



Research article

Optogenetic actuation in ChR2-transduced fibroblasts alter excitation-contraction coupling and mechano-electric feedback in coupled cardiomyocytes: a computational modeling study

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Supplementary

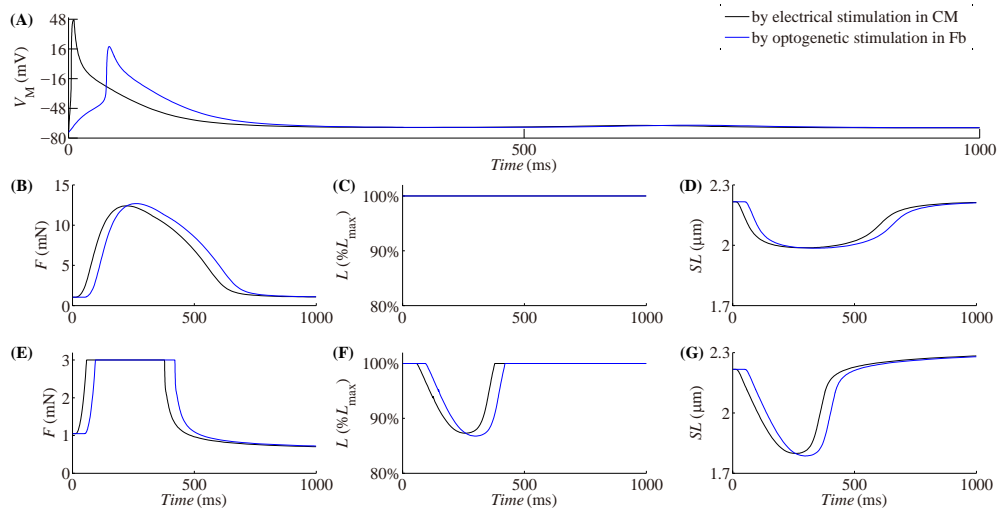


Figure S1. Membrane potentials and mechanical parameters in the CM. The electrical stimulation applied to the CM is a rectangular electrical pulse (6 ms, -15 pA/pF). The optogenetic stimulation applied to ChR2-transduced Fbs is a light pulse (50 ms, 5 mW/mm²). (A) V_M . (B-D) Time-dependent signals of isometric F , L and SL . (E-G) Time-dependent signals of isotonic F , L and SL applied an afterload of 3 mN.

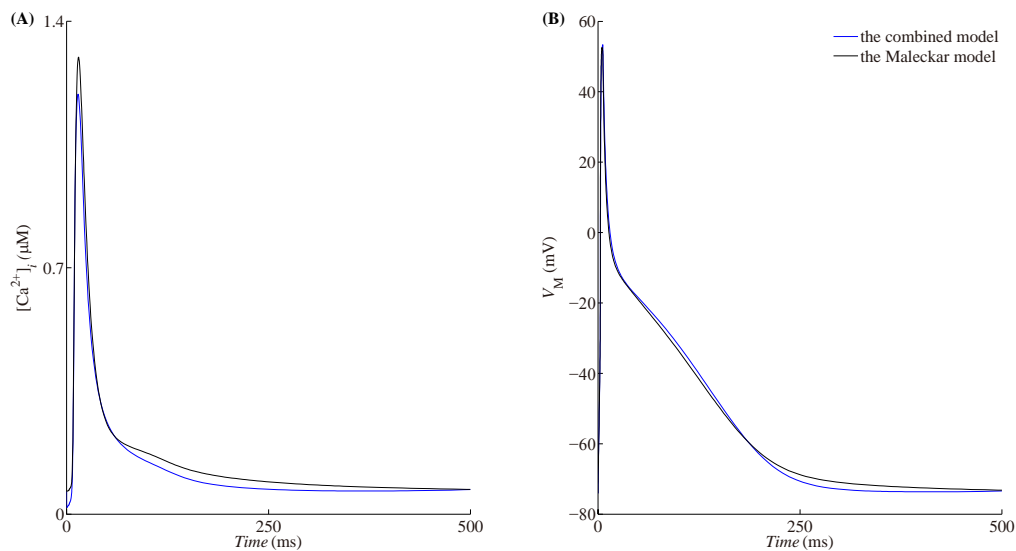


Figure S2. Illustration of $[Ca^{2+}]_i$ (A) and V_M (B) in the combined model and in the model of Maleckar et al. [1].

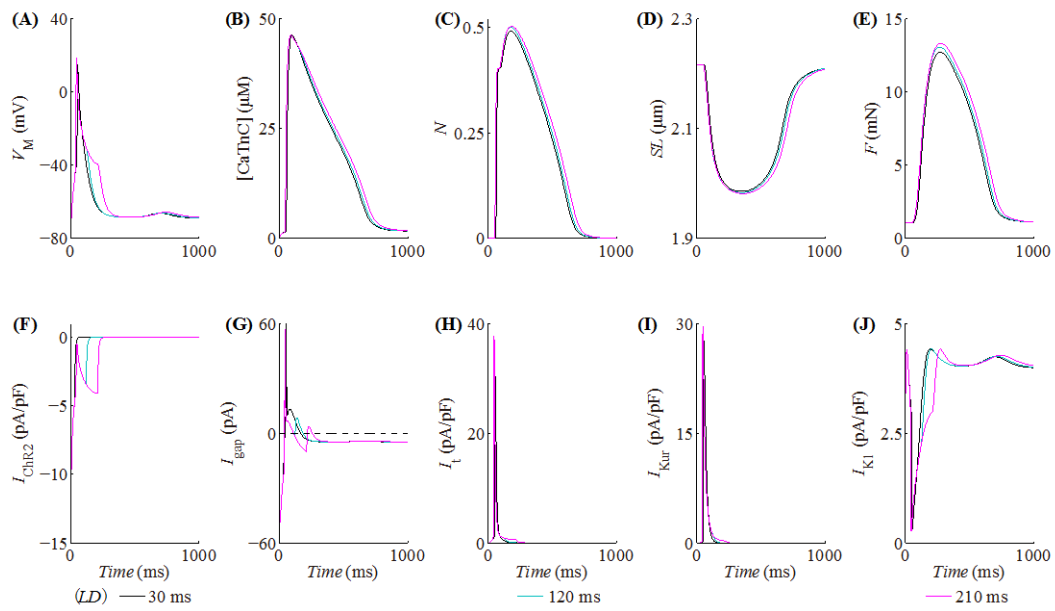


Figure S3. Electrophysiological and mechanical parameters (in isometric contraction) related to ECC at a LD of 30, 120 and 210 ms. LI is fixed to 5 mW/mm^2 .

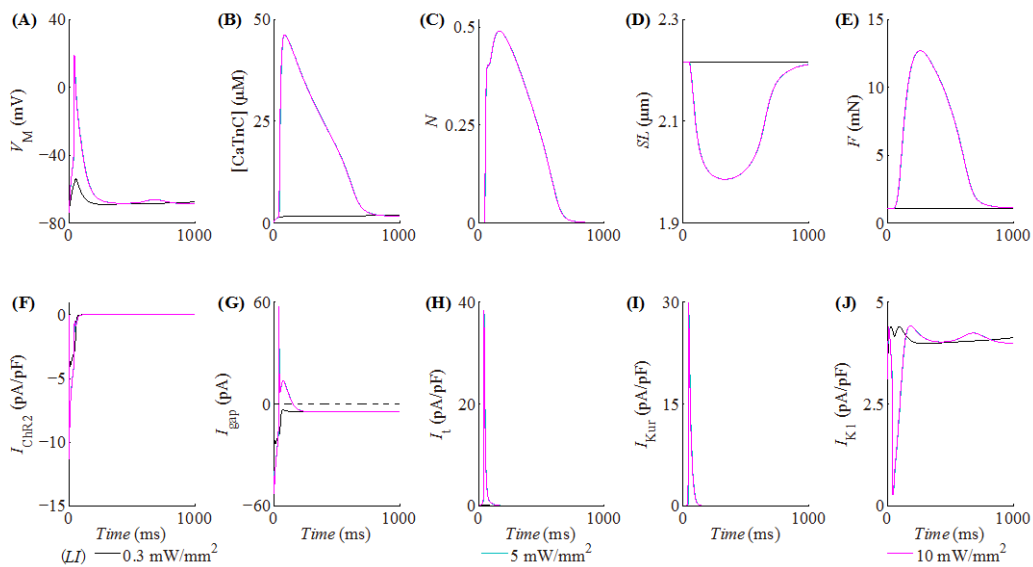


Figure S4. Electrophysiological and mechanical parameters (in isometric contraction) related to ECC at a LI of 0.3, 5 and 10 mW/mm^2 . LD is fixed to 50 ms.

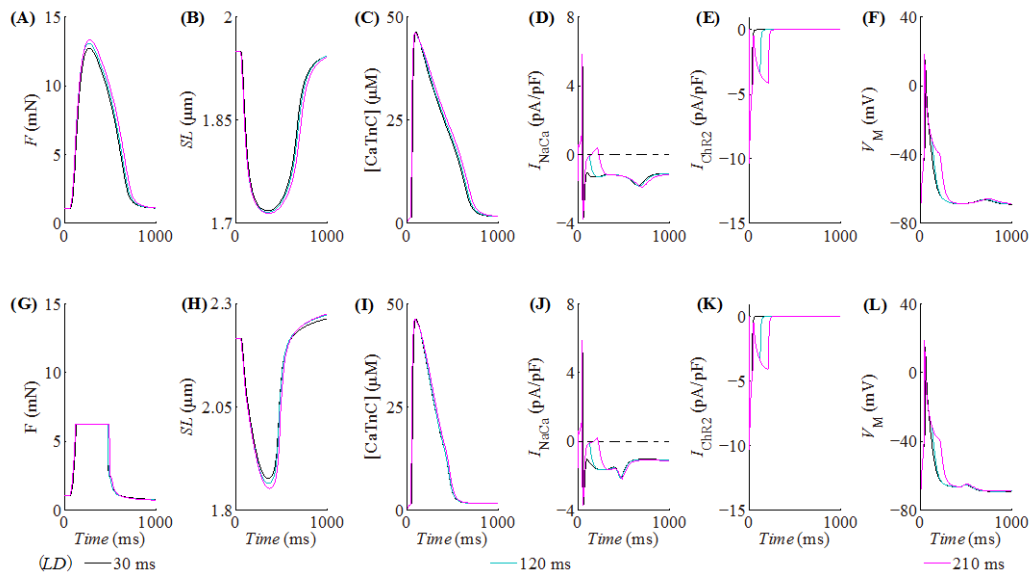


Figure S5. Electrophysiological and mechanical parameters (in isometric and isotonic contractions) related to MEF at a LD of 30, 120 and 210 ms. LI is fixed to 5 mW/mm^2 .

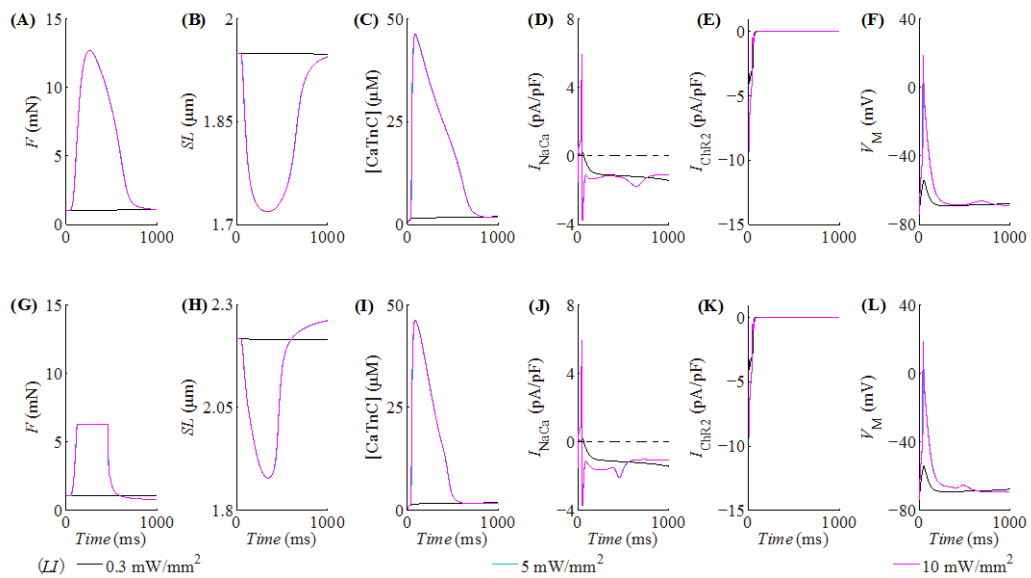


Figure S6. Electrophysiological and mechanical parameters (in isometric and isotonic contractions) related to MEF at a LI of 0.3, 5 and 10 mW/mm^2 . LD is fixed to 50 ms.

Model Equations

Table 1 through 29 contained all the equations, parameters values and initial conditions necessary to carry out the simulations presented in this manuscript. Unless otherwise noted, the units are as follows: time in second (s), voltage in millivolt (mV), concentration in millimolar (mM), current in picoampere (pA), conductance in nanosiemens (nS), capacitance in picofarads (pF), volume in nanoliters (nL), temperature in kelvin (K), and sarcomere length in micron (μm).

As described in our manuscript, the model consists of three modules: the excitation-contraction coupling (ECC) module in the cardiomyocyte (CM), the optical module in fibroblasts (Fbs) and the CM-Fb coupling module.

Module 1: ECC in CM

This module consists of two blocks: an electrophysiological block and a mechanical block. The former is based on the ‘Maleckar-Trayanova’ (MT) ionic model, and the latter is based on an updated version of the ‘Ekaterinburg-Oxford’ (EO) model [1,2]. We replace the MT’s description of Ca^{2+} buffering system with the EO’s and modify the formulation of intracellular Ca^{2+} concentration. Then we integrate the two blocks to compose the ECC module.

1) Electrophysiological block

Table 1. Na^+ current: I_{Na}

$I_{\text{Na}} = P_{\text{Na}} m^3 (0.9h_1 + 0.1h_2) [Na^+]_c V_M \frac{F^2}{RT} \frac{e^{(V_M - E_{\text{Na}})F/RT} - 1.0}{e^{V_M F/RT} - 1.0}$	
$\bar{m} = \frac{1.0}{1.0 + e^{(V_M + 27.12)/-8.21}}$	$\bar{h} = \frac{1.0}{1.0 + e^{(V_M + 63.6)/5.3}}$
$\frac{dm}{dt} = \frac{\bar{m} - m}{\tau_m}$	$\tau_m = 0.000042e^{-((V_M + 25.57)/28.8)^2} + 0.000024$
$\frac{dh_1}{dt} = \frac{\bar{h} - h_1}{\tau_{h_1}}$	$\tau_{h_1} = \frac{0.03}{1.0 + e^{(V_M + 35.1)/3.2}} + 0.0003$
$\frac{dh_2}{dt} = \frac{\bar{h} - h_2}{\tau_{h_2}}$	$\tau_{h_2} = \frac{0.12}{1.0 + e^{(V_M + 35.1)/3.2}} + 0.003$

Table 2. Ca^{2+} current: I_{CaL}

$I_{\text{CaL}} = \bar{g}_{\text{CaL}} d_L [f_{\text{Ca}} f_{L1} + (1 - f_{\text{Ca}}) f_{L2}] (V_M - E_{\text{Ca,app}})$	
$\bar{d}_L = \frac{1.0}{1.0 + e^{(V_M + 9.0)/-5.8}}$	$\bar{f}_L = \frac{1.0}{1.0 + e^{(V_M + 27.4)/7.1}}$
$\frac{dd_L}{dt} = \frac{\bar{d}_L - d_L}{\tau_{d_L}}$	$\tau_{d_L} = 0.0027e^{-((V_M + 35.0)/30.0)^2} + 0.002$

$$\begin{aligned} \frac{df_{L1}}{dt} &= \frac{\bar{f}_L - f_{L1}}{\tau_{f_{L1}}} & \tau_{f_{L1}} &= 0.161e^{-((V_M+40.0)/14.4)^2} + 0.01 \\ \frac{df_{L2}}{dt} &= \frac{\bar{f}_L - f_{L2}}{\tau_{f_{L2}}} & \tau_{f_{L2}} &= 1.3323e^{-((V_M+40.0)/14.2)^2} + 0.0626 \\ f_{Ca} &= \frac{[Ca^{2+}]_d}{[Ca^{2+}]_d + k_{Ca}} \end{aligned}$$

Table 3. Transient and ultra-rapidly delayed rectifier K^+ currents: I_t and I_{Kur}

$$\begin{aligned} I_t &= \bar{g}_t r s (V_M - E_K) \\ \bar{r} &= \frac{1.0}{1.0 + e^{(V_M-1.0)/-11.0}} & \bar{s} &= \frac{1.0}{1.0 + e^{(V_M+40.5)/11.5}} \\ \frac{dr}{dt} &= \frac{\bar{r} - r}{\tau_r} & \tau_r &= 0.0035e^{-(V_M/30.0)^2} + 0.0015 \\ \frac{ds}{dt} &= \frac{\bar{s} - s}{\tau_s} & \tau_s &= 0.025635e^{-((V_M+52.45)/15.89)^2} + 0.01414 \\ I_{Kur} &= \bar{g}_{Kur} r_{Kur} s_{Kur} (V_M - E_K) \\ \bar{r}_{Kur} &= \frac{1.0}{1.0 + e^{(V_M+6.0)/-8.6}} & \bar{s}_{Kur} &= \frac{1.0}{1.0 + e^{(V_M+7.5)/10.0}} \\ \frac{dr_{Kur}}{dt} &= \frac{\bar{r}_{Kur} - r_{Kur}}{\tau_{r_{Kur}}} & \tau_{r_{Kur}} &= \frac{0.009}{1.0 + e^{(V_M+5.0)/12.0}} + 0.0005 \\ \frac{ds_{Kur}}{dt} &= \frac{\bar{s}_{Kur} - s_{Kur}}{\tau_{s_{Kur}}} & \tau_{s_{Kur}} &= \frac{0.59}{1.0 + e^{(V_M+60.0)/10.0}} + 3.05 \end{aligned}$$

Table 4. Delayed rectifier K^+ currents: $I_{K,s}$ and $I_{K,r}$

$$\begin{aligned} I_{K,s} &= \bar{g}_{K,s} n (V_M - E_K) \\ \bar{n} &= \frac{1.0}{1.0 + e^{(V_M-19.9)/-12.7}} & \tau_n &= 0.7 + 0.4e^{-((V_M-20.0)/20.0)^2} \\ \frac{dn}{dt} &= \frac{\bar{n} - n}{\tau_n} \\ I_{K,r} &= \bar{g}_{K,r} p_a p_i (V_M - E_K) \\ \bar{p}_a &= \frac{1.0}{1.0 + e^{(V_M+15.0)/-6.0}} & p_i &= \frac{1.0}{1.0 + e^{(V_M+55.0)/24.0}} \\ \frac{dp_a}{dt} &= \frac{\bar{p}_a - p_a}{\tau_{p_a}} & \tau_{p_a} &= 0.03118 + 0.21718e^{-((V_M+20.1376)/22.1996)^2} \end{aligned}$$

Table 5. Inward rectifier K^+ currents: I_{K1}

$$I_{K1} = \bar{g}_{K1} [K^+]_c^{0.4457} \frac{V_M - E_K}{1.0 + e^{1.5(V_M - E_K + 3.6)F/RT}}$$

Table 6. Background inward currents: $I_{B,Na}$ and $I_{B,Ca}$

$$I_{B,Na} = \bar{g}_{B,Na}(V_M - E_{Na})$$

$$I_{B,Ca} = \bar{g}_{B,Ca}(V_M - E_{Ca})$$

Table 7. Pump and exchanger currents: I_{NaK} , I_{CaP} , and I_{NaCa}

$$I_{NaK} = \bar{I}_{NaK} \frac{[K^+]_c}{[K^+]_c + k_{NaK,K}} \cdot \frac{[Na^+]_i^{1.5}}{[Na^+]_i^{1.5} + k_{NaK,Na}^{1.5}} \cdot \frac{V_M + 150.0}{V_M + 200.0}$$

$$I_{CaP} = \bar{I}_{CaP} \frac{[Ca^{2+}]_i}{[Ca^{2+}]_i + k_{CaP}}$$

$$I_{NaCa} = k_{NaCa} \frac{[Na^+]_i^3 [Ca^{2+}]_c e^{\gamma V_M F/RT} - [Na^+]_c^3 [Ca^{2+}]_i e^{(\gamma-1.0)V_M F/RT}}{1.0 + d_{NaCa}([Na^+]_c^3 [Ca^{2+}]_i + [Na^+]_i^3 [Ca^{2+}]_c)}$$

Table 8. Intracellular ion concentrations: $[Na^+]_i$, $[K^+]_i$, and $[Ca^{2+}]_i$

$$\frac{d[Na^+]_i}{dt} = -\frac{I_{Na} + I_{B,Na} + 3I_{NaK} + 3I_{NaCa}}{Vol_i F}$$

$$\frac{d[K^+]_i}{dt} = -\frac{I_t + I_{Kur} + I_{K1} + I_{K,s} + I_{K,r} - 2I_{NaK}}{Vol_i F}$$

$$\frac{d[Ca^{2+}]_i}{dt} = -\frac{-I_{di} + I_{B,Ca} + I_{CaP} - 2I_{NaCa} + I_{up} - I_{rel}}{2Vol_i F} - \left(\frac{d[CaTnC]}{dt} + \frac{dB_1}{dt} + \frac{dB_2}{dt} \right)$$

$$\frac{d[Ca^{2+}]_d}{dt} = -\frac{I_{CaL} + I_{di}}{2.0 Vol_d F}$$

$$I_{di} = ([Ca^{2+}]_d - [Ca^{2+}]_i) \frac{2F Vol_d}{\tau_{di}}$$

Table 9. Cleft space ion concentrations: $[Na^+]_c$, $[K^+]_c$, and $[Ca^{2+}]_c$

$$\frac{d[Na^+]_c}{dt} = \frac{[Na^+]_b - [Na^+]_c}{\tau_{Na}} + \frac{I_{Na} + I_{B,Na} + 3I_{NaK} + 3I_{NaCa}}{Vol_c F}$$

$$\frac{d[K^+]_c}{dt} = \frac{[K^+]_b - [K^+]_c}{\tau_K} + \frac{I_t + I_{Kur} + I_{K1} + I_{K,s} + I_{K,r} - 2I_{NaK}}{Vol_c F}$$

$$\frac{d[Ca^{2+}]_c}{dt} = \frac{[Ca^{2+}]_b - [Ca^{2+}]_c}{\tau_{Ca}} + \frac{I_{CaL} + I_{B,Ca} + I_{CaP} - 2I_{NaCa}}{2.0 Vol_c F}$$

Table 10. Intracellular Ca^{2+} buffering

$$\frac{dB_1}{dt} = b_{1on} \cdot (B_{1tot} - B_1) \cdot [Ca^{2+}]_i - b_{1off} \cdot B_1$$

$$\frac{dB_2}{dt} = b_{2on} \cdot (B_{2tot} - B_2) \cdot [Ca^{2+}]_i - b_{2off} \cdot B_2$$

Table 11. Ca^{2+} handling by the sarcoplasmic reticulum

$$I_{\text{up}} = \bar{I}_{\text{up}} \frac{[\text{Ca}^{2+}]_i / k_{\text{cyca}} - k_{\text{xcs}}^2 [\text{Ca}^{2+}]_{\text{up}} / k_{\text{srca}}}{([\text{Ca}^{2+}]_i + k_{\text{cyca}}) / k_{\text{cyca}} + k_{\text{xcs}} ([\text{Ca}^{2+}]_{\text{up}} + k_{\text{srca}}) / k_{\text{srca}}}$$

$$I_{\text{tr}} = ([\text{Ca}^{2+}]_{\text{up}} - [\text{Ca}^{2+}]_{\text{rel}}) \frac{2F \text{Vol}_{\text{rel}}}{\tau_{\text{tr}}}$$

$$I_{\text{rel}} = \alpha_{\text{rel}} \left(\frac{F_2}{F_2 + 0.25} \right)^2 ([\text{Ca}^{2+}]_{\text{rel}} - [\text{Ca}^{2+}]_i)$$

$$\frac{dO_{\text{Calse}}}{dt} = 480.0 [\text{Ca}^{2+}]_{\text{rel}} (1.0 - O_{\text{Calse}}) - 400.0 O_{\text{Calse}}$$

$$\frac{d[\text{Ca}^{2+}]_{\text{rel}}}{dt} = \frac{I_{\text{tr}} - I_{\text{rel}}}{2F \text{Vol}_{\text{rel}}} - 31.0 \frac{dO_{\text{Calse}}}{dt}$$

$$\frac{d[\text{Ca}^{2+}]_{\text{up}}}{dt} = \frac{I_{\text{up}} - I_{\text{tr}}}{2F \text{Vol}_{\text{up}}}$$

$$\frac{dF_1}{dt} = r_{\text{recov}} (1.0 - F_1 - F_2) - r_{\text{act}} F_1$$

$$\frac{dF_2}{dt} = r_{\text{act}} F_1 - r_{\text{inact}} F_2$$

$$r_{\text{act}} = 203.8 \left[\left(\frac{[\text{Ca}^{2+}]_i}{[\text{Ca}^{2+}]_i + k_{\text{rel},i}} \right)^4 + \left(\frac{[\text{Ca}^{2+}]_d}{[\text{Ca}^{2+}]_d + k_{\text{rel},d}} \right)^4 \right]$$

$$r_{\text{inact}} = 33.96 + 339.6 \left(\frac{[\text{Ca}^{2+}]_i}{[\text{Ca}^{2+}]_i + k_{\text{rel},i}} \right)^4$$

Table 12. Kinetics of CaTnC complexes

$$\frac{d[\text{CaTnC}]}{dt} = a_{\text{on}} \cdot (\text{TnC}_{\text{tot}} - [\text{CaTnC}]) \cdot [\text{Ca}^{2+}]_i - a_{\text{off}} \cdot e^{-k_A \cdot [\text{CaTnC}]} \cdot \Pi(N_A) \cdot [\text{CaTnC}]$$

$$\Pi(N_A) = \begin{cases} 1 & \text{if } N_A \leq 0 \\ \Pi_{\text{min}}^{N_A} & \text{if } 0 < N_A \leq 1 \\ \Pi_{\text{min}} & \text{otherwise} \end{cases}$$

$$N_A = \frac{\text{TnC}_{\text{tot}} \cdot N}{L_{\text{oz}} \cdot [\text{CaTnC}]}$$

Table 13. Reversal potentials: E_{Na} , E_{K} and E_{Ca}

$$E_{\text{Na}} = \frac{RT}{F} \log \frac{[\text{Na}^+]_c}{[\text{Na}^+]_i}$$

$$E_{\text{K}} = \frac{RT}{F} \log \frac{[\text{K}^+]_c}{[\text{K}^+]_i}$$

$$E_{\text{Ca}} = \frac{RT}{2F} \log \frac{[\text{Ca}^{2+}]_c}{[\text{Ca}^{2+}]_i}$$

2) Mechanical block

Table 14. Force

$$\begin{aligned}
 F_{CE} &= \lambda \cdot p_v \cdot N \\
 F_{SE} &= \beta_1 \cdot (e^{\alpha_1 \cdot (l_2 - l_1)} - 1) \\
 F_{PE} &= \beta_2 \cdot (e^{\alpha_2 \cdot l_2} - 1) \\
 F_{XSE} &= \beta_3 \cdot (e^{\alpha_3 \cdot l_3} - 1) \\
 F_{\text{sample}} &= F_{XSE}
 \end{aligned}$$

Table 15. Length

$$\begin{aligned}
 l &= l_2 + l_3 \\
 \frac{dl_1}{dt} &= v \\
 \frac{dl_2}{dt} &= w \\
 \frac{dl_3}{dt} &= \begin{cases} -w & \text{if isometry} \\ 0 & \text{if isotony} \end{cases}
 \end{aligned}$$

Table 16. Contractile element (CE) velocity

$$\begin{aligned}
 alp_p &= \begin{cases} \alpha_{vp1} & \text{if } v \leq 0 \\ \alpha_{vps} & \text{otherwise} \end{cases} \\
 k_{pvis} &= \begin{cases} \beta_{vp1} \cdot e^{\alpha_{vp1} \cdot l_1} & \text{if } v \leq 0 \\ \beta_{vps} \cdot e^{\alpha_{vps} \cdot l_1} & \text{otherwise} \end{cases} \\
 \Phi_x & \\
 = \begin{cases} \frac{\lambda \cdot K_k \cdot p_v + alp_p \cdot k_{pvis} \cdot v^2 + w \cdot (\alpha_2 \cdot \beta_2 \cdot e^{\alpha_2 \cdot l_2} + \alpha_3 \cdot \beta_3 \cdot e^{\alpha_3 \cdot l_3})}{\lambda \cdot N \cdot p_{\text{prime}_v} + k_{pvis}} & \text{if isometry} \\ \frac{\lambda \cdot K_k \cdot p_v + alp_p \cdot k_{pvis} \cdot v^2 + w \cdot \alpha_2 \cdot \beta_2 \cdot e^{\alpha_2 \cdot l_2}}{\lambda \cdot N \cdot p_{\text{prime}_v} + k_{pvis}} & \text{if iso} \end{cases} \\
 \frac{dv}{dt} &= \Phi_x
 \end{aligned}$$

Table 17. Parallel element (PE) velocity

$$\begin{aligned}
 alp_s &= \begin{cases} \alpha_{vs1} & \text{if } w \leq v \\ \alpha_{vss} & \text{otherwise} \end{cases} \\
 k_{svis} &= \begin{cases} \beta_{vs1} \cdot e^{\alpha_{vs1} \cdot (l_2 - l_1)} & \text{if } w \leq v \\ \beta_{vss} \cdot e^{\alpha_{vss} \cdot (l_2 - l_1)} & \text{otherwise} \end{cases}
 \end{aligned}$$

$$\frac{dw}{dt} = \begin{cases} \frac{\Phi_x - alp_s \cdot (w - v)^2 - \alpha_1 \cdot \beta_1 \cdot e^{\alpha_1 \cdot (l_2 - l_1)} \cdot (w - v) + w \cdot (\alpha_2 \cdot \beta_2 \cdot e^{\alpha_2 \cdot l_2} + \alpha_3 \cdot \beta_3 \cdot e^{\alpha_3 \cdot l_3})}{k_{svis}} & \text{if isometry} \\ \frac{k_{svis} \cdot (\Phi_x - alp_s \cdot (w - v)^2) - \alpha_1 \cdot \beta_1 \cdot e^{\alpha_1 \cdot (l_2 - l_1)} \cdot (w - v) - w \cdot \alpha_2 \cdot \beta_2 \cdot e^{\alpha_2 \cdot l_2}}{k_{svis}} & \text{if isotony} \end{cases}$$

Table 18. Average crossbridge force

$$v_1 = \frac{v_{max}}{10}$$

$$\gamma_2 = \frac{a \cdot d_h \cdot \left(\frac{v_1}{v_{max}}\right)^2}{3a \cdot d_h - \frac{(a+1) \cdot v_1}{v_{max}}}$$

$$P_{star}$$

$$= \begin{cases} \frac{a \cdot \left(1 + \frac{v}{v_{max}}\right)}{a - \frac{v}{v_{max}}} & \text{if } v \leq \\ 1 + d_h - \frac{d_h^2 \cdot a}{\frac{a \cdot d_h \cdot \left(\frac{v}{v_{max}}\right)^2 + \frac{(a+1) \cdot v}{v_{max}} + a \cdot d_h} \gamma_2} & \text{otherwise} \end{cases}$$

$$G_{star} = \begin{cases} 1 + \frac{0.6v}{v_{max}} & \text{if } v \leq 0 \\ \frac{P_{star}}{(0.4a+1) \cdot v} + 1 & \text{if } 0 < v \leq v_1 \\ \frac{P_{star} \cdot e^{-\alpha_G \cdot \left(\frac{v-v_1}{v_{max}}\right)^{\alpha_P}}}{(0.4a+1) \cdot v} + 1 & \text{otherwise} \end{cases}$$

$$\text{case}_1 = \frac{a \cdot (0.4 + 0.4a)}{v_{max} \cdot (0.4(a+1))^2}$$

$$\text{case}_2 = \frac{a \cdot \left(1 + 0.4a + \frac{1.2v}{v_{max}} + 0.6 \left(\frac{v}{v_{max}}\right)^2\right)}{v_{max} \cdot \left(\left(a - \frac{v}{v_{max}}\right) \cdot \left(1 + \frac{0.6v}{v_{max}}\right)\right)^2}$$

$$\text{case}_3 = \frac{0.4a + 1}{a \cdot v_{max}}$$

$$\text{case}_4 = \frac{1}{v_{max}} \cdot e^{-\alpha_G \cdot \left(\frac{v-v_1}{v_{max}}\right)^{\alpha_P}} \cdot \left(\frac{0.4a+1}{a} + \alpha_G \cdot \alpha_P \cdot \left(1 + \frac{(0.4a+1) \cdot v}{a \cdot v_{max}}\right) \cdot \left(\frac{v-v_1}{v_{max}}\right)^{\alpha_P-1}\right)$$

$$p_{\text{prime}_v} = \begin{cases} \text{case}_1 & \text{if } v \leq -v_{\max} \\ \text{case}_2 & \text{if } -v_{\max} < v \leq 0 \\ \text{case}_3 & \text{if } 0 < v \leq v_1 \\ \text{case}_4 & \text{otherwise} \end{cases}$$

$$p_v = \frac{P_{\text{star}}}{G_{\text{star}}}$$

Table 19. Crossbridge kinetics

$$M_A = \frac{\left(\frac{[\text{CaTnC}]}{\text{TnC}_{\text{tot}}}\right)^\mu \cdot (1 + k_\mu^\mu)}{\left(\frac{[\text{CaTnC}]}{\text{TnC}_{\text{tot}}}\right)^\mu + k_\mu^\mu}$$

$$\text{temp}_{n1} = g_1 \cdot l_1 + g_2$$

$$n_1 = \begin{cases} 0 & \text{if } \text{temp}_{n1} < 0 \\ \text{temp}_{n1} & \text{if } \text{temp}_{n1} < 1 \\ 1 & \text{otherwise} \end{cases}$$

$$L_{\text{oz}} = \begin{cases} \frac{l_1 + S_0}{0.46 + S_0} & \text{if } l_1 > 0.55 \\ S_0 + 0.55 & \text{otherwise} \end{cases}$$

$$v_{\text{st}} = x_{\text{st}} \cdot v_{\max}$$

$$q_v = \begin{cases} q_1 - \frac{q_2 \cdot v}{v_{\max}} & \text{if } v \leq 0 \\ \frac{(q_4 - q_3) \cdot v}{v_{\text{st}}} + q_3 & \text{if } 0 < v \leq v_{\text{st}} \\ \frac{q_4}{\left(1 + \frac{\beta_Q \cdot (v - v_{\text{st}})}{v_{\max}}\right)^{\alpha_Q}} & \text{otherwise} \end{cases}$$

$$k_{p_v} = \kappa \cdot \kappa_0 \cdot q_v \cdot m_0 \cdot G_{\text{star}}$$

$$k_{m_v} = \kappa_0 \cdot q_v \cdot (1 - \kappa \cdot m_0 \cdot G_{\text{star}})$$

$$K_\kappa = k_{p_v} \cdot M_A \cdot n_1 \cdot L_{\text{oz}} \cdot (1 - N) - k_{m_v} \cdot N$$

$$\frac{dN}{dt} = K_\kappa$$

Module 2: Optical activity in Fb

This module contains an electrophysiological model of atrial Fbs (represented by the Maleckar et al.) transduced with Chr2 currents (represented by Williams et al.) [3,4].

Table 20. Time- and voltage-dependent K^+ current: I_{Kv_Fb}

$$I_{Kv_Fb} = \bar{g}_{Kv,Fb} r_{Kv} s_{Kv} (V_{Fb} - E_{K,Fb})$$

$$\bar{r}_{Kv} = (1.0 + e^{-(V_{Fb}+20.0)/11.0})^{-1} \quad \bar{s}_{Kv} = (1.0 + e^{(V_{Fb}+23.0)/7.0})^{-1}$$

$$\frac{dr_{Kv}}{dt} = \frac{\bar{r}_{Kv} - r_{Kv}}{\tau_{r_{Kv}}} \quad \tau_{r_{Kv}} = 0.0203 + 0.138 \cdot e^{-((V_{Fb}+20.0)/25.9)^2}$$

$$\frac{ds_{Kv}}{dt} = \frac{\bar{s}_{Kv} - s_{Kv}}{\tau_{s_{Kv}}} \quad \tau_{s_{Kv}} = 1.574 + 5.268 \cdot e^{-((V_{Fb}+23.0)/22.7)^2}$$

$$E_{K,Fb} = \frac{RT}{F} \log \frac{[K^+]_{c,Fb}}{[K^+]_{i,Fb}}$$

Table 21. Time-independent inward-rectifying K^+ current: I_{K1_Fb}

$$I_{K1_Fb} = \bar{g}_{K1,Fb} \left(\frac{\alpha_{K1}}{\alpha_{K1} + \beta_{K1}} \right) (V_{Fb} - E_{K,Fb})$$

$$\alpha_{K1} = 0.1 (1.0 + e^{0.06(V_{Fb} - E_{K,Fb} - 200.0)})^{-1}$$

$$\beta_{K1} = \frac{3.0 \cdot e^{0.0002(V_{Fb} - E_{K,Fb} + 100.0)} + e^{0.1(V_{Fb} - E_{K,Fb} + 10.0)}}{1.0 + e^{-0.5(V_{Fb} - E_{K,Fb})}}$$

Table 22. Na^+ - K^+ pump current: I_{NaK_Fb}

$$I_{NaK_Fb} = \bar{I}_{NaK,Fb} \cdot \frac{[K^+]_{c,Fb}}{[K^+]_{c,Fb} + k_{mK,Fb}} \cdot \left(\frac{[Na^+]_{i,Fb}}{[Na^+]_{i,Fb} + k_{mNa,Fb}} \right)^{1.5} \cdot \frac{V_{Fb} + 150.0}{V_{Fb} + 200.0}$$

Table 23. Background inward current: I_{B,Na_Fb}

$$I_{B,Na_Fb} = \bar{g}_{B,Na,Fb} (V_{Fb} - E_{Na,Fb})$$

$$E_{Na,Fb} = \frac{RT}{F} \log \frac{[Na^+]_{c,Fb}}{[Na^+]_{i,Fb}}$$

Table 24. Intracellular ion concentrations: $[Na^+]_{i,Fb}$, and $[K^+]_{i,Fb}$

$$\frac{d[Na^+]_{i,Fb}}{dt} = - \frac{I_{B,Na_Fb} + 3I_{NaK_Fb}}{Vol_{i,Fb} F}$$

$$\frac{d[K^+]_{i,Fb}}{dt} = - \frac{I_{K1_Fb} + I_{Kv_Fb}}{Vol_{i,Fb} F}$$

Table 25. Model of I_{Chr2}

$$I_{Chr2} = \bar{g}_{Chr2} \cdot G(V) \cdot (O_1 + 0.1O_2) \cdot (V_{Fb} - E_{Chr2})$$

$$O_1 + O_2 + C_1 + C_2 = 1$$

$$\frac{dC_1}{dt} = G_r \cdot C_2 + G_{d1} \cdot O_1 + k_1 \cdot C_1$$

$$\begin{aligned} \frac{dO_1}{dt} &= k_1 \cdot C_1 - (G_{d1} + e_{12}) \cdot O_1 + e_{21} \cdot O_2 \\ \frac{dO_2}{dt} &= k_2 \cdot C_2 - (G_{d2} + e_{21}) \cdot O_2 + e_{12} \cdot O_1 \\ \frac{dC_2}{dt} &= G_{d2} \cdot O_2 - (k_2 + G_r) \cdot C_2 \\ e_{12} &= \left(0.011 + 0.005 \cdot \ln \left(1 + \frac{Irradiance}{0.024} \right) \right) \cdot 1000 \\ e_{21} &= \left(0.008 + 0.004 \cdot \ln \left(1 + \frac{Irradiance}{0.024} \right) \right) \cdot 1000 \\ k_1 &= \varepsilon_1 \cdot \sigma_{ret} \cdot Irradiance \cdot \frac{\lambda_1}{w_{loss} \cdot hc} \cdot p \\ k_2 &= \varepsilon_2 \cdot \sigma_{ret} \cdot Irradiance \cdot \frac{\lambda_1}{w_{loss} \cdot hc} \cdot p \\ \frac{dp}{dt} &= \frac{S0 - p}{\tau_{ChR2}} \\ S0 &= 0.5(1 + \tanh(120(100 \cdot Irradiance - 0.1))) \\ G(V) &= \frac{10.6408 - 14.6408e^{-\frac{V_{Fb}}{42.7671}}}{V_{Fb}} \\ G_{d1} &= \left(0.075 + 0.043 \tanh \left(-\frac{V_{Fb} + 20}{20} \right) \right) \cdot 1000 \\ G_r &= (4.34 \times 10^{-5} \times e^{-0.02115V_{Fb}}) \cdot 1000 \end{aligned}$$

Module 3: CM-Fb coupling

Table 26. Transmembrane potential of CM and Fb

$$\begin{aligned} \frac{dV_M}{dt} &= -\frac{1}{C_M} \left[I_M + \sum_{i=1}^n G_{gap}(V_M - V_{Fbi}) \right] \\ \frac{dV_{Fbi}}{dt} &= -\frac{1}{C_{Fb}} [I_{Fbi} + G_{gap}(V_{Fbi} - V_M)] \end{aligned}$$

$$I_M = I_{Na} + I_{CaL} + I_t + I_{Kur} + I_{K1} + I_{K,r} + I_{K,s} + I_{B,Na} + I_{B,Ca} + I_{NaK} + I_{CaP} + I_{NaCa}$$

$$I_{Fbi} = I_{Kv_Mfb} + I_{K1_Mfb} + I_{NaK_Mfb} + I_{B,Na_Mfb} + I_{ChR2}$$

Table 27. Parameter values

$[Na^+]_b = 130.0 \text{ mM}$	$k_{NaCa} = 0.0374842 \text{ pA}/(\text{mM})^4$
$[K^+]_b = 5.4 \text{ mM}$	$\gamma = 0.45$
$[Ca^{2+}]_b = 1.8 \text{ mM}$	$d_{NaCa} = 0.0003 \text{ (mM)}^{-4}$
$E_{Ca,app} = 60.0 \text{ mV}$	$\bar{I}_{up} = 2800.0 \text{ pA}$
$k_{Ca} = 0.025 \text{ mM}$	$k_{cyca} = 0.0003 \text{ mM}$
$R = 8314.0 \text{ mJ/MK}$	$K_{srca} = 0.5 \text{ mM}$

$T = 306.15 \text{ K}$	$K_{\text{xcs}} = 0.4$
$F = 96487.0 \text{ C/M}$	$\tau_{\text{tr}} = 0.01 \text{ s}$
$C_{\text{M}} = 60 \text{ pF}$	$\alpha_{\text{rel}} = 200000.0 \text{ pA (mM)}^{-1}$
$\text{Vol}_i = 0.005884 \text{ nL}$	$k_{\text{rel},i} = 0.0003 \text{ mM}$
$\text{Vol}_c = 0.136 \text{ Vol}_i$	$k_{\text{rel},d} = 0.003 \text{ mM}$
$k_{\text{CaP}} = 0.0002 \text{ mM}$	$r_{\text{recov}} = 0.815 \text{ s}^{-1}$
$\text{Vol}_d = 0.02 \text{ Vol}_i$	$q_1 = 17.3 \text{ s}^{-1}$
$\text{Vol}_{\text{rel}} = 0.0000441 \text{ nL}$	$q_2 = 259.0 \text{ s}^{-1}$
$\text{Vol}_{\text{up}} = 0.0003969 \text{ nL}$	$q_3 = 17.3 \text{ s}^{-1}$
$\tau_{\text{Na}} = 14.3 \text{ s}$	$q_4 = 15.0 \text{ s}^{-1}$
$\tau_{\text{K}} = 10.0 \text{ s}$	$x_{\text{st}} = 0.964285$
$\tau_{\text{Ca}} = 24.7 \text{ s}$	$a = 0.25$
$\tau_{\text{di}} = 0.01 \text{ s}$	$\alpha_1 = 14.6 (\mu\text{m})^{-1}$
$\bar{I}_{\text{NaK}} = 68.55 \text{ pA}$	$\alpha_2 = 14.6 (\mu\text{m})^{-1}$
$k_{\text{NaK,K}} = 1.0 \text{ mM}$	$\alpha_3 = 48.0 (\mu\text{m})^{-1}$
$k_{\text{NaK,Na}} = 11.0 \text{ mM}$	$\beta_1 = 0.84 \text{ mN}$
$\bar{I}_{\text{CaP}} = 4.0 \text{ pA}$	$\beta_2 = 0.0018 \text{ mN}$
$\alpha_{\text{vp}_1} = 16.0 (\mu\text{m})^{-1}$	$\beta_3 = 0.015 \text{ mN}$
$\alpha_{\text{vp}_s} = 16.0 (\mu\text{m})^{-1}$	$d_{\text{h}} = 0.5$
$\beta_{\text{vp}_1} = 0.0015 \text{ mN}\cdot\text{s}/\mu\text{m}$	$k_{\mu} = 0.6$
$\beta_{\text{vp}_s} = 0.0015 \text{ mN}\cdot\text{s}/\mu\text{m}$	$\kappa = 0.705$
$\alpha_{\text{vs}_1} = 39.0 (\mu\text{m})^{-1}$	$\kappa_0 = 3.0$
$\alpha_{\text{vs}_s} = 46.0 (\mu\text{m})^{-1}$	$\lambda = 30.0 \text{ mN}$
$\beta_{\text{vs}_1} = 0.008 \text{ mN}\cdot\text{s}/\mu\text{m}$	$m_0 = 0.9$
$\beta_{\text{vs}_s} = 0.006 \text{ mN}\cdot\text{s}/\mu\text{m}$	$\mu = 3.0$
$g_1 = 0.6 (\mu\text{m})^{-1}$	$v_{\text{max}} = 5.5 \mu\text{m/s}$
$g_2 = 0.52$	$C_{\text{Fb}} = 6.3 \text{ pF}$
$\text{TnC}_{\text{tot}} = 0.07 \text{ mM}$	$\text{Vol}_{i,\text{Fb}} = 0.00137 \text{ nL}$
$B_{1\text{tot}} = 0.08 \text{ mM}$	$[\text{Na}^+]_{\text{c,Fb}} = 130.011 \text{ mM}$
$B_{2\text{tot}} = 0.1 \text{ mM}$	$[\text{K}^+]_{\text{c,Fb}} = 5.3581 \text{ mM}$
$a_{\text{on}} = 70000.0 (\text{mM}\cdot\text{s})^{-1}$	$k_{\text{mK,Fb}} = 1.0 \text{ mmol}$
$a_{\text{off}} = 200.0 \text{ s}^{-1}$	$k_{\text{mNa,Fb}} = 11.0 \text{ mmol}$
$b_{1\text{on}} = 100000.0 (\text{mM}\cdot\text{s})^{-1}$	$\bar{I}_{\text{NaK,Fb}} = 10.36 \text{ pA}$
$b_{1\text{off}} = 182.0 \text{ s}^{-1}$	$E_{\text{ChR2}} = 0.0 \text{ mV}$
$b_{2\text{on}} = 1000.0 (\text{mM}\cdot\text{s})^{-1}$	$\varepsilon_1 = 0.8535$
$b_{2\text{off}} = 3.0 \text{ s}^{-1}$	$\varepsilon_1 = 0.14$
$\Pi_{\text{min}} = 0.03$	$\sigma_{\text{ret}} = 12.0 \times 10^{-20} \text{ m}^2$
$S_0 = 1.14 \mu\text{m}$	$\lambda_1 = 470.0 \text{ nm}$
$\alpha_{\text{G}} = 1.0$	$w_{\text{loss}} = 1.3$
$\alpha_{\text{P}} = 4.0$	$hc = 1.986446 \times 10^{-50} \text{ kg}\cdot\text{m}^3/\text{s}$
$\alpha_{\text{Q}} = 10.0$	$G_{\text{d2}} = 50.0 \text{ s}^{-1}$
$\beta_{\text{Q}} = 5.0$	$\tau_{\text{ChR2}} = 0.0013 \text{ s}$
$k_{\text{A}} = 40.0 (\text{mM})^{-1}$	

Table 28. Maximum conductance values

$P_{Na} = 0.0018 \text{ nL/s}$	$\bar{g}_{B,Na} = 0.060599 \text{ nS}$
$\bar{g}_{CaL} = 6.75 \text{ nS}$	$\bar{g}_{B,Ca} = 0.078681 \text{ nS}$
$\bar{g}_t = 8.25 \text{ nS}$	$\bar{g}_{Kv,Fb} = 1.575 \text{ nS}$
$\bar{g}_{Kur} = 2.25 \text{ nS}$	$\bar{g}_{K1,Fb} = 3.038 \text{ nS}$
$\bar{g}_{K,s} = 1.0 \text{ nS}$	$\bar{g}_{B,Na,Fb} = 0.05985 \text{ nS}$
$\bar{g}_{K,r} = 0.5 \text{ nS}$	$\bar{g}_{ChR2} = 0.4 \text{ nS/pF}$
$\bar{g}_{K1} = 3.1 \text{ nS}$	

Table 29. Initial conditions

$V_M = -74.03 \text{ mV}$	$F_1 = 0.4701$
$[Na^+]_c = 130.0221 \text{ mM}$	$F_2 = 0.0028$
$[K^+]_c = 5.5602 \text{ mM}$	$O_{Calse} = 0.4315$
$[Ca^{2+}]_c = 1.8158 \text{ mM}$	$[CaTnC] = 0.00015 \text{ mM}$
$[Na^+]_i = 8.5168 \text{ mM}$	$B_1 = 0.0 \text{ mM}$
$[K^+]_i = 129.486 \text{ mM}$	$B_2 = 0.0 \text{ mM}$
$[Ca^{2+}]_i = 6.5 \times 10^{-5} \text{ mM}$	$v = 0.0 \text{ } \mu\text{m/s}$
$[Ca^{2+}]_d = 7.1 \times 10^{-5} \text{ mM}$	$w = 0.0 \text{ } \mu\text{m/s}$
$[Ca^{2+}]_{up} = 0.6492 \text{ mM}$	$[K^+]_{i,Fb} = 129.4349 \text{ mM}$
$[Ca^{2+}]_{rel} = 0.6326 \text{ mM}$	$[Na^+]_{i,Fb} = 8.5547 \text{ mM}$
$m = 3.289 \times 10^{-3}$	$V_{Fb} = -47.75 \text{ mV}$
$h_1 = 0.8772$	$r_{Kv} = 0.0743$
$h_2 = 0.8739$	$s_{Kv} = 0.9717$
$d_L = 1.4 \times 10^{-5}$	$O_1 = 0.0$
$f_{L1} = 0.9986$	$O_2 = 0.0$
$f_{L2} = 0.9986$	$C_1 = 1.0$
$r = 1.089 \times 10^{-3}$	$C_2 = 0.0$
$s = 0.9486$	$p = 0.0$
$r_{Kur} = 3.67 \times 10^{-4}$	$N = 7.455 \times 10^{-8}$
$s_{Kur} = 0.9673$	$l_1 = 0.436 \text{ } \mu\text{m}$
$n = 4.374 \times 10^{-3}$	$l_2 = 0.436 \text{ } \mu\text{m}$
$p_a = 5.3 \times 10^{-5}$	$l_3 = 0.089 \text{ } \mu\text{m}$

Glossary

Module 1: ECC in CM			
I_{Na}	Na^+ current	I_{di}	Ca^{2+} diffusion current from the diffusion-restricted subsarcolemmal space to the cytosol
I_{CaL}	L-type Ca^{2+} current	I_{up}	Sarcoplasmic reticulum Ca^{2+} uptake current
I_t	Transient outward K^+ current	I_{CaP}	Sarcolemmal Ca^{2+} pump current
I_{Kur}	Sustained outward K^+ current	I_{NaCa}	Na^+ - Ca^{2+} exchange current
$I_{K,s}$	Slow delayed rectifier K^+ current	I_{tr}	Sarcoplasmic reticulum Ca^{2+} translocation current (from uptake to release compartment)

$I_{K,r}$	Rapid delayed rectifier K^+ current	I_{rel}	Sarcoplasmic reticulum Ca^{2+} release current
I_{K1}	Inwardly rectifying K^+ current	$[Na^+]_b$	Na^+ concentration in bulk (bathing) medium
$I_{B,Na}$	Background Na^+ current	$[K^+]_b$	K^+ concentration in bulk (bathing) medium
$I_{B,Ca}$	Background Ca^{2+} current	$[Ca^{2+}]_b$	Ca^{2+} concentration in bulk (bathing) medium
I_{NaK}	Na^+ - K^+ pump current	$[Na^+]_c$	Na^+ concentration in the extracellular cleft space
$[K^+]_c$	K^+ concentration in the extracellular cleft space	$\tau_{f_{L1}}, \tau_{f_{L2}}$	Fast and slow inactivation time constants for I_{CaL}
$[Ca^{2+}]_c$	Ca^{2+} concentration in the extracellular cleft space	τ_r	Activation time constant for I_t
$[Na^+]_i$	Na^+ concentration in the intracellular medium	τ_s	Inactivation time constant for I_t
$[K^+]_i$	K^+ concentration in the intracellular medium	$\tau_{r_{Kur}}$	Activation time constant for I_{Kur}
$[Ca^{2+}]_i$	Ca^{2+} concentration in the intracellular medium	$\tau_{s_{Kur}}$	Inactivation time constant for I_{Kur}
$[Ca^{2+}]_d$	Ca^{2+} concentration in the restricted subsarcolemmal space	τ_n	Activation time constant for $I_{K,s}$
$[Ca^{2+}]_{up}$	Ca^{2+} concentration in the sarcoplasmic reticulum uptake compartment	τ_{p_a}	Activation time constant for $I_{K,r}$
$[Ca^{2+}]_{rel}$	Ca^{2+} concentration in the sarcoplasmic reticulum release compartment	O_{calse}	Fractional occupancy of the calsequestrin buffer (in the sarcoplasmic reticulum release compartment) by Ca^{2+}
E_{Na}	Equilibrium (Nernst) potential for Na^+	R	Universal gas constant
E_K	Equilibrium (Nernst) potential for K^+	T	Absolute temperature
E_{Ca}	Equilibrium (Nernst) potential for Ca^{2+}	F	Faraday's constant
$E_{Ca,app}$	Apparent reversal potential for I_{CaL}	C_M	Membrane capacitance of CM
\bar{g}_{CaL}	Maximum conductance for I_{CaL}	V_M	Membrane voltage of CM
\bar{g}_t	Maximum conductance for I_t	Vol_c	Volume of the extracellular cleft space
\bar{g}_{Kur}	Maximum conductance for I_{Kur}	Vol_i	Total cytosolic volume
$\bar{g}_{K,s}$	Maximum conductance for $I_{K,s}$	Vol_d	Volume of the diffusion-restricted subsarcolemmal space
$\bar{g}_{K,r}$	Maximum conductance for $I_{K,r}$	Vol_{up}	Volume of the sarcoplasmic reticulum uptake compartment
\bar{g}_{K1}	Maximum conductance for I_{K1}	Vol_{rel}	Volume of the sarcoplasmic reticulum release compartment
$\bar{g}_{B,Na}$	Maximum conductance for $I_{B,Na}$	τ_{Na^+}	Time constant of diffusion of Na^+ , K^+ , and Ca^{2+} from the bulk medium to the extracellular cleft space
$\bar{g}_{B,Ca}$	Maximum conductance for $I_{B,Ca}$	$\tau_{Ca^{2+}}$	Time constant of diffusion from the restricted subsarcolemmal space to the cytosol
m	Activation gating variable for I_{Na}	\bar{I}_{NaK}	Maximum Na^+ - K^+ pump current
h_1, h_2	Fast and slow inactivation gating variables for I_{Na}	$k_{NaK,K}$	Half-maximum K^+ binding concentration for I_{NaK}
d_L	Activation gating variable for I_{CaL}	$k_{NaK,Na}$	Half-maximum Na^+ binding concentration for I_{NaK}
f_{L1}, f_{L2}	Fast and slow inactivation gating variables for I_{CaL}	\bar{I}_{CaP}	Half-maximum Ca^{2+} binding concentration for I_{CaP}
f_{Ca}	$[Ca^{2+}]_d$ -dependent ratio of fast (f_{L1}) to slow (f_{L2}) inactivation of I_{CaL}	k_{CaP}	Half-maximum Ca^{2+} binding concentration for I_{CaP}
k_{Ca}	Half-maximum Ca^{2+} binding concentration for f_{Ca}	k_{NaCa}	Scaling factor for I_{NaCa}

r	Activation gating variable for I_t	γ	Position of energy barrier controlling voltage dependence of I_{NaCa}
s	Inactivation gating variable for I_t	d_{NaCa}	Denominator constant for I_{NaCa}
r_{Kur}	Activation gating variable for I_{Kur}	\bar{I}_{up}	Maximum sarcoplasmic reticulum uptake current
s_{Kur}	Inactivation gating variable for I_{Kur}	k_{cyca}	Half-maximum binding concentration for $[Ca^{2+}]_i$ to I_{up}
p_i	Inactivation gating variable (instantaneous) for $I_{K,r}$	k_{srca}	Half-maximum binding concentration for $[Ca^{2+}]_{up}$ to I_{up}
$\bar{m}, \bar{h}_1, \dots$	Steady-state value of m, h_1 , etc	k_{xcs}	Ratio of forward to back reactions for I_{up}
F_1	Relative amount of “inactive precursor” in the I_{rel} formulation	τ_{tr}	Time constant of diffusion of Ca^{2+} from sarcoplasmic reticulum uptake to release compartment
F_2	Relative amount of “activator” in the I_{rel} formulation	α_{rel}	Scaling factor for I_{rel}
P_{Na}	Relative permeability to Na^+	$k_{rel,i}$	Half-activation $[Ca^{2+}]_i$ for I_{rel}
τ_m	Activation time constant for I_{Na}	$k_{rel,d}$	Half-activation $[Ca^{2+}]_d$ for I_{rel}
τ_{h_1}, τ_{h_2}	Fast and slow inactivation time constants for I_{Na}	r_{recov}	Recovery rate constant for the sarcoplasmic reticulum release channel
τ_{d_L}	Activation time constant for I_{CaL}	B_1	Concentration of Ca^{2+} bound with “fast” Ca^{2+} binding ligands
B_2	Concentration of Ca^{2+} bound with “slow” Ca^{2+} binding ligands	τ_{s_1}, τ_{s_2}	Rapidly and slowly recovering inactivation time constants for I_t
b_{1on}	Rate constant for Ca^{2+} bound with “fast” Ca^{2+} binding ligands	p_a	Activation gating variable for $I_{K,r}$
b_{2on}	Rate constant for Ca^{2+} bound with “slow” Ca^{2+} binding ligands	n	Activation gating variable for $I_{K,s}$
b_{1off}	Maximum rate constant for unbound with “fast” Ca^{2+} binding ligands	s_1, s_2	Rapidly and slowly recovering inactivation gating variables for I_t
b_{2off}	Maximum rate constant for unbound with “slow” Ca^{2+} binding ligands	alp_p	Parameter of Φ_x function
B_{1tot}	Total concentration of Ca^{2+} bound with “fast” Ca^{2+} binding ligands	$\alpha_{vp_1}, \alpha_{vp_s}, \beta_{vp_1}, \beta_{vp_s}$	Constants of the dependencies of k_{pvis} on the sarcomere length
B_{2tot}	Total concentration of Ca^{2+} bound with “slow” Ca^{2+} binding ligands	k_{pvis}	Length-dependent coefficient of viscosity for damper viscous element
$[CaTnC]$	Ca^{2+} -troponin complexes concentration	w	Velocity of PE deformation
TnC_{tot}	Total concentration of TnC	Φ_x	Derivative of velocity of CE deformation
α_{on}	Rate constant for CaTnC association	K_K	Derivative of cross-bridges concentration
α_{off}	Maximum rate constant for CaTnC dissociation	p_{prime_v}	Parameter of Φ_x function
k_A	Cooperativity parameter	alp_s	Parameter of w function
N_A	Average fraction of the attached cross-bridges per one CaTnC complex	$\alpha_{vs_1}, \alpha_{vs_s}, \beta_{vs_1}, \beta_{vs_s}$	Constants of the dependencies of k_{svis} on the sarcomere length
$\Pi(N_A)$	Dependence defining cooperativity of the contractile proteins	k_{svis}	Length-dependent coefficient of viscosity for damper viscous element
Π_{min}	Parameter of $\Pi(N_A)$ function	v_1	Parameter of p_v function
τ_{act}	Parameter of F_1 and F_2 function	v_{max}	Parameter of p_v function
τ_{inact}	Parameter of F_1 and F_2 function	γ_2	Parameter of P_{star} function
F_{CE}	Contractile element (sarcomere) (CE) force	a	Parameter of p_v function
F_{SE}	Serial elastic element (SE) force	d_h	Parameter of P_{star} function
F_{PE}	Parallel elastic element (PE) force	P_{star}	Dependence of the steady-state

F_{XSE}	Extra serial elastic element (XSE) force	G_{star}	sarcomere force on the sarcomere shortening/lengthening velocity Dependence of the steady-state sarcomere stiffness on the velocity
F_{sample}	Sample force	α_G	Parameter of G_{star} function
l	Deviation of the sample length from its slack length	α_P	Parameter of G_{star} function
l_1	Deviation of contractile length from its slack length	case ₁	Parameter of p_{prime_v} function
l_2	Deviation of parallel elastic element length from its slack length	case ₂	Parameter of p_{prime_v} function
l_3	Deviation of extra series element length from its slack length	case ₃	Parameter of p_{prime_v} function
α_1	Exponential coefficient of F_{SE}	case ₄	Parameter of p_{prime_v} function
α_2	Exponential coefficient of F_{PE}	M_A	End-to-end interaction between adjacent tropomyosin segments in the case if both of them affected by the respective CaTnC complexes formation
α_3	Exponential coefficient of F_{XSE}	k_u	Parameter of M_A function
β_1	Linear coefficient of F_{SE}	μ	Parameter of M_A function
β_2	Linear coefficient of F_{PE}	temp _{n1}	Parameter of n_1 function
β_3	Linear coefficient of F_{XSE}	g_1	Parameter of n_1 function
λ	Scale parameter of F_{CE}	g_2	Parameter of n_1 function
p_v	Steady-state dependence of an average crossbridge's force on the velocity of sarcomere shortening/lengthening	n_1	Probability of that a myosin head can find a vacant site on the actin filament
N	Cross-bridges concentration	v	Velocity of CE deformation
L_{oz}	Instantaneous length of thick and thin filament overlap zone	q_4	Parameter of q_v function
S_0	Parameter of L_{oz} function	α_Q	Parameter of q_v function
v_{st}	Parameter of q_v function	β_Q	Parameter of q_v function
x_{st}	Parameter of q_v function	k_{p_v}	Parameter of K_κ function
q_v	Stationary relation "stiffness-velocity" for the sample	κ	Function required for variation the ratio between rates of cross-bridge attachment and detachment
q_1	Parameter of q_v function	κ_0	Parameter of κ function
q_2	Parameter of q_v function	m_0	Fraction of strongly attached Xb in steady state isometric conditions
q_3	Parameter of q_v function	k_{m_v}	Parameter of K_κ function

Module 2: Optical activity in Fb

$I_{K_v_Fb}$	Time- and voltage-dependent K^+ current in Fb	E_{ChR2}	Reversal potential for ChR2
I_{K1_Fb}	Inward-rectifying K^+ current in Fb	O_1	Open state probability
I_{NaK_Fb}	Na^+ - K^+ pump current in Fb	O_2	Open state probability
I_{B,Na_Fb}	Background Na^+ current in Fb	C_1	Closed state probability
$[Na^+]_{c,Fb}$	Na^+ concentration in the Fb's extracellular cleft space	C_2	Closed state probability
$[K^+]_{c,Fb}$	K^+ concentration in the Fb's extracellular cleft space	G_r	Rate constant for $C_2 \rightarrow C_1$ transition
$[Na^+]_{i,Fb}$	Na^+ concentration in the Fb's intracellular medium	G_{d1}	Rate constant for $O_1 \rightarrow C_1$ transition
$[K^+]_{i,Fb}$	K^+ concentration in the Fb's intracellular medium	k_1	Light-sensitive rate constant for $C_1 \rightarrow O_1$ transition
$E_{Na,Fb}$	Equilibrium (Nernst) potential for Na^+ in Fb	e_{12}	Rate constant for $O_1 \rightarrow O_2$ transition
$E_{K,Fb}$	Equilibrium (Nernst) potential for K^+	e_{21}	Rate constant for $O_2 \rightarrow O_1$ transition

	in Fb		
$\bar{g}_{Kv,Fb}$	Maximum conductance for I_{Kv_Fb}	k_2	Light-sensitive rate constant for $C_2 \rightarrow O_2$ transition
$\bar{g}_{K1,Fb}$	Maximum conductance for I_{K1_Fb}	G_{d2}	Rate constant for $O_2 \rightarrow C_2$ transition
$\bar{g}_{B,Na,Fb}$	Maximum conductance for I_{b,Na_Fb}	<i>Irradiance</i>	Light irradiance
$Vol_{i,Fb}$	Total cytosolic volume of Fb	ϵ_1	Quantum efficiency for photon absorption from C_1
r_{kv}	Activation gating variable for I_{Kv_Fb}	ϵ_2	Quantum efficiency for photon absorption from C_2
s_{kv}	Inactivation gating variable for I_{Kv_Fb}	σ_{ret}	Absorption cross-section for retinal
$\tau_{r_{Kv}}$	Activation time constant for I_{Kv_Fb}	λ_1	Wavelength of max absorption for retinal
$\tau_{s_{Kv}}$	Inactivation time constant for I_{Kv_Fb}	w_{loss}	Scaling factor for losses of photons due to scattering or absorption
α_{K1}, β_{K1}	Fractional open probability of the I_{K1_Fb} channel	hc	Product of Planck's constant and the speed of light
$\bar{I}_{NaK,Fb}$	Maximum $Na^+ - K^+$ pump current in Fb	p	State-variable, time- and irradiance-dependent activation function for ChR2
$k_{mK,Fb}$	Half-maximum K^+ binding concentration for I_{NaK_Fb}	$S0$	Parameter of p function
$k_{mNa,Fb}$	Half-maximum Na^+ binding concentration for I_{NaK_Fb}	τ_{ChR2}	Time constant of ChR2 activation
$\bar{r}_{Kv}, \bar{s}_{Kv}, \dots$	Steady-state value of $r_{Kv}, s_{Kv},$ etc	I_{ChR2}	ChR2 current
C_{Fb}	Membrane capacitance of Fb	\bar{g}_{ChR2}	Max conductance for I_{ChR2}
V_{Fb}	Membrane voltage of Fb	$G(V)$	Voltage-dependent rectification function
Module 3: CM-Fb coupling			
G_{gap}	Gap junctional conductance between Fb and CM	I_M	Net membrane current of the CM
n	Number of Fbs coupled to each CM	I_{Fb}	Net membrane current of the Fb

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