( $1 \times 10^{-5}$  mol/L) in THF and CPNs I–IV solutions (10  $\mu\text{g/mL}$ ) were measured and showed in Figure 4. The relative photophysical data were summarized in Table 2. The quantum yields ( $\Phi_s$ ) were calculated by the reference method using coumarin307 ( $\Phi = 0.56$  in methanol) as the standard according to the literature described [36]. From Figure 4 and Table 2, we can find that the absorption and fluorescence spectra of CPs and CPNs are both red-shifted with the increase of side-chain length, and their Stokes shifts ( $\Delta\lambda$ ) showed the same trend. The reason may be attributed to the enhanced conjugation because of longer side chains [37]. The  $\Phi_s$  enhanced with the increased side chain length from 0.25 to 0.40 for CPs and from 0.23 to 0.58 for CPNs. But the values of P3 reached 0.66, which may be affected by the higher DP. In addition, different length side chains of CPs and CPNs supplied various fluorescence colors from green to yellow in Figure 5, but the fluorescence of CPNs is brighter than that of the corresponding CPs. These results indicated that the synthesized CPs and CPNs have better absorbance and fluorescence emission properties.



**Figure 4.** (a and b): The UV absorption spectra of P1–P4 in THF (10  $\mu\text{mol/L}$ ) and CPNs I–IV solution (10  $\mu\text{g/mL}$ ); (c and d) The fluorescence emission spectra of P1–P4 in THF (10  $\mu\text{mol/L}$ ) and CPNs I–IV solution (10  $\mu\text{g/mL}$ ).





**Figure 5.** (a) The fluorescence photographs of CPs (I–IV) in THF at 365 nm; (b) The fluorescence photographs of the CPNs I–IV aqueous at 365 nm.

**Table 2.** The photophysical data of P1–P4 and their nanoparticles (CPNs I–CPNs IV).

	$\lambda_{max}^A$ (nm)	$\lambda_{max}^F$ (nm)	$\Delta\lambda$ (nm)	$\Phi$
P1	410	472	62	0.25
P2	417	473	56	0.33
P3	420	507	87	0.66
P4	440	542	102	0.40
CPNs I	397	525	128	0.23
CPNs II	406	558	152	0.42
CPNs III	409	557	148	0.47
CPNs IV	415	571	155	0.58

$\lambda_{max}^A$  is the wavelength of maximum absorption;  $\lambda_{max}^F$  is the wavelength of maximum fluorescence emission;  $\Delta\lambda$  was Stokes shift;  $\Phi$  is quantum yield.

#### 4. Conclusions

In summary, four CPs of different side chains (P1-ethyl, P2-butyl, P3-octyl, P4-hexadecyl) were prepared by Wittig–Horner reaction based on carbazole and hydroquinone with narrow PDI. The DP declined with the increase of the side chain length except P3. The data of DLS, TEM and the absorbance and fluorescence emission showed the increased side chain length of CPs will be benefit to longer wavelength absorbance, better fluorescence and morphology for CPNs. In addition, the nanoparticle size is lightly affected by simply functional group and the side chain length. These results will be benefit to the design of CPs and the preparation of CPNs.

#### Acknowledgments

This project was supported by the National Natural Science Foundation of China (NSFC 51403111, 11774188). We are particularly grateful to State Key Laboratory of Crystal Materials in Shandong University for help in fluorescence measuring.



## Conflict of interest

There is no conflict of interest regarding the publication of this manuscript.

## References

1. Lyu Y, Pu K (2017) Recent advances of activatable molecular probes based on semiconducting polymer nanoparticles in sensing and imaging. *Adv Sci* 4: 1600481–1600487.
2. Cui Q, Wang X, Yang Y, et al. (2016) Binding-directed energy transfer of conjugated polymer materials for dual-color imaging of cell membrane. *Chem Mater* 28: 4661–4669.
3. Chan YH, Wu C, Ye F, et al. (2011) Development of ultrabright semiconducting polymer dots for ratiometric pH sensing. *Anal Chem* 83: 1448–1455.
4. Wu PJ, Chen JL, Chen CP, et al. (2013) Photoactivated ratiometric copper(II) ion sensing with semiconducting polymer dots. *Chem Commun* 49: 898–900.
5. Zhang P, Lu H, Chen H, et al. (2016) Cationic conjugated polymers-induced quorum sensing of bacteria cells. *Anal Chem* 88: 2985–2988.
6. Wu PJ, Kuo SY, Huang YC, et al. (2014) Polydiacetylene-enclosed near-infrared fluorescent semiconducting polymer dots for bioimaging and sensing. *Anal Chem* 86: 4831–4839.
7. Zhou X, Liang H, Jiang P, et al. (2016) Multifunctional phosphorescent conjugated polymer dots for hypoxia imaging and photodynamic therapy of cancer cells. *Adv Sci* 3: 1500155–1500166.
8. Palner M, Pu K, Shao S, et al. (2015) Semiconducting polymer nanoparticles with persistent near-infrared luminescence for in vivo optical imaging. *Angew Chem Int Edit* 127: 11639–11642.
9. Zhang D, Wu M, Zeng Y, et al. (2016) Lipid micelles packaged with semiconducting polymer dots as simultaneous MRI/photoacoustic imaging and photodynamic/photothermal dual-modal therapeutic agents for liver cancer. *J Mater Chem B* 4: 589–599.
10. Lyu Y, Xie C, Chechetka SA, et al. (2016) Semiconducting polymer nanobioconjugates for targeted photothermal activation of neurons. *J Am Chem Sci* 138: 9049–9052.
11. Cai X, Liu X, Liao LD, et al. (2016) Encapsulated conjugated oligomer nanoparticles for real-time photoacoustic sentinel lymph node imaging and targeted photothermal therapy. *Small* 12: 4873–4880.
12. Cheng L, He W, Gong H, et al. (2013) PEGylated micelle nanoparticles encapsulating a non-fluorescent near-infrared organic dye as a safe and highly-effective photothermal agent for in vivo cancer therapy. *Adv Funct Mater* 23: 5893–5902.
13. Li K, Liu B (2010) Water-soluble conjugated polymers as the platform for protein sensors. *Polym Chem* 1: 252–259.
14. Moon JH, McDaniel W, MacLean P, et al. (2007) Live-cell-permeable poly(p-phenylene ethynylene). *Angew Chem Int Edit* 46: 8223–8225.
15. Feng X, Yang G, Liu L, et al. (2012) A convenient preparation of multi-spectral microparticles by bacteria-mediated assemblies of conjugated polymer nanoparticles for cell imaging and barcoding. *Adv Mater* 24: 637–641.
16. Khidre RE, Abdou WM (2016) Wittig–Horner reagents: powerful tools in the synthesis of 5- and 6-heterocyclic compounds; shedding light on their application in pharmaceutical chemistry. *Turk J Chem* 40: 225–247.

17. Wu C, Jin Y, Schneider T, et al. (2010) Ultrabright and bioorthogonal labeling of cellular targets using semiconducting polymer dots and click chemistry. *Angew Chem Int Edit* 49: 9436–9440.
18. Wu C, Schneider T, Zeigler M, et al. (2010) Bioconjugation of ultrabright semiconducting polymer dots for specific cellular targeting. *J Am Chem Sci* 132: 15410–15417.
19. Wu C, Hansen SJ, Hou Q, et al. (2011) Design of highly emissive polymer dot bioconjugates for in vivo tumor targeting. *Angew Chem Int Edit* 50: 3430–3434.
20. Li K, Pan J, Feng SS, et al. (2009) Generic strategy of preparing fluorescent conjugated-polymer-loaded poly(dl-lactide-co-glycolide) nanoparticles for targeted cell imaging. *Adv Funct Mater* 19: 3535–3542.
21. Howes P, Green M, Levitt J, et al. (2010) Phospholipid encapsulated semiconducting polymer nanoparticles: their use in cell imaging and protein attachment. *J Am Chem Sci* 132: 3989–3996.
22. Yang G, Liu L, Yang Q, et al. (2012) A multifunctional cationic pentathiophene: Synthesis, organelle-selective imaging, and anticancer activity. *Adv Funct Mater* 22: 736–743.
23. Ni D, Yang D, Ma S, et al. (2013) Side chains and backbone structures influence on 4,7-dithien-2-yl-2,1,3-benzothiadiazole (DTBT)-based low-bandgap conjugated copolymers for organic photovoltaics. *Front Optoelectron* 6: 418–428.
24. Subramaniyan S, Xin H, Kim FS, et al. (2011) Effects of side chains on thiazolothiazole-based copolymer semiconductors for high performance solar cells. *Adv Energy Mater* 1: 854–860.
25. Zhang W, Shiotsuki M, Masuda T (2007) Synthesis and properties of polymer brush consisting of poly(phenylacetylene) main chain and poly(dimethylsiloxane) side chains. *Polymer* 48: 2548–2553.
26. Zhu L, Jiang C, Chen G, et al. (2017) Side chain engineering: The effect on the properties of isoindigo-based conjugated polymers contain different length and structure alkyl chains on nitrogen atom. *Org Electron* 49: 278–285.
27. Hwang KH, Kim DH, Min HC, et al. (2016) Effect of side chain position and conformation of quinacridone–quinoxaline based conjugated polymers on photovoltaic properties. *J Ind Eng Chem* 34: 66–75.
28. Wang YJ, Larsson M, Huang WT, et al. (2016) The use of polymer-based nanoparticles and nanostructured materials in treatment and diagnosis of cardiovascular diseases: Recent advances and emerging designs. *Prog Polym Sci* 57: 153–178.
29. Kamaly N, Xiao Z, Valencia PM, et al. (2012) Targeted polymeric therapeutic nanoparticles: design, development and clinical translation. *Chem Soc Rev* 41: 2971–3010.
30. Razzellhollis J, Fleischli F, Jahnke AA, et al. (2017) Effects of side-chain length and shape on polytellurophene molecular order and blend morphology. *J Phys Chem C* 121: 11675–11679.
31. Duan C, Willems REM, Franeker JJV, et al. (2016) Effect of side chain length on the charge transport, morphology, and photovoltaic performance of conjugated polymers in bulk heterojunction solar cells. *J Mater Chem A* 4: 1855–1866.
32. Jiang YB, Gao C, Zhang X, et al. (2018) A highly selective and sensitive fluorescence probe with A- $\pi$ -D- $\pi$ -A structure for detection of Ag<sup>+</sup>. *J Mol Struct* 1163: 33–40.
33. Zhang X, Sun YM, Yu XQ, et al. (2009) Synthesis and nonlinear optical properties of several new two-photon photopolymerization initiators about dibenzothiophene derivatives. *Synthetic Met* 159: 2491–2496.
34. Shin CK, Lee H (2004) Effect of alkyl side-chain length and solvent on the luminescent characteristics of poly(3-n-alkylthiophene). *Synthetic Met* 140: 177–181.

35. Lecollinet G, Delorme N, Edely M, et al. (2009) Self-assembled monolayers of bisphosphonates: influence of side chain steric hindrance. *Langmuir* 25: 7828–7835.
36. Meng XL, Zhang X, Yao JS, et al. (2013) Fluorescence and fluorescence imaging of two schiff derivatives sensitive to  $\text{Fe}^{3+}$  induced by single- and two-photon excitation. *Sensor Actuat B-Chem* 176: 488–496.
37. Fu B, Baltazar J, Sankar AR, et al. (2014) Enhancing field-effect mobility of conjugated polymers through rational design of branched side chains. *Adv Funct Mater* 24: 3734–3744.



AIMS Press

© 2018 the Author(s), licensee AIMS Press. This is an open access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>)