



*Research article*

## Columnar structure of SV40 minichromosome

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**Abstract:** Like the sequence of the strongest 601 clone nucleosome of Lowary and Widom, the SV40 genome sequence contains tracks of YR dinucleotides separated by small integers of the 10.4n base series (10, 11, 21 and 30 bases). The tracks, however, substantially exceed the nucleosome DNA size and, thus, correspond to more extended structure - columnar chromatin. The micrococcal nuclease digests of the SV40 chromatin do not show uniquely positioned individual nucleosomes. This confirms the columnar structure of the minichromosome, as well as earlier electron microscopy studies.

**Keywords:** nucleosome; sequence periodicity; YR dinucleotides; YR tracks; stacked nucleosomes; rotational positioning; nucleosome sliding; nucleosome mapping

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### 1. Introduction

SV40 minichromosome is classical object for chromatin structure studies that, in particular, provided convincing EM pictures of the nucleosomes organized in the beads-on-string manner [1,2]. The efforts towards determination of the expected unique positions of the nucleosomes along the 5243 bases long genome of the virus gave inconclusive results, both by computational mapping [3], and by cloning of the micrococcal nuclease (MNase) digest nucleosome DNA size fragments [4]. It was found that 22 of 41 cloned and mapped fragments overlap with other fragments sharing from ~25 to ~125 bases. “Nucleosomes do not occupy unique positions in SV40 minichromosomes” [4]. It appears, thus, that the nucleosomes of SV40 are sliding, so that each individual pattern of the beads is different, and no specific nucleosome repeat length, as in eukaryotes in general, is observed.

There is an alternative view at the chromatin structure—organization of the “nucleosomes” in tight oligonucleosomes [5], also called columnar structures [6–9], or meganucleosomes [10]. In this case, the nucleosomes excised by MNase from the columns in the process of their disintegration

would slide like in SV40 minichromosome, occupying many nearly equivalent alternative positions shifted by 10–11 bases. Such separation (i.e., overlapping) has been first observed by Ponder & Crawford [11].

What we call columnar structures, as referred above, is manifested in the extended ~10 base ladder of nuclease digestion of chromatin [5,10,12], well beyond the nucleosome DNA size. The columns are also seen directly by various versions of electron microscopy of the minimally perturbed samples. They appear as smooth 10 nm fibers [13–15]. Nucleosomes in the fibers seem to be “closely packed forming a continuous 100 Å filament” [13]. The existence of the columns is suggested by observation of long, far exceeding nucleosome size 10–11 base periodic sequence regions in various chromatins [7–9]. Finally, the nucleosome repeat lengths of various eukaryotic organisms, derived by MNase digestion of the chromatins, display discrete values, with increments of ~10 base-pairs [16,17]. The lengths, actually, closely follow 10.4xn series (Trifonov, in press).

The nucleosomes can be mapped on genome sequences by matching to standard periodic consensus pattern  $(RRRRRYYYYY)_n$  [18–20]. In case of the columns the mapping reveals RR/YY oscillations significantly beyond the nucleosome DNA size [9]. Apart from the RR/YY oscillation the pattern contains also the periodic YR dinucleotides which appear to make significant contribution, especially due to TA periodicity [21,22]. There is no mapping algorithm available which would take into account the corresponding (unknown) weights of the RR/YY and YR components. One can apply, however, for the nucleosome (column) mapping purposes two separate algorithms based on the above  $(RRRRRYYYYY)_n$  consensus (which includes YR as a part of the signal) and on the YR-periodic tracks [23]. The prominent strong nucleosome forming sequence, clone 601 [21], is good example of both components present [22].

In this study we used both algorithms, combining the data in one map, with conclusion that the SV40 minichromosome consists of several connected columnar structures, rather than of solitary beads of the nucleosomes.

## 2. Materials and Methods

The 5243 base sequence of the SV-40 genome is taken from NCBI Genbank.

The construction of the YR tracks follows the rule suggested by the crystal structure of the 601 clone nucleosome [24]: The sequence bound by histone octamers consists of 10-mers or 11-mers starting with YR dinucleotides, occasionally separated by ordinary sequence 10–11mers (up to three 10–11 base period separations). Such succession of YR 10-11-mers makes a YR track.

The nucleosome mapping by the  $(RRRRRYYYYY)_n$  probe is described in [20] and can be implemented via server <http://strn-nuc.haifa.ac.il:8080/mapping/home.jsf>

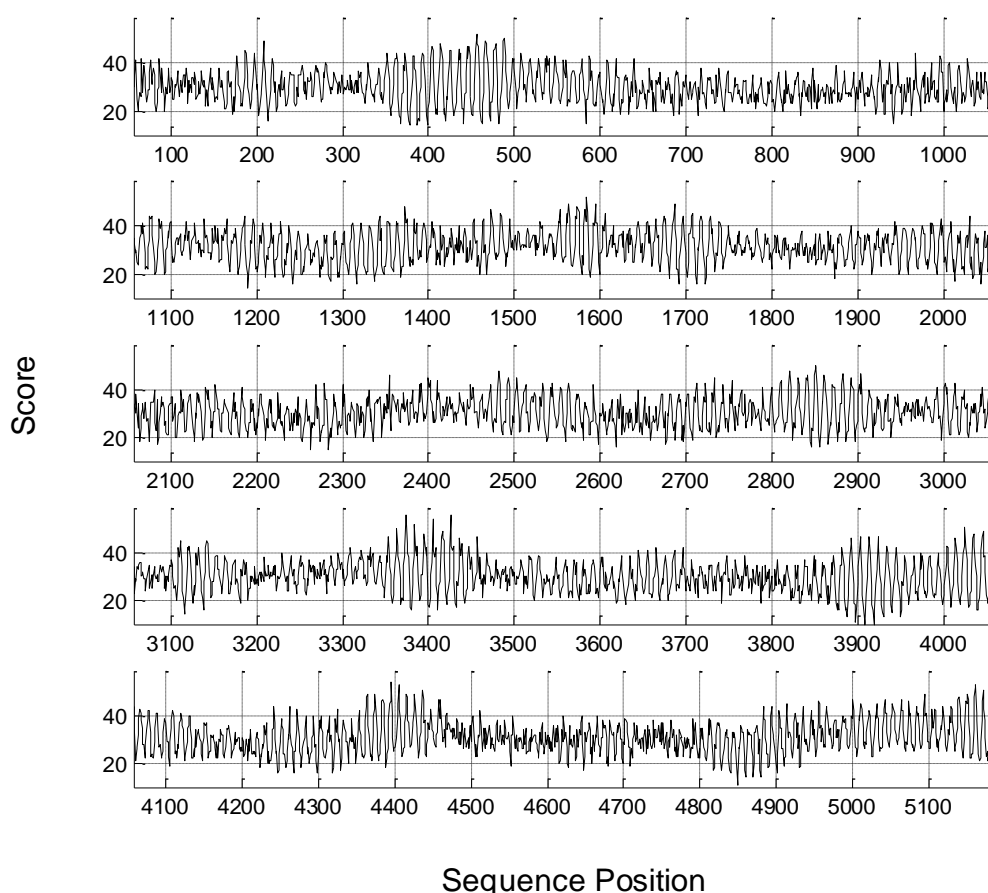
## 3. Results and Discussion

### 3.1. Sequence evidence in favor of columns in SV40 chromatin

As it follows from crystal data on the 601 clone nucleosomes [24], the YR•YR stacks of the nucleosome DNA are located in positions “minor groove *in*”. They interact with arginines of the histones and, thus, serve as “anchors” uniquely determining the inner side of the bound nucleosome

DNA. The special role the YR dinucleotides should play in the nucleosome positioning has been advocated by Zhurkin and his colleagues since 1979 [25–28].

The YR elements of the clone 601 form a “track” of CG, 5TA and 3TG separated from one another by integer number of bases corresponding to one to three periods of nucleosome DNA of average value 10.4 bases (10, 11, 21 and 30 bases). Such YR track, if found in DNA sequence, would correspond to region bound to histone octamers. The periodically arranged YR elements would keep DNA in specific rotational setting and provide stability to the nucleosomes and columns, in addition to the contribution of the alternating RRRRRYYYYY pattern.



**Figure 1. Nucleosome map of SV40 chromosome calculated with the (RRRRYYYYY)<sub>n</sub> probe (Tripathi et al., 2015, <http://strn-nuc.haifa.ac.il:8080/mapping/home.jsf>). The amplitudes correspond to the simple dinucleotide match counts of respective sequence segments to the sequence probe.**

In Figure 1 the map of the SV40 chromatin DNA periodicity is shown derived by the application of the (RRRRYYYYY)<sub>n</sub> mapping to the sequence. The regions with 10–11 base oscillations correspond to either nucleosomes or to their tight oligomers, columns. The largest RR/YY periodic region spans ~540 bases (sequence coordinates 4800–5243 and further to ~100, over the circular genome sequence start). Numerous peaks of the map indicate the locations of the pseudodyads, central points of the calculated nucleosome positions, separated one from another by

10–11 bases. These are positions “minor groove *out*” [9]. Respectively, the minima indicate positions “minor groove *in*” where the YR elements would be preferentially located. The sections of the map where the RR/YY periodicity is not obvious, however, still show the periodic distribution of YR dinucleotides following one another at one- to three-period distances, as in the clone 601. Altogether, the periodically distributed RR/YY and YR elements make long regions, which would correspond to the columnar chromatin structures.

In Figure 2 the full map of the YR-tracks identified in the SV40 genome is shown. The tracks are of various sizes, from 31 bases (track 17) to 725 bases (track 9). The tracks longer than tight dinucleosome DNA size, ~250 bases [5], occupy together about half of the SV40 genome (2564 bases). The longest track 9 would correspond to the tight oligonucleosomes involving 5 to 6 units ~125 bases each [5], stacked together in one column.

The construction of the tracks from the sequence positions of YR elements and their match to positions of minima in the RR/YY nucleosome map allowed for the gaps between the YR 10–11-mers of the size two or three periods (20–22 and 30–33 bases) as in the 601 clone nucleosome. The larger gaps between the tracks would mean the discontinuity in the columnar organization. Remarkably, however, there seems to be no discontinuity, since the sizes of the observed gaps are all combinations of the 10-mers and 11-mers (see Table 1). That is, although the 10–11-mers of the gaps are not decorated by the YR elements at their starts, length-wise they can be accommodated to the column, thus, fusing the YR tracks in a single long structure - a continuous genome size column.

**Table 1. Gaps of ordinary sequences between the YR tracks in SV40.**

Gap size(bases)	Number of cases
40 ( $4 \times 10$ )	7
41 ( $3 \times 10 + 11$ )	1
42 ( $2 \times 10 + 2 \times 11$ )	2
43 ( $10 + 3 \times 11$ )	2
44 ( $4 \times 11$ )	2
54 ( $10 + 4 \times 11$ )	1
55 ( $5 \times 11$ )	3
60 ( $6 \times 10$ )	3
66 ( $6 \times 11$ )	2

Perhaps it is not a mere coincidence that the starting 8 bases of the Figure 2 and ending 3 bases together make 11 bases, which means that the whole genome consists of 10-mers and 11-mers, as the continuous columnar structure would suggest. It is quite possible that the long smooth 10 nm filaments observed by electron microscopy in vitrified samples of SV40 chromatin [14], indeed, correspond to the columns as suggested by periodic distribution of the YR elements in the nucleotide sequence (this work). Moreover, as EM of vitrified samples of metaphase arrested eukaryotic cells suggests, chromosomes seem to be “formed by the compact association of 11 nm filaments, or

portions thereof” [29]. In other words, it may well be that the columnar organization of chromatin is common feature of eukaryotic chromosomes in general, not just of the SV40 minichromosomes.

### 3.2. Relation to experimental nucleosome mapping data

The overlapping nucleosome positions in [4] are all confined to the long YR track regions (Figure 2), though few are located in approximate nucleosome size tracks: track 1 which actually contains 2 overlapping MNase nucleosomes (at positions 221 and 247), track 7 (nucleosome at 1768), track 12 (3762), track 14 (4374 and 4392) and track 16 (4876). The frequent overlapping of the nucleosomes defined by ~145 base fragments resulting from MNase digest suggests that the “nucleosomes” may center at any peak of the RR/YY map with as many alternative positions as number of the peaks. In this sense they may slide, in discrete steps, all along the DNA sequence of the column, making series of overlapping “nucleosomes”. The longest YR track 9 harbors at least three pairs of such sliding nucleosomes (2198, 2274; 2495, 2588; and 2734, 2752). The authors of [4] conclude: “nucleosomes do not occupy unique positions in SV40 minichromosomes”.

Since practically all sequence territory of the SV40 minichromosome is covered by the periodic YR-tracks (and RR/YY oscillations) it is not surprising that practically all nucleosomes experimentally identified in [4] are located within the periodic regions. The experimental accuracy ( $\pm$  ~5 bases) does not allow to observe all the alternative positions of the nucleosome centers which should be separated by 10–11 bases. More accurate mapping does show the alternatives [11] for the nucleosomes which cover the unique BamHI restriction site. The site is, indeed, located within the periodic YR track 11. The corresponding RR/YY map shows about 10 alternative center positions for the nucleosomes covering the site (coordinates 2450–2600). In view of the above the term “nucleosome”, actually, describes any one of the particles, products of MNase digestion of the columnar structure, centered at one of ~400 sequence locations (according to the peaks of the nucleosome map).

## 4. Conclusion

The RR/YY nucleosome map of the SV40 minichromosome and the periodic distribution of YR dinucleotides along the SV40 genome, similar to the distribution of these elements in the strongest known 601 clone nucleosome, suggests that the SV40 chromatin consists of long (perhaps, even full-length) continuous columnar structures, rather than of individual beads-on-string nucleosomes.

1-GCCTCGGC	GCTGCAATTT	CCAGGAATGGC	GTGAATTTAT
CTCTGCATAAA	452-TGTGAAGGGG	921-TGTAGATTTG	TGAAAAATTTG
<b>Track 18</b>	AAGATACTGT	931-TATAGGCCAG	AGGCTCCTGG
TAAAAAAAT	472-TGACGGGAAA	ATGATTACTA	TGGTGCAAATC
TAGTCAGCCA	482-CGCAAAAAC	951-TGATATTTTAT	AAAGAACTGCT
TGGGCGGAGA	CAGAAAGGTT	TTCTGGAGTA	CCTCAGTGGAT
ATGGCGGAAC	AACTGAAAAAC	973-CAAACCTTTGT	GTTGCCTTTAC
62-TGGGCGGAGT	CAGAAAGTTA	TCACAGTGTT	TTCTAGGCCTG
TAGGGCGGGGA	ACTGTAAGT	CAGTATCTTG	<b>Track 5</b>
83-TGGGCGGAGT	TTAGTCTTTT	ACCCAGACAT	1468-TAGTGTGCGAA
TAGGGCGGGG	TGTCTTTTAT	1015-TGGGTCCAA	TACTTCTGCTC
ACTATGGTTG	TTCAGGTCCA	CCTTTTAA	1490-TAAAAGCTTAT
CTGACTAATT	TGGTGCTGC	TGCCATTCT	GAAGATGGCCC
GAGATGCATG	TTAACACTGT	CAGCTTTT	CAACAAAAAGA
CTTGCATAC	584-TGGGGACCT	GGCGTGAATA	AAAGGAAGTGT
TTCTGCCTGC	AATTGCTACTG	CAAAATGACA	TCCAGGGCAG
<b>Track 1</b>	605-TGTCTGAAGC	TTCTAGGCTC	CTCCAAAAAAA
TGGGAGCCT	615-TGCTGCTGCT	ACCTCACAGGA	CCAAAGGAACC
GGGACTTTC	ACTGGATTTT	GCTTGAAAGAA	AGTGCAAGTGC
173-CACACCTGGT	635-CAGTAGCTGA	GAACCCAAAGA	<b>Track 6</b>
183-TGCTGACTAAT	AATTGCTGCTG	<b>Track 4</b>	1578-CAAAGCTCGT
194-TGAGATGCAT	GAGAGCCGC	TATTTAAGGGA	CATAAAAGGA
GCTTTGCATA	666-TGCTGCAATG	CAGTTTGGA	GGAATAGAAG
CTTCTGCCTGC	AAGTGCAACT	AGGTTTTTAGA	TTCTAGGAGT
225-TGGGAGCCT	687-TGCATCTGTT	GGAACTACT	TAAAAGTGGAG
GGGACTTTC	GCTACTGTTG	TGACAGTAA	TAGACAGCTTC
CACACCTAAC	AAGGCCAAC	TTAATGCTCC	ACTGAGGTGG
TGACACACAT	AACCTCTGAG	TGTTAATTGG	AGTGCTTTTT
TCCACAGCTG	GCAATGCTG	1192-TATAACTCTT	AAATCCTCAA
GTTCTTTCCG	CTATAGGCCT	1202-TACAAGATTAC	TGGCAATCC
CCTCAGAAGG	CCTCCACAGG	TACTCTACTT	<b>Track 7</b>
TACCTAACCA	CCTATGCTGTG	1223-TGCTCCCAT	1681-TGATGAACAT
AGTTCCTCTT	ATATCTGGGGC	1233-TAGGCCTACAA	1691-CAAAAAGGCTT
TCAGAGGTTA	TCCTGCTGCTA	1244-TGGTGAGACA	AAGTAAAAGCT
TTTCAGGCCA	<b>Track 3</b>	AGTAGCCAACA	1713-TAGCAGCTGAA
<b>Track 2</b>	TAGCTGGATT	GGGAAGGGTTG	AAACAGTTTA
TGGTGCTGCG	TGAGCTTTTAC	CAAAATATCATT	1734-CAGATGACTCT
CCGGCTGTCA	812-TGCAAACTGTG	1287-TGGCACACC	CCAGACAAAG
356-CGCCAGGCCTC	ACTGGTGTGAG	1297-TATGATAATAT	AACAACTGCCT
367-CGTTAAGGTT	834-CGCTGTTGCT	1308-TGATGAAGCA	TGCTACAGTG
CGTAGGTCATG	CAAGTGGGGTA	GACAGTATTC	TGGCTAGAATT
GACTGAAAGTA	TAGATTTTTTA	AGCAAGTAAC	CCTTTGCCTAA
AAAAACAGCT	GTGACTGGGAT	1338-TGAGAGGTGGG	TTTAAATGAGG
410-CAACGCCTTTT	CACAAAGTTTC	AAGCTCAAAGC	ACTTAACTTG
421-TGTGTTTGT	TACTGTTGGTT	1360-CAAGTCTTAA	<b>Track 8</b>
432-TAGAGCTTTT	899-TATATCAACAA	TGTGCAGTCAG	TGAAATATTT

TGATGTGGAA	TGTTGACATT	GACTGTGAGGA	TATTTGAGTTC
GCTGTTACTGT	TGTGGGCTGTT	CTGAGGGGCTT	TTTTATTTAGG
TAAAACTGAG	2317-TACCAACTT	GAAATGAGCCT	TGTTTCTTTTC
GTTATTGGGG	CTGGAACACAG	<b>Track 10</b>	TAAGTTTACCT
1872-TAACTGCTATG	CAGTGAAGG	2815-TGGACTGTG	TAACTACTGCCA
TTAAACTTGCA	GACTTCCCAGA	AATCAATGCC	TCCAAAATAATC
TTCAGGGACA	TATTTTAAAAAT ○	2835-TGTTTCATGCC	CCTTAAATTGT ○
CAAAAACTCA	TACCCTTAGA	CTGAGTCTTC	CCAGGTATTA
TGAAAATGGTG	AAGCGGTCTG	2856-CATGTTCTTCT	ATTCCTTGACC
CTGAAAACC	TGAAAACCCC	CCCACCATCT	TGAAGGCAAAAT
CATTCAAGGGT	TACCAATTT	TCATTTTAT	CTCTGGACTCC
1947-CAATTTTCAT	CCTTTTGTTA	2888-CAGCATTTTC	CCTCCAGTGCC
TTTTTTGCTGT	AGTGACCTAAT	CTGGCTGTCT	CTTTACATCCT
TGTTGGGGAAC	TACAGGAGG	TCATCATCAT	<b>Track 11</b>
CTTTGGAGCTG	ACACAGAGGG	2918-CATCACTGTTT	CAAAAACTAC
1991-CAGGGTGTGTT	TGATGGGCA	CTTAGCCAATC	3411-TAAAACTGGT
AGCAAATACA	GCCTATGATTG	2940-TAAAACTCAA	3422-CAATAGCTAC
GGACCAAATAT	GAATGTCCTCT	TTCCCATAGC	TCCTAGCTCA ○
CCTGCTCAAAC	2486-CAGTAGAGG ○	CACATTAAACT	AAGTTCAGCC
	AGGTTAGGGTT	TCATTTTGTGA	3452-TGTCCAAGGG
<b>Track 9</b>	2507-TATGAGGACAG	TACACTGACAA	CAAATTAACA
2035-TGTAACCCCA	2517-CAGAGGAGCT	ACTAAACTCTT	TTTAAAGCTT ○
AAAAATGCTA	TCCTGGGGATC	TGTCCAATCT	TCCCCCACA
2055-CAGTTGACAGT	CAGACATGATA	CTCTTCCAC	TAATTCAGC
2066-CAGCAGATGAA	AGATACATTGA	TCCACAATTC ○	AAAGCAGCTGC
2077-CACTGACCAC	2560-TGAGTTTGA	TGCTCTGAAT	TAATGTAGTTT
AAGGCTGTTT	CAAACCACAAC	ACTTTGAGCA	3524-TACCACTATCA
TGATAAGGA	2581-TAGAATGCAG ○	AACTCAGCCA	ATTGGTCCTT
2107-TAATGCTTATC	TGAAAAAATG	CAGGTCTGTAC	TAAACAGCCAG
CAGTGGAGTGC	CTTTATTTGT	CAAATTAACAT	TATCTTTTTT
2129-TGCTGAGGTT	GAAATTTGTGA	AAGAAGCAAAG	TAGGAATGTTG
TCCAAGTAAA	TGCTATTGCTT	CAATGCCACTT	3577-TACACCATGCA
AATGAAAACAC	TATTTGTAACC	3109-TGAATTATTC	TTTTAAAAAGT
TAGATATTTT	ATTATAAGCTG	TCTTTTCTAA	CATACACCAC
GGAACCTACA	CAATAACAAG	CAAAAACTCAC	TCAATCCATTT
2181-CAGGTGGGGA	TTAACAACAA	3140-TGCGTTCCAGG	3620-TGGCAACAAA
AAATGTGCCT ○	2677-CAATTGCATT	CAATGCTTTAA	3631-CAGTGTAGCC
CCTGTTTTGCA	2687-CATTTTATGT	ATAATCTTTG ○	AAGCAACTCC
2212-CATTACTAAC	TTCAGGTTCA	GGCCTAAAATC	AGCCATCCAT
ACAGCAACCA	GGGGGAGGTG	3183-TATTTGTTTTA	TCTTCTATGT ○
2232-CAGTGCTTCTT	TGGGAGGTTT	CAATCTGGCC	<b>Track 12</b>
GATGAGCAGGG	TTTAAAGCAAG ○	TGCAGTGTTT	3671-CAGCAGAGCC
2254-TGTTGGGCCCT	TAAAACCTCTA	TAGGCACACTG	TGTAGAACCA
2265-TGTGCAAAGC ●	CAAAATGTGGTA ○	TACTCATTC	AACATTATAT
2275-TGACAGCTTG	TGGCTGATTAT	TGTTGACTATT	CCATCCATC
2285-TATGTTTCTGC	GATCATGAACA	CCAGGGGGAAA	CAAAAGATCAT

TA	AATCTGTT		TA	ATAGCAGA		ATCTCTGTAGG		AGAAGGTCCAT		
TC	TAAACATT		CA	CTCTATGCC		TA	GTTTGTCC	<b>Track 18</b>		
3743-TC	TTCTCTAGT		TG	TGTGGAGTA		AATTATGTCA		TA	GCTGCAAA	
TA	ATTGTAGGC	○	AG	AAAAAACAG		CA	CCCAGAAGA		GATTCCTCTCT	
TA	FCAACCCG		4236-TA	TGTTATGAT		TA	AGGTCCTT		GTTTAAACTT	
CT	TTTTAGCT		4247-TA	TAACTGTTA		CA	CAAAGATC	●	5157-TA	TCCATCTTT
AAA	CAGTAT		4258-TG	CCTACTTA		AAGTCCAAAC			GCAAAGCTTTT	
CA	ACAGCCTGT		4268-TA	AAGTTAC		CA	CATTCTAAA		5179-TG	CAAAAGCC
TG	GCATATGG		AGA	ATATTTT		GCAATCGAAG			TA	GGCTCCA
TTTT	TTGGTT		TCC	ATAATTT		CA	GTAGCAAT	○	AAAA	AGCCTC
TT	TGCTGTCA		TCT	TGTATAG		CA	ACCCACAC		CTC	ACTACTC
GCA	AATATAG		<b>Track 14</b>			AAGTGGATCTT		5220-TG	GAATAGCT	
<b>Track 13</b>			CA	GTCAGCT		TCCTGTATAAT		5230-CA	GAGGCCGAG	
CA	GCAATTGCA		TTTT	CCTTTG		TTTCTATTTT		GCG	-5243	
3856-TA	ATGCTTTT		TG	GTGTAAT		<b>Track 16</b>				
CA	TGGTACTTA		AG	CAAAGCAA		CA	TGCTTCAT			
3877-TA	GTTGGCTGGG		GCA	AGAGTTC		CCT	CAGTAAG			
CT	GTTCTTTT	○	TA	TACTAAA		CA	CAGCAAGCA			
3899-TA	ATACATTT		4368-CA	CAGCATGA	●	4828-TA	TGCAGTTAG			
3909-TA	AACACATTT		CT	CAAAAACT		CA	GACATTTTC			
3920-CA	AAACTGTA		4389-TA	GCAATTCTG	●	TTT	GCACACT			
CT	GAAATTC		AAG	GAAAGTCC		4860-CA	GCCATTG			
AAG	TACATCC		TT	GGGTCTTC		TTT	GCAGTAC	●		
3950-CA	AGCAATAA		4422-TA	CCTTCTCT		ATT	GCATCAA			
3960-CA	ACACATCAT		TCT	TTTTTTGG		4890-CA	CCAGGATT			
3971-CA	CATTTTGT		AG	GAGTAGAA		TA	AGGAAGAAG			
TT	CATTGCA		TG	TTGAGAGT		4911-CA	AATACCTC			
TA	CTCTGTTA		CA	GCAGTAGC		AGT	TGCATCC			
CA	AGCTTCCA		CT	CATCATCA		4931-CA	GAGCCTC			
GG	ACTTGTT	○	CT	AGATGGCA		CA	AAGTCAGGT			
TA	GTTTCTCT		TTT	CTTCTGA		4952-TG	ATGAGCATA			
TG	CTTCTCT		GCA	AAACAGG		TTTT	ACTCCA			
GG	ATAAAAT		TTTT	CCTCAT		TCT	TCCATTTT			
4052-CA	TGCTCCTTT		<b>Track 15</b>			CT	TGTACAGAG			
AACC	CACCTGG		TA	AAGGCATT		TA	TTCATTTTC			
4074-CA	AACTTTCT		CC	ACCACTGC		TT	CATTTTTC			
CA	ATAACAGAA	○	TCC	CATTCAT		TT	CATCTCCTC			
AAT	GATCTCT	○	4553-CA	GTTCCATAG		CT	TATCAGGA			
AGT	CAAGGCAC		GTT	GGAATCTA		<b>Track 17</b>				
TA	TACATCAA		AA	ATACACAAA		TG	AACTCCT			
TA	TTCCTTAT		CA	ATTAGAAT		TG	CATTTTTT			
TA	ACCCCTTA		CA	GTAGTTTAA		TA	AATATGCCT			
CA	AATTAATA		CA	CATTATACA		TT	CTCATCAGA			
AG	CTAAAGGTA		CT	TAAAAATTT		GG	AATATCC			
CA	CAATTTTG		TA	TATTACCT		CC	AGCACTCC			
AG	CA	TAGTTAT	TA	GAGCTTTAA	○	TT	CAAGACCT			



**Figure 2.** The YR tracks of the SV40 genome, in format 10 or 11 bases in line. The numbers on the left correspond to the sequence positions of the map minima, i. e., positions “minor groove *in*”. Small circles on the right show approximate positions (centers) of strong (●) and moderate (○) nucleosomes experimentally mapped in (Ambrose et al., 1990). Note that some 10-11-mers start with YR, while others are ordinary sequence segments of the same lengths.

### Conflict of Interest

All authors declare that there are no conflicts of interest.

### References

1. Griffith JD (1975) Chromatin structure: deduced from a minichromosome. *Science* 187: 1202–1203.
2. Bellard M, Oudet P, Germond JE, et al. (1976) Subunit structure of simian-virus-40 minichromosome. *Eur J Biochem* 70: 543–553.
3. Mengeritsky G, Trifonov EN (1984) Nucleotide sequence-directed mapping of the nucleosomes of SV40 chromatin. *Cell Bioph* 6: 1–9.
4. Ambrose C, Lowman H, Rajadhyaksha A, et al. (1990) Location of nucleosomes in simian virus 40 chromatin. *J Mol Biol* 214: 875–884.
5. Tatchell K, van Holde KE (1978) Compact oligomers and nucleosome phasing. *Proc Natl Acad Sci USA* 75: 3583–3587.
6. Fajkus J, Trifonov EN (2001) Columnar packing of telomeric nucleosomes. *Biochem Biophys Res Comm* 280: 961–963.
7. Salih B, Trifonov EN (2015) Strong nucleosomes of *A. thaliana* concentrate in centromere regions. *J Biomol Str Dyn* 33: 10–13.
8. Salih B, Trifonov EN (2015). Strong nucleosomes reside in meiotic centromeres of *C. elegans*. *J Biomol Str Dyn* 33: 365–373.
9. Nibhani R, Trifonov EN (2015) Reading sequence-directed computational nucleosome maps. *J Biomol Str Dyn* 33:1558–1566.
10. Gu SG, Goszczynski B, McGhee JD, et al. (2013) Unusual DNA packaging characteristics in endoreduplicated *Caenorhabditis elegans* oocytes defined by in vivo accessibility to an endogenous nuclease activity. *Epigenet Chromatin* 6: 37.
11. Ponder BAJ, Crawford LV (1977) Arrangement of nucleosomes in nucleoprotein complexes from polyoma-virus and SV40. *Cell* 11: 35–49.
12. Zhang T, Talbert PB, Zhang W, et al. (2013) The CentO satellite confers translational and rotational phasing on cenH3 nucleosomes in rice centromeres. *Proc Natl Acad Sci U S A* 110: E4875–4883.
13. Rattner JB, Hamkalo BA (1978) Higher-order structure in metaphase chromosomes *Chromosoma* 69: 373–379.
14. Dubochet J, Adrian M, Schultz P, et al. (1986) Cryo-electron microscopy of vitrified SV40 minichromosomes: the liquid drop model. *EMBO J* 5: 519–528.
15. Wanner G, Formanek H (2000) A new chromosome model. *J Struct Biol* 132: 147–161.
16. Karpov VL, Bavykin SG, Preobrazhenskaya OV, et al. (1982) Alignment of nucleosomes along DNA and organization of spacer DNA in drosophila chromatin. *Nucl Acids Res* 10: 4321–4337.

17. Widom J (1992) A relationship between the helical twist of DNA and the ordered positioning of nucleosomes in all eukaryotic cells. *Proc Natl Acad Sci U S A* 89:1095–1099.
18. Rapoport AE, Frenkel ZM, Trifonov EN (2011) Nucleosome positioning pattern derived from oligonucleotide compositions of genomic sequences. *J Biomol Str Dyn* 28: 567–574.
19. Frenkel ZM, Bettecken T, Trifonov EN (2011) Nucleosome DNA sequence structure of isochores. *BMC Genomics* 12: 203.
20. Tripathi V, Salih B, Trifonov EN (2015) Universal full-length nucleosome mapping sequence probe. *J Biomol Str Dyn* 33: 666–673.
21. Lowary PT, Widom J (1998) New DNA sequence rules for high affinity binding to histone octamer and sequence directed nucleosome positioning. *J Mol Biol* 276: 19–42.
22. Trifonov EN, Nibhani R (2015) Review fifteen years of search for strong nucleosomes. *Biopolymers*, 103: 432–437.
23. Nibhani R, Trifonov EN, TA-periodic (“601”-like) centromeric nucleosomes of *A.thaliana*. *J Biomol Str Dyn* [in press].
24. Vasudevan D, Chua EYD, Davey CA (2010) Crystal structures of nucleosome core particles containing the '601' strong positioning sequence. *J Mol Biol* 403: 1–10.
25. Zhurkin VB (1982) Periodicity in DNA primary structure and specific alignment of nucleosomes. *Stud Biophys* 87: 151–152.
26. Zhurkin VB (1983) Specific alignment of nucleosomes on DNA correlates with periodic distribution of purine pyrimidine and pyrimidine purine dimers. *FEBS Lett* 158: 293–297.
27. Zhurkin VB, Lysov YP, Ivanov VI (1979) Anisotropic flexibility of DNA and the nucleosomal structure. *Nucl Acids Res* 6: 1081–1096.
28. Wang D, Ulyanov NB, Zhurkin VB (2010) Sequence-dependent kink-and-slide deformations of nucleosomal DNA facilitated by histone arginines bound in the minor groove. *J Biomol Str Dyn* 27: 843–859.
29. McDowall AW, Smith JM, Dubochet J (1986) Cryo-electron microscopy of vitrified chromosomes in situ. *EMBO J* 5: 1395–1402.
30. Trifonov EN Nucleosome repeat lengths and columnar chromatin structure. *J Biomol Str Dyn* [in press].



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